Novel Cheminformatics Methods for Modeling Biomolecular Data in High Dimension Low Sample Size (HDLSS) Chemistry Space

Tong-Ying Wu

A dissertation submitted to the faculty of the University of North Carolina at Chapel Hill in partial fulfillment of the requirements for the degree of Doctor of Philosophy in the Department of Biomedical Engineering.

Chapel Hill
2011

Approved By:
Alexander Tropsha, Ph.D.
David Lalush, Ph.D.
J. S. Marron, Ph.D.
Jack Snoeyink, Ph.D.
Shawn Gomez, Ph.D.
ABSTRACT

TONG-YING WU: Novel Cheminformatics Methods for Modeling Biomolecular Data in High Dimension Low Sample Size (HDLSS) Chemistry Space
(Under the direction of Dr. Alexander Tropsha)

The increasing availability of biological and chemical data has led to a critical need for cheminformatics and bioinformatics tools to analyze the data. However, not all datasets contain sufficient information for significant analysis. One problem is High Dimension Low Sample Size (HDLSS), where the number of structural characteristics, e.g., molecular descriptors, that can be calculated from a single compound (high dimensionality) far exceeds the number of compounds (low sample size). A major challenge associated with modeling HDLSS data is overfitting, and specialized tools are required to overcome the statistical difficulties inherent to HDLSS. We improved the Distance Weighted Discrimination (DWD) statistical learning method through a new variable selection technique for estimating the intrinsic dimension of a dataset, i.e., the minimum number of descriptors to classify data. Compared to SVM and DWD without variable selection, DWD with variable selection achieved higher prediction accuracy on several benchmarking datasets and allowed the identification of key molecular features that contribute to investigated biological properties, e.g., inhibition of AmpC β-lactamase and binding affinity for the various serotonin receptors.

For analyzing and modeling stereochemistry-dependent datasets, we developed chiral atom-pair descriptors (3D chiral atom-pair), which were calculated from three-dimensional molecular structures. QSAR models built with these descriptors, versus either 3D non-chiral atom-pair or 2D Dragon descriptors, more accurately predicted antimalarial activity and binding
affinities of small molecules toward various receptors. Our method not only led to the
identification of a subset of chiral atoms that are expected to affect the selected biological
property, e.g., antimalarial activity, but also enabled the development of models that would not be
possible otherwise.

To aid automatic protein function annotation, especially in the case of functional
homologs, we developed new protein descriptors based solely on protein's structure. Our method
showed sensitivity comparable to that of ScanPROSITE. When predicted annotations from both
ScanPROSITE and our method were combined into a consensus model, we observed a significant
gain in prediction reliability and the successful functional annotation of proteins with low
sequence similarity.
Dedication

To my family for their continuous support in overcoming the difficulties in life. To my grandmother, who passed away prior to my pursuit of a Ph.D. degree, for always believing in me. My love for her and sadness at her passing have fueled my determination to be an active member in the medical and pharmaceutical research rather than just bring the life saving technologies to people.
Acknowledgments

Numerous people contributed to the research projects described in this dissertation, and I am grateful to all of them for their discussion and support. I would like to acknowledge my research chair, Dr. Alexander Tropsha, for his supervision, guidance, and funding that made this research possible. This research could not have been completed without Dr. J.S. Marron’s expertise in Object Oriented Data Analysis, especially for HDLSS data. Dr. Jack Snoeyink’s expertise in computational geometry and Dr. Shawn Gomez’s suggestions on protein function annotation facilitated the study of the protein structure-function relationship. In order to study the contribution of stereochemistry in biological activities, Dr. Weifan Zheng’s guidance in chirality descriptors was critical. I also like to express my gratitude to my committee chair, Dr. David Lalush, and the Director in Graduate Studies in BME, Dr. Paul Dayton, for their effort to resolve the challenges that I faced during my study. I also would like to thank Dr. Denis Fourches, Dr. Eugene Muratov, Dr. Alexander Sedykh, Dr. Alexander Golbraikh and Mr. Stephen J. Bush for their feedback and criticisms for improving this manuscript. Discussions with Dr. Wei Wang, Dr. Leonard McMillan, Dr. Jan Prins, Dr. Brian Kuhlman, Dr. Tim Elston, and Dr. Yufeng Liu also made impacts on this research.

Additionally, I owe my deepest gratitude to the following people for their guidance in computer-aided drug design: Dr. Markus Boehm and Dr. Gregory A. Bakken from Pfizer for their knowledge in lead identification; Dr. Christian Lemmen, Dr. Holger Claussen, and Dr. Markus Lilienthal from Bio SolveIT for their help to implement an efficient similarity search method in large databases; Dr. Chris Bizon from Renaissance Computing Institute and Dr. Dana E. Vanderwall from Bristol-Myers Squibb for their knowledge in multi-property lead optimization;
Dr. Martha S. Head, Dr. Kaushik Raha, Dr. Deepak Bandyopadhyay from GlaxoSmithKline (GSK) for their knowledge in the structure-based drug design and the identification of the protein binding sites; Dr. Daniel J. Price and Dr. Eugene L. Stewart from GSK and Dr. Zheng P. Yang from Boehringer Ingelheim for their knowledge in the overall computer-aided drug design.
# Table of Contents

List of Tables .............................................................................................................. xi
List of Figures .............................................................................................................. xii
List of Abbreviations ................................................................................................... xiv
Chapter 1 .................................................................................................................... 1
  1.1 Background Information ................................................................................... 1
    1.1.1 Growth of Publically Available Data ......................................................... 1
    1.1.2 Overview of Computational Methods Employed in Drug Discovery .......... 2
    1.1.3 Applications of Classification Methods in Cheminformatics and Bioinformatics .... 3
  1.2 Research Motivation ......................................................................................... 5
Chapter 2 .................................................................................................................... 7
  Variable Selection Based Classification Method for Imbalanced and HDLSS Data .......... 7
    2.1 Motivation ........................................................................................................ 7
    2.2 Overview of Variable Selection ..................................................................... 9
    2.3 Overview of the Proposed Method ................................................................ 10
      2.3.1 Variable Evaluation Component .............................................................. 11
      2.3.2 Variable Selection (Ranking) Component ................................................ 12
    2.5 Simulation ...................................................................................................... 13
      2.5.1 Experimental Design .............................................................................. 13
      2.5.2 Simulation Result .................................................................................... 14
    2.6 QSAR Studies ................................................................................................. 18
    2.7 QSAR Datasets Description ........................................................................... 19
      2.7.1 AmpC β-lactamases Dataset Description (110 Compounds) .................... 19
      2.7.2 5-HT Datasets Description ................................................................... 20
    2.8 QSAR Modeling Results ................................................................................ 21
      2.8.1 AmpC B-lactamase Modeling Result (110 Compounds) ............................ 21
<table>
<thead>
<tr>
<th>Section</th>
<th>Title</th>
<th>Page</th>
</tr>
</thead>
<tbody>
<tr>
<td>2.8.2</td>
<td>5-HT_{1B} Modeling Result</td>
<td>24</td>
</tr>
<tr>
<td>2.8.3</td>
<td>5-HT_{1D} Modeling Result</td>
<td>28</td>
</tr>
<tr>
<td>2.8.4</td>
<td>5-HT_{2B} Modeling Result</td>
<td>32</td>
</tr>
<tr>
<td>2.8.5</td>
<td>5-HT_{6} Modeling Result</td>
<td>34</td>
</tr>
<tr>
<td>2.9</td>
<td>Conclusion</td>
<td>36</td>
</tr>
<tr>
<td>Chapter 3</td>
<td>Novel Three-Dimensional Chirality Atom-Pair Descriptors</td>
<td>37</td>
</tr>
<tr>
<td>3.1</td>
<td>Motivation / Background</td>
<td>37</td>
</tr>
<tr>
<td>3.2</td>
<td>Overview of Current Chirality Descriptors</td>
<td>40</td>
</tr>
<tr>
<td>3.2.1</td>
<td>Prior Implementation of the Chirality Descriptors</td>
<td>41</td>
</tr>
<tr>
<td>3.2.2</td>
<td>Three Dimensional Chiral Atom-Pair Descriptors</td>
<td>42</td>
</tr>
<tr>
<td>3.3</td>
<td>Dataset Description</td>
<td>45</td>
</tr>
<tr>
<td>3.3.1</td>
<td>Peptide Transporter 1 (PEPT1) Dataset</td>
<td>45</td>
</tr>
<tr>
<td>3.3.2</td>
<td>AmpC β–lactamases Dataset (149 Compounds)</td>
<td>47</td>
</tr>
<tr>
<td>3.3.3</td>
<td>Artemisinin Dataset</td>
<td>49</td>
</tr>
<tr>
<td>3.4</td>
<td>Evaluation of Descriptor Significance and Generalizability</td>
<td>50</td>
</tr>
<tr>
<td>3.5</td>
<td>Modeling Results</td>
<td>51</td>
</tr>
<tr>
<td>3.5.1</td>
<td>PEPT1 Dataset</td>
<td>51</td>
</tr>
<tr>
<td>3.5.2</td>
<td>AmpC β–lactamase Dataset (149 Compounds)</td>
<td>54</td>
</tr>
<tr>
<td>3.5.3</td>
<td>Artemisinin Dataset</td>
<td>56</td>
</tr>
<tr>
<td>3.6</td>
<td>Conclusion</td>
<td>59</td>
</tr>
<tr>
<td>Chapter 4</td>
<td>Novel Protein Descriptors</td>
<td>61</td>
</tr>
<tr>
<td>4.1</td>
<td>Growth of Protein Structure Data</td>
<td>61</td>
</tr>
<tr>
<td>4.2</td>
<td>Overview of Current Methods for Protein Function Annotation</td>
<td>64</td>
</tr>
<tr>
<td>4.2.1</td>
<td>Sequence-Based Methods</td>
<td>64</td>
</tr>
<tr>
<td>4.2.2</td>
<td>Structure-Based Methods</td>
<td>65</td>
</tr>
<tr>
<td>4.3</td>
<td>Motivation</td>
<td>68</td>
</tr>
<tr>
<td>4.4</td>
<td>Novel Protein Descriptors</td>
<td>68</td>
</tr>
<tr>
<td>4.4.1</td>
<td>Classification Scheme of Amino Acids (Ten Classes) and Geometric Properties</td>
<td>69</td>
</tr>
<tr>
<td>4.4.2</td>
<td>Geometric Property of α–helix vs. β sheet</td>
<td>72</td>
</tr>
<tr>
<td>4.5</td>
<td>Selection of Enzyme Diversity Set</td>
<td>75</td>
</tr>
<tr>
<td>Section</td>
<td>Title</td>
<td>Page</td>
</tr>
<tr>
<td>---------</td>
<td>-------</td>
<td>------</td>
</tr>
<tr>
<td>4.6</td>
<td>Selection of Enzyme Homologs</td>
<td>76</td>
</tr>
<tr>
<td>4.6.1</td>
<td>Superoxide Dismutase (EC 1.15.1.1)</td>
<td>77</td>
</tr>
<tr>
<td>4.6.2</td>
<td>Cellulase (EC 3.2.1.4)</td>
<td>78</td>
</tr>
<tr>
<td>4.6.3</td>
<td>β-lactamase (EC 3.5.2.6)</td>
<td>79</td>
</tr>
<tr>
<td>4.6.4</td>
<td>Methyltransferases (EC 2.1.1)</td>
<td>80</td>
</tr>
<tr>
<td>4.6.5</td>
<td>Protein-Tyrosine Kinases (EC 2.7.10)</td>
<td>81</td>
</tr>
<tr>
<td>4.6.6</td>
<td>Protein-Serine/Threonine Kinases (EC 2.7.11)</td>
<td>82</td>
</tr>
<tr>
<td>4.7</td>
<td>Modeling Results</td>
<td>84</td>
</tr>
<tr>
<td>4.7.1</td>
<td>Superoxide Dismutase (EC 1.15.1.1)</td>
<td>84</td>
</tr>
<tr>
<td>4.7.2</td>
<td>Cellulase (EC 3.2.1.4)</td>
<td>89</td>
</tr>
<tr>
<td>4.7.3</td>
<td>β-lactamase (EC 3.5.2.6)</td>
<td>92</td>
</tr>
<tr>
<td>4.7.4</td>
<td>Methyltransferases (EC 2.1.1)</td>
<td>96</td>
</tr>
<tr>
<td>4.7.5</td>
<td>Protein-Tyrosine Kinases (EC 2.7.10)</td>
<td>97</td>
</tr>
<tr>
<td>4.7.6</td>
<td>Protein-Serine/Threonine Kinase (EC 2.7.11)</td>
<td>98</td>
</tr>
<tr>
<td>4.8</td>
<td>Evaluating the Stability of the Delaunay Simplices</td>
<td>99</td>
</tr>
<tr>
<td>4.9</td>
<td>Comparison with PROSITE</td>
<td>102</td>
</tr>
<tr>
<td>4.9.1</td>
<td>Test Set Selection Criteria for Both E.C. 1.15.1.1 and E.C. 3.5.2.6</td>
<td>102</td>
</tr>
<tr>
<td>4.9.2</td>
<td>Comparison Results</td>
<td>103</td>
</tr>
<tr>
<td>4.10</td>
<td>Conclusion</td>
<td>105</td>
</tr>
<tr>
<td>Chapter 5</td>
<td>Summary</td>
<td>108</td>
</tr>
</tbody>
</table>
List of Tables

Table 2. 1. 5-HT receptor families and subtypes. .................................................................20
Table 2. 2. The 49 unique descriptors selected by all 5 models. ........................................22
Table 2. 3. The unique descriptors selected by all 5 models from each of the three clusters. ......27
Table 2. 4. The unique descriptors selected by all 5 models from each of the three clusters. ......31
Table 2. 5. The 79 unique descriptors selected by all 5 models for the 5-HT_{2B} dataset ...........33
Table 2. 6. The 87 unique descriptors selected by all 5 models for the 5-HT_{6} dataset ............35

Table 3. 1. List of top ten single enantiomer blocker drugs in ranking order .........................39
Table 3. 2. The 17 atom types defined in the atom pair descriptors .......................................41

Table 4. 1. Possible categories of tetrahedra ........................................................................70

**Table 4. 2.** Classification of amino acids ........................................................................71
Table 4. 3. Fourteen PDB entries retrieved based on the selection criteria from EC 1.15.1.1......77
Table 4. 4. Eighteen PDB entries retrieved based on the selection criteria for EC 3.2.1.4 ........78
Table 4. 5. Twelve PDB entries retrieved based on the selection criteria for EC 3.5.2.6 ...........79
Table 4. 6. Thirty-two PDB entries retrieved based on the selection criteria for EC 2.1.1 ..........80
Table 4. 7. Eleven PDB entries retrieved based on the selection criteria for EC 2.7.10 ..........81
Table 4. 8. List of subfamilies associated with EC 2.7.11 .....................................................82
Table 4. 9. Twenty-two PDB entries retrieved based on the selection criteria for EC 2.7.11 ....83
Table 4. 10. The list of PDB entries selected for the two positive classes ...............................103
List of Figures

Figure 2. 1. Informative descriptor vs. noise. .................................................................13
Figure 2. 2. Prediction results of the 30 simulation runs. ..............................................15
Figure 2. 3. Identification of the optimal model through the z-score ...............................16
Figure 2. 4. Model analysis of the simulated data. ..........................................................17
Figure 2. 5. Performance comparison between the methods (AmpC dataset) .......................21
Figure 2. 6. Common structure features of the AmpC inhibitors .....................................23
Figure 2. 7. Dendrogram of the 5-HT_1B dataset .............................................................24
Figure 2. 8. Data distribution of the 5-HT_1B dataset in Dragon chemical descriptor space .......25
Figure 2. 9. Performance comparison through five-fold external cross validation for the 5-HT_1B dataset ........................................................................26
Figure 2. 10. Dendrogram of the 5-HT_1D dataset ............................................................28
Figure 2. 11. Data distribution of the 5-HT_1D dataset in Dragon chemical descriptor space ....29
Figure 2. 12. Performance comparison through five-fold external cross validation for the 5-HT_1D dataset ........................................................................30
Figure 2. 13. Performance comparison through five-fold external cross validation for the 5-HT_2B dataset ........................................................................32
Figure 2. 14. Performance comparison through five-fold external cross validation for the 5-HT_6 dataset ........................................................................34

Figure 3. 1. Two enantiomers of a generic amino acid. .........................................................37
Figure 3. 2. Growing trend of chiral technology .................................................................38
Figure 3. 3. Activity distribution of the PEPT1 binding substrates ....................................46
Figure 3. 4. Isoindole-like scaffold (top) vs. isoindole (bottom) .........................................47
Figure 3. 5. The two enantiomeric pairs observed in the additional AmpC dataset ..............48
Figure 3. 6. Structure of artemisinin ..................................................................................49
Figure 3. 7. Degrees of chirality and the corresponding significance for the PEPT1 dataset ....51
Figure 3. 8. Five-fold external cross-validation results of the PEPT1 dataset .....................53
Figure 3. 9. Degrees of chirality and the corresponding significance for AmpC β-lactamase dataset (149 compounds) .................................................................54
Figure 3. 10. Five-fold external cross-validation result of AmpC β-lactamase dataset (149 compounds) ........................................................................55
Figure 3. 11. Degrees of chirality and corresponding significance for the artemisinin dataset ...57
Figure 3. 12. Five-fold external cross-validation result of the artemisinin dataset ...............58

Figure 4. 1. Annual growth of structures in Protein Data Bank (PDB) ...............................62
Figure 4. 2. Annual growth of protein structures with unknown function .............................63
Figure 4. 3. Illustration of the potential benefit by encoding the geometric property of tetrahedra for proteins .................................................................72
Figure 4. 4. α-helix vs. β-sheet .....................................................................................73
Figure 4. 5. Difference in geometric properties of tetrahedra between α-helix and β sheet ....74
Figure 4. 6. Distribution of the six major enzyme groups in the enzyme diversity set. 
Figure 4. 7. Identifying the most significant model through z-scores for the EC1.15.1.1 dataset.
Figure 4. 8. The five most significant features associated with SOD.
Figure 4. 9. Mapping the four identified tetrahedral categories onto structures associated with SOD binding to Fe$^{3+}$ and Mn$^{2+}$.
Figure 4. 10. Mapping the four identified tetrahedral categories onto structures associated with SOD binding to Cu$^{2+}$/Zn$^{2+}$ and Ni$^{2+}$.
Figure 4. 11. Mapping the four identified tetrahedral categories onto PDB entry, 1TO4.
Figure 4. 12. Identifying the most significant model through z-score for the EC 3.2.1.4 dataset.
Figure 4. 13. The twenty most significant features associated with cellulase.
Figure 4. 14. Mapping the 18 identified tetrahedral categories onto the structures associated with cellulases.
Figure 4. 15. Identifying the most significant model through z-score for the EC 3.5.2.6 dataset.
Figure 4. 16. The ten most significant features associated with β-lactamase.
Figure 4. 17. Mapping the six identified tetrahedral categories onto the 1M2X and 1NYM structure entries (EC 3.5.2.6 family).
Figure 4. 18. Mapping the six identified tetrahedral categories onto the 3G35 structure entry (EC 3.5.2.6 family).
Figure 4. 19. Identifying the most significant model through z-score for the EC 2.1.1 dataset.
Figure 4. 20. Identifying the most significant model through z-score for the EC 2.7.10 dataset.
Figure 4. 21. Identifying the most significant model through z-scores for the EC 2.7.11 dataset.
Figure 4. 22. Mapping the four categories of tetrahedron found in the modified structures onto the original 1TO4 PDB structure.
Figure 4. 23. Benchmarking results.
# List of Abbreviations

<p>| 2D | Two-Dimensional |
| 3D | Three-Dimensional |
| 5-HT | 5-hydroxytryptamine |
| Ala | Alanine |
| APair | Regular Atom-pair (without chiral atom types) |
| Arg | Arginine |
| Asn | Asparagine |
| Asp | Aspartic Acid |
| BCC | Bond-charge Corrections |
| BLAST | Basic Local Alignment Search Tool |
| Ca$^{2+}$ | Calcium (II) Cation |
| CADD | Computer-aided Drug Design |
| CAMD | Computer-aided Molecular Design |
| cAMP | Cyclic Adenosine Monophosphate |
| cAP | Chiral Atom-pair |
| CCR | Correct Classification Rate |
| CDD | Collaborative Drug Discovery |
| cGMP | Cyclic Guanosine Monophosphate |
| Cu | Cupper |
| Cu$^{2+}$ | Copper (II) Cation; Cupric Cation |
| Cys | Cysteine |
| DNA | Deoxyribonucleic Acid |
| DWD | Distance Weighted Discrimination; weighted Distance Weighted Discrimination |
| DWD (No V.S.) | wDWD without Variable Selection |
| DWD (V.S.) | wDWD with Variable Selection |
| DWD_VS | wDWD with Variable Selection |
| E.C. | Enzyme Classification or Enzyme Commission |
| ET | Evolutionary Trace |
| FDA | US Food and Drug Administration |
| Fe | Iron |
| Fe$^{3+}$ | Iron (III) Cation; Ferric Cation |
| Gln | Glutamine |
| Glu | Glutamate |
| Gly | Glycine |
| GPSS | Global Protein Surface Survey |
| HDLSS | High Dimension Low Sample Size |
| His | Histidine |
| HMM | Hidden Markov Model |
| Ile | Isoleucine |
| Leu | Leucine |</p>
<table>
<thead>
<tr>
<th>Symbol</th>
<th>Term</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lys</td>
<td>Lysine</td>
</tr>
<tr>
<td>Met</td>
<td>Methionine</td>
</tr>
<tr>
<td>Mn</td>
<td>Manganese</td>
</tr>
<tr>
<td>Mn&lt;sup&gt;2+&lt;/sup&gt;</td>
<td>Manganese Cation</td>
</tr>
<tr>
<td>MSA</td>
<td>Multiple Sequence Alignment</td>
</tr>
<tr>
<td>Ni</td>
<td>Nickel</td>
</tr>
<tr>
<td>Ni&lt;sup&gt;2+&lt;/sup&gt;</td>
<td>Nickel(II) Cation; Nickelous Cation</td>
</tr>
<tr>
<td>NMR</td>
<td>Nuclear Magnetic Resonance</td>
</tr>
<tr>
<td>PCA</td>
<td>Principal Component Analysis</td>
</tr>
<tr>
<td>PDB</td>
<td>Protein Data Bank</td>
</tr>
<tr>
<td>PDSP</td>
<td>Psychoactive Drug Screening Program</td>
</tr>
<tr>
<td>PEPT1</td>
<td>Peptide Transporter 1</td>
</tr>
<tr>
<td>Phe</td>
<td>Phenylalanine</td>
</tr>
<tr>
<td>PINTS</td>
<td>Patterns in Non-homologous Tertiary Structures</td>
</tr>
<tr>
<td>Pro</td>
<td>Proline</td>
</tr>
<tr>
<td>PSI-BLAST</td>
<td>Position-Specific Iterative BLAST</td>
</tr>
<tr>
<td>pvSOAR</td>
<td>Pocket and Void Surfaces of Amino Acid Residues</td>
</tr>
<tr>
<td>QSAR</td>
<td>Quantitative Structure-Activity Relationship</td>
</tr>
<tr>
<td>RA</td>
<td>Relative Activity</td>
</tr>
<tr>
<td>RMS</td>
<td>Root Mean Square</td>
</tr>
<tr>
<td>RMSD</td>
<td>Root-mean-square Deviation</td>
</tr>
<tr>
<td>RNA</td>
<td>Ribonucleic Acid</td>
</tr>
<tr>
<td>SCOP</td>
<td>Structural Classification of Proteins</td>
</tr>
<tr>
<td>Ser</td>
<td>Serine</td>
</tr>
<tr>
<td>SOD</td>
<td>Superoxide Dismutase</td>
</tr>
<tr>
<td>SVM</td>
<td>Support Vector Machine</td>
</tr>
<tr>
<td>Thr</td>
<td>Threonine</td>
</tr>
<tr>
<td>Trp</td>
<td>Tryptophan</td>
</tr>
<tr>
<td>Tyr</td>
<td>Tyrosine</td>
</tr>
<tr>
<td>V.S.</td>
<td>Variable Selection</td>
</tr>
<tr>
<td>Val</td>
<td>Valine</td>
</tr>
<tr>
<td>wDWD</td>
<td>weighted Distance Weighted Discrimination</td>
</tr>
<tr>
<td>Zn</td>
<td>Zinc</td>
</tr>
<tr>
<td>Zn&lt;sup&gt;2+&lt;/sup&gt;</td>
<td>Zinc Cation</td>
</tr>
</tbody>
</table>
Chapter 1
Introduction

1.1 Background Information

1.1.1 Growth of Publically Available Data

Due to technological advancements within the past two decades, rapid synthesis and high-throughput screening of large chemical libraries have become routine procedures in the pharmaceutical industry, which has resulted in a massive increase of data for chemical compounds, and their targets, pathways, and associated data. These data were largely proprietary and therefore rarely available to the academic research community. However, within the past decade, high-throughput screening has become increasingly common in academia, and the resulting data are typically published and often made publically available in relevant databases. Additionally, some pharmaceutical companies are also beginning to put some of their datasets in the public domain.

Several publically available databases have been created. PubChem \(^{97}\) is a public database that launched as a result of the cheminformatics initiatives from the National Institutes of Health (NIH) in October, 2004. It contains data regarding chemical molecules whose activities have been experimentally measured using biological assays. As of mid January, 2011, PubChem provides open access to data from over 31 million pure and characterized chemical compounds and close to 75 million substance mixtures, extracts, complexes, and other uncharacterized substances. Two other public databases, PDSP \(^{112}\) and ChEMBL \(^{142}\), are popular within the cheminformatics community. PDSP currently contains 55,440 competitive inhibition assay data for psychoactive
compounds, and ChEMBL includes 8,372 targets, 1,000,470 distinct compounds, and 4,668,202 activities.

In 2010, after seeing the potential of open innovation, GlaxoSmithKline publically released 13,471 molecules that had been screened for activity against malaria. This move marks the first large-scale public release of experimentally tested chemical compounds by a pharmaceutical company. These compounds and their associated screening data are available via CDD (Collaborative Drug Discovery) Public\textsuperscript{40}. As of January, 2011, CCD Public contains 69 datasets for a total of about 1.5 million small molecules\textsuperscript{40}.

In addition to small-molecule databases, many public databases provide access to larger biological molecules. For instance, Protein Data Bank (PDB) is a repository for the three-dimensional x-ray crystallography or NMR spectroscopy data of large biological molecules such as proteins and nucleic acids. As of January 13, 2011, the PDB contains 70,494 protein structure entries.

1.1.2 Overview of Computational Methods Employed in Drug Discovery

Cheminformatics and bioinformatics tools, such as statistical classification methods, have proven to be reliable in handling and analyzing large datasets\textsuperscript{16}. However, the explosion of publicly available biological and chemical data has lead to a critical need for modifications of existing and developments of new cheminformatics and bioinformatics tools for integration of the data\textsuperscript{101}. These publicly available data could serve as a platform for computer-aided drug design (CADD) that refers to discovery of new molecules with desirable properties through computational methodologies. There are two major types of methods utilized in CADD: structure-based and ligand-based. Structure-based methods utilize knowledge of the three dimensional structure of a biological target (e.g., protein). However, many targets lack experimental structures, in which case, a homology model based on the experimental structure
from a related protein may be used. Even worse, in many cases the biological target associated with a disorder is unknown, and structure-based drug design cannot be used.

On the other hand, ligand-based methods do not require three-dimensional structural information of biological targets. Instead, ligand-based methods, which are also referred to as ligand-based drug designs (LBDD), only require one or more chemical compounds that display a particular experimentally measured activity, thus allowing a broader range of applicability than its structure-based counterpart. More specifically, LBDD identifies the structural characteristics for a molecular compound, usually referred to as descriptors. These descriptors describe the multi-dimensional features of a compound, e.g., molecular weight, topology, volume, and are in turn applied to estimate biological activity.

The assumption in LBDD is that structurally similar compounds will possess similar biological activity. This structural similarity can be assessed either globally or locally using descriptors. A global similarity search can work with only a single active compound, making it especially useful in earlier phases of CADD where one does not have enough information about the biological targets and few binding ligands are available. However, if only one active compound is used, a global similarity search will utilize all descriptors, including those irrelevant to the biological activity of interest. In contrast, local similarity search methods can identify molecular descriptors relevant to the biological activity, but they also require more compounds known to have the requested biological activity. Because more and more experimental screening data are being made available, LBDD using local similarity is becoming increasingly applicable.

1.1.3 Applications of Classification Methods in Cheminformatics and Bioinformatics

Biological activities can be generalized into two types: continuous and categorical. For modeling categorical data, there are two learning methods to group data: unsupervised and supervised. In unsupervised learning (referred to as cluster analysis), the problem is to analyze a single dataset and decide how and whether the observations in the dataset can be divided into
groups. Supervised learning (referred to as classification) is a method of assigning unknown entities into known groups. The goal is to learn from training sets and then apply the knowledge to test sets. Thus, entities from test sets are placed into established groups, i.e., active or inactive, based on their measurable quantitative characteristics. Although both classification and cluster analysis determine which group a compound belongs to based on the quantitative measurements, the sorting of groups associated with classification attempts to identify the contributions of the quantitative measures to the established groups. In fact, classification, as a method for information extraction, has been applied to many fields, including cheminformatics and bioinformatics, which are important in the drug discovery process.

In bioinformatics, classification is applied from microarray gene expression data to proteins. In gene expression analysis, the goal is to separate signal from noise in high-throughput gene expression studies. Another important application of classification in bioinformatics is classifying proteins into groups. The grouping, which is well established, can be based on the similarities in structures or functions of proteins. The functional grouping of proteins can be found by the Enzyme Classification (EC) number while Structural Classification of Proteins (SCOP) provides the structural grouping of proteins. Given a group of proteins, the goal of classification is to identify common patterns (or motifs) that are conserved.

In cheminformatics, quantitative structure-activity relationship (QSAR) modeling relies on machine learning methods to correlate molecular descriptors to well-defined biological activities, which can be categorized into groups, such as inhibitors, weak inhibitors, and non-inhibitors. In the scenario of categorized biological activity, classification is used to build models from the molecules in the training set and then to predict the biological activities of unknown molecules through these models. These models select molecular descriptors that are relevant to the biological activity from a population of descriptors determined either empirically or by computational methods. Selected descriptors, which encode structural or property parameters, provide clues to understanding the structural requirements for compounds to exhibit biological
activity. With the amount of biomolecular data available to the public, classification is ideal to analyze these data and to identify important molecular features attributing to the given biological activity.

1.2 Research Motivation

Modern QSAR studies are characterized by the use of multiple descriptors of chemical structure combined with linear or non-linear machine learning methods in an attempt to build predictive QSAR models through rigorous model validation. As summarized by Tropsha et al.\textsuperscript{136}, the major differences between various QSAR paradigms are due to the different molecular descriptors and the algorithms used to establish a correlation between descriptor values and biological activity; however, there appears to be no universal QSAR paradigm that produces the best QSAR models for any datasets. The combination of the significant increase in publicly available datasets of biologically active compounds and the critical need to improve the hit rate of experimental compound screening has created a strong need to develop reliable computational QSAR modeling techniques and specific end-point predictors, i.e. a specific set of variables expected to predict a biological activity (end-point).

The challenges associated with modeling these biomolecular data include (1) high dimension low sample size (HDLSS), i.e., when the embedded dimension (e.g., number of descriptors) far exceeds the sample size (e.g., number of molecules in the dataset), and (2) imbalanced categorical data, i.e., when the number of samples in one class far exceeds that of the other. HDLSS data is challenging largely due to the high likelihood of overfitting. Overfitting occurs when a statistical model captures noise bias toward the training set instead of identifying specific end-point predictors that describe the underlying biological activity relationship. Thus, overfitting generally lowers the prediction accuracy for samples that are not part of the training set.
Another challenge associated with modeling the biomolecular data is the imbalanced class distribution of categorized biological activity, which means that there is at least one class of instances which significantly outnumbers other classes. This challenge is also referred in this dissertation as the imbalanced categorical characteristic. Some modern classification methods, which optimize the overall misclassification rate of the whole training set, do not perform well on data with imbalanced categorical characteristic, because such methods generally assume a relatively balanced class distribution and put too much strength on the majority class.

There is a middle ground between using simplistic models that are traditional in biochemistry and letting powerful computers free to do data mining on a huge number of potential features. In this dissertation, we demonstrate the benefit of choosing models that attempt to capture a (relatively) small number of chosen features that are then subjected to statistical analysis. Specifically, we show that

(i) estimating the intrinsic dimension of a dataset can improve DWD statistical learning, and overcome the statistical difficulties inherent to biological data with high dimension, low sample size (HDLSS) and imbalanced categorical characteristics;

(ii) novel, three-dimensional chiral atom-pair descriptors for stereochemistry-dependent datasets produce more accurate QSAR models; and

(iii) new protein descriptors based solely on structure aid automatic function annotation, especially in cases of function homologs with low sequence similarity.
Chapter 2

Variable Selection Based Classification Method for Imbalanced and HDLSS Data

2.1 Motivation

Classification \(^{50}\) as a method for information extraction is a statistical tool that has been applied to many fields, including QSAR and micro-array analysis. The typical biomolecular data associated with QSAR and micro-array analysis have the characteristics of High Dimension Low Sample Size (HDLSS) and imbalanced categorical data. The defining characteristic of HDLSS is when the dimension of the data vectors is larger (often much larger) than the sample size (the number of data vectors available). A major challenge associated with modeling HDLSS data is the problem of overfitting, which occurs in the event that a statistical model is driven by noise artifacts in the training set instead of the underlying relationship. Thus, overfit models generally have poor predictive performance for unseen data. It has been demonstrated that the Support Vector Machine \(^{140}\) (SVM), which is a clever and powerful discrimination method, suffers from the data piling effect at the margin (overfitting) for HDLSS data, and as a result, generalizability is diminished \(^{92}\).

The defining characteristic of imbalanced categorical data is when at least one class of instances (major class) significantly outnumbers other classes (minor classes). Ding suggests a five percent threshold to distinguish a significantly imbalanced categorical dataset from a moderately imbalanced one \(^{47}\). Based on Ding’s threshold, a binary classification dataset is considered significantly imbalanced if the size of a minor class is no more than five percent of the entire data size. One major problem associated with modeling imbalanced categorical data is that some modern statistical learning methods, e.g., the decision tree and support vector machine,
optimize the overall misclassification rate and treat all classes equally. Optimization of this type of classification criterion can be problematic because the minority classes tend to be ignored or discounted during the classification due to their small proportions. This can be a serious problem if those minority classes are important, which is often the case in QSAR and micro-array analysis. Current solutions to classify the imbalanced categorical data include upsampling the minor classes, downsampling the major classes, and/or changing the optimization criteria (such as the correct classification rate, or CCR). CCR, which is the average of sensitivity and specificity in the case of binary classification, is an optimization criterion that can be applied to both balanced and imbalanced categorical data. However, the ultimate goal should be finding an algorithm that classifies the data without removing, re-sampling or modifying the set.

To address problems associated with HDLSS and imbalanced categorical characteristics, a modification of the existing Distance Weighted Discrimination (DWD) has been proposed. DWD, which is a linear classifier and operates by splitting a high-dimensional input space with a hyperplane, performs binary classification by projecting the data onto a DWD direction. This DWD direction, which is a real vector of weights corresponding to the input space, is relatively insensitive to the imbalanced categorical dataset; however, the previous optimization to determine the location of the hyper-plane (threshold), which separates the positive class from the negative class, was not optimized for the case of imbalanced categorical data. In more recent work, the location of the hyper-plane has now been optimized using weighted Distance Weighted Discrimination (wDWD) for both balanced and imbalanced categorical data.

However there is still a problem that even wDWD does not fully address: the actual separation may exist in a lower dimension space instead of in the full feature space. In other words, not all the features are important to a given biological property, i.e., only some of the chemical descriptors may be relevant. Although both versions of DWD assign weights (loadings) to features (or descriptors in QSAR), those weights are typically non-zero values. Therefore features that are not relevant to the biological property are still considered, which can result in
overfitting, especially in the HDLSS setting. Our proposed method intends to strengthen this aspect. Due to the classification performance improvement of wDWD over the original DWD and because the loading values assigned in both methods are identical, wDWD is utilized exclusively throughout this research. From this point forward, DWD will refer to wDWD.

2.2 Overview of Variable Selection

In working with HDLSS data, potential overfitting is a serious problem. To minimize the effect of overfitting, we implemented a method of variable selection to identify specific end-point predictors (predictive features or descriptors) that describe the underlying biological activity relationship. Although variable selection can be applied to both unsupervised and supervised learning, the focus of this research is the selection of a subset of relevant features for building predictive learning models.

As pointed out by Guyon et al, the primary objectives of variable selection are:

- To improve the prediction performance of the models
- To provide faster and more cost-effective models by identifying the predictor variables
- To obtain a better understanding of the underlying process that generated the data

These objectives are especially critical for datasets with tens or hundreds of thousands of available variables, which are frequently encountered in the field of bioinformatics and cheminformatics.

Techniques for variable selection can be summarized into three different types: filter, wrapper, and embedded. Filter techniques select subsets of variables as a preprocessing step using selection criteria that are independent of the chosen classifier, possibly based on information gain (univariate) and/or correlation (multivariate). However, by discounting the interaction with the classifier, filter techniques can also limit classification performance. In addition, univariate
versions of the filter technique are even more likely to achieve lower classification performance because they discount the dependencies between features.

Wrapper techniques utilize the machine learning of interest to measure the quality of feature subsets according to their predictive power and can therefore be combined with any learning machine. Wrapper techniques such as sequential forward selection (deterministic) and simulated annealing (stochastic) can improve classification performance by interacting with classifiers. However, the performance gain comes at the price of higher computational cost as the method is typically computationally intensive. Another critical drawback associated with wrapper techniques is the higher risk of overfitting and getting stuck in a local optimum.

Embedded techniques perform variable selection in the training process and are usually specific to given learning machines. In contrast to filter and wrapper techniques, the learning and the feature selection procedures in embedded techniques cannot be separated. The advantage of embedded techniques over wrapper techniques is in computational complexity. The benefit of being less computationally intensive makes embedded techniques more attractive than wrapper techniques. Feature selection using the weight vector of SVM is an example of an embedded technique. Other embedded techniques include decision trees and weighted naïve Bayes.

### 2.3 Overview of the Proposed Method

To address the problems associated with imbalanced HDLSS categorical data, we implemented a variable evaluation and selection method to couple with DWD. This variable evaluation and selection method contains two components: variable evaluation and variable selection. The variable evaluation component utilizes a permutation test to evaluate how well a set of descriptors can separate two classes without setting aside additional data. In this permutation test, a value indicating the significance of a descriptor set to a categorized biological activity is calculated by comparing the separation obtained from the original label to the separations obtained from a population of permuted labels. This value of significance is then
used to compare a different set of top ranked descriptors and to estimate the intrinsic dimension by identifying the most significant set of top ranked descriptors.

The variable selection component utilizes an embedded variable selection technique that takes advantage of the weight vector from DWD, which is a linear classifier that makes a classification decision based on the value obtained from the dot product of the weight vector and the feature vector (descriptors of a molecule). Each descriptor has a corresponding weight by design; therefore, based on the absolute value of the weights, it is possible to generate a ranked list of the descriptors. This ranked list serves as a reference for selecting different sets of top ranked descriptors, e.g., top 10 and top 20, to incorporate into the new DWD models. Each of these DWD models is then evaluated for significance through the variable evaluation component of the method, and only the most significant model with the corresponding descriptors is selected.

2.3.1 Variable Evaluation Component

To evaluate how well a set of descriptors can separate two classes, a procedure known as y-randomization, i.e., positive and negative class labels are randomly assigned to each compound, is performed $N$ times ($N = 1,000$ for QSAR studies). For each y-randomization, the original ratio of positive to negative labels is maintained, and a new model is generated (random model) with the same set of descriptors. Then the classes are projected onto the DWD direction where the decision for binary classification occurs. To quantitatively estimate the separation between the two classes, a mean-difference is computed by calculating the distance between the center of the positive class and the center of the negative class in the projected DWD direction, which is generated based on a given set of descriptors $^{144}$. By comparing the model built with original labels (original model) against the population of random models, the significance of the original model can be quantified by two different $p$-values: an empirical and a Gaussian fit.

Both empirical and Gaussian fit $p$-values estimate the likelihood that a random model will have better separation than the original model, but the Gaussian fit $p$-value improves the
significance estimation over the empirical p-value by approximating a Gaussian distribution curve based on the mean-difference values of the random models and calculating the area under the curve with mean-difference values better than that of the original model. However, when comparing the different sets of top-ranked descriptors, there is a potential problem that the likelihood cannot be used to accurately estimate significance because both the empirical and Gaussian p-values could be too small to be detected. To solve this problem, we implemented an alternate criterion, a z-score, to indicate the significance of a set of top ranked descriptors. This z-score, i.e., the number of standard deviations of the original model mean-difference from the population mean for the mean-difference values of the y-randomization models, will yield a value in scenarios when both empirical and Gaussian p-values are too small to detect; thus the z-score is more suitable when comparing different sets of descriptors.

2.3.2 Variable Selection (Ranking) Component

The proposed variable selection technique relies on the initial weight vector of DWD obtained from the full descriptor set to generate a ranked list. This ranked list is sorted in a descending order based on the absolute value of the weights. Since each weight corresponds to a descriptor, new classification models are built with a different number of top ranked descriptors.

The variable selection technique utilizes a greedy algorithm to search for the optimal set of top ranked descriptors to incorporate in the final model. The initial search step is to identify the interval where the combination of top ranked descriptors is likely to achieve the most significance. The search begins by considering a coarse logarithmic series of sets of partial descriptors, e.g., the top 500 descriptors, top 200 descriptors, and top 100 descriptors, which are incorporated in the new models and evaluated for model significance. In the region where the z-score values are much higher, further searches with smaller step size are performed to identify the optimal set of top ranked descriptors for the final model.
2.5 Simulation

2.5.1 Experimental Design

A simulation was designed to benchmark DWD with and without variable selection for imbalanced HDLSS categorical data to evaluate the performance gained by coupling DWD with variable selection. In addition, a version of SVM without variable selection was added into the benchmark to compare the difference in performance between DWD and SVM.

The data for this simulation was designed to be imbalanced, with the training set containing 21 actives and 80 inactives while the external set contained 200 actives and 800 inactives. To ensure the HDLSS characteristic of the data, there were 731 features associated with each entry (active or inactive). The simulation was designed to contain only 50 informative descriptors. The values of the informative descriptors for the actives were randomly sampled from the normal distribution with a mean.

![Figure 2.1. Informative descriptor vs. noise.](image)

The y-axes in both plots, which are the density estimations, are associated with the curves and not the markers (blue and red circles). The locations of the markers are based on the order of the molecules in the dataset. The feature (descriptor) in the top plot encodes some information to separate the positive class (blue circles) from the negative class (red circles) while the feature in the right plot is pure noise (bottom plot).
of -0.50 and a standard deviation of 1.0, denoted as N(-0.50, 1.0). For the inactives, these values were sampled from N(0.50, 1.0). As for the remaining descriptors, which were designed to be noise, the values were randomly sampled from N(0.0, 1.0). The distributions of both the active class and the inactive class in informative descriptor vs. noise are shown in the figure below (Figure 2.1). The purpose of this simulation is to test how well the DWD with variable selection could retrieve the 50 features that do contain information to separate the two classes. In addition, this simulation is designed to avoid a single magic descriptor that could separate the two classes; the most significant separation should incorporate a significant portion of the 50 features while minimizing the noise. This simulation was replicated 30 times.

In addition, five-fold cross-validation was performed in the training set during the modeling procedure for both SVM and DWD to tune the penalty parameter, which adjusted the associated projected direction (e.g., DWD direction) for each corresponding linear classifier. The values considered for the penalty parameter range from 2 to 1,024 with the value doubled in each step. After identifying the optimal value for the penalty parameter, a single model was built using all the available data within a given training set. Each resulting model was then used to predict the actives and inactives in the corresponding test set.

### 2.5.2 Simulation Result

The prediction results of the test sets from the 30 simulation runs are summarized by the box-and-whisker plots shown in Figure 2.2, which shows the sensitivity, specificity, and correct classification rate (CCR) over the 30 test runs. The results of the simulation showed that all three methods performed similarly in specificity. With the implemented variable selection (V.S.), DWD consistently achieved a higher sensitivity and CCR than the other two methods.
Analyzing the 30 optimal models obtained from DWD-VS indicates that the number of descriptors incorporated in the model was frequently 30 or 35; however, one instance out of 30 simulations yielded an optimal model with 40 descriptors. Figure 2.3 identifies the optimal model and the searching process for the optimal set of top ranked descriptors in one of the simulation runs. The initial search considered a coarse logarithmic series of partial descriptor sets and identified the interval where the optimal number of top ranked descriptors to incorporate into the model was likely to occur. Another search with a step size of ten was applied to this particular interval, which covered the range between the top 20 and the top 200 ranked descriptors, and identified that the highest z-score was associated with the model built with the top 30 ranked

![Test Set Results (30 Runs)](image)

**Figure 2.2.** Prediction results of the 30 simulation runs.

The box-and-whisker plots show the sensitivity, specificity, and CCR of the 30 test sets. The median values are indicated by circles with a black dot inside. The lower and upper ends of the boxes indicate the lower quartile and upper quartile, respectively, with standard whisker lengths of 1.5 times of the interquartile range, i.e., the height of the box. Values outside the whiskers are considered as performance outliers and represented by red pluses.
descriptors. One last search with a step size of 5 was performed in the region between the top 20 to the top 110 ranked descriptors in order to approximate the location of the optimal z-score.

![Z-Score vs. Top Ranked Descriptors](image1)

![Z-Score vs. Top Ranked Descriptors](image2)

**Figure 2.3.** Identification of the optimal model through the z-score.

The initial search considered a coarse logarithmic series of partial descriptor sets and identified the interval where the optimal number of top ranked descriptors to incorporate into the model was likely to occur (left plot). Once the region of high z-scores was identified, additional searches were performed with smaller increments of the number of top ranked descriptors. In this particular run, the optimal model was the one that incorporated the 30 top ranked descriptors (right plot).

To further analyze the performance of the implemented variable selection, recall and true negative rate were calculated based on the descriptors selected by each of the optimal models obtained from DWD-VS. The recall is the percentage of the 50 informative descriptors selected
by the model, and the true negative rate or the noise removal rate is the percentage of the noise excluded by the model. The true negative rate for each of the 30 simulation runs was close to 100%; however, the recall for the informative descriptors ranged only from 0.58 to 0.78, with a median of 0.60 (Figure 2.4). The recall and the true negative rate calculated from the 30 simulation run indicated that the implemented variable selection was capable of improving the classification result by removing most of the noise while retaining the majority of the informative descriptors.

The outcome of this simulation shows that the z-score proved to be a strong indicator for model performance and a helpful parameter for model optimization. Since the calculation of the z-score does not require setting aside additional data, the resulting models also have the

![Model Analysis](image)

**Figure 2.4.** Model analysis of the simulated data.

In this simulation, the optimal model usually selected 30 out of the 50 informative descriptors while removing most of the noise.
advantage of using all the data available. This advantage is critical for modeling the imbalanced categorical dataset in the HDLSS setting.

### 2.6 QSAR Studies

For the QSAR studies, all three methods, i.e., SVM without VS and DWD both with and without VS, were applied to build binary classification models for five different datasets. The first dataset contains 110 compounds that include inhibitors and non-inhibitors for AmpC β-lactamase. The other four datasets are the binders and non-binders for the different families of serotonin receptors (5-HT receptors). For each of these datasets, Dragon^3^ descriptors, which include both physicochemical and structural properties of molecules, were generated. Descriptors with zero variance were removed from the generated descriptor matrices. For highly correlated descriptors within the descriptor matrix that achieve pairwise correlation close to 1.00 from one another, an additional step was taken by selecting only one descriptor as representative. The resulting descriptor matrix and the corresponding categorized biological activity for each dataset were partitioned into modeling and test sets, and all three methods were built with the same modeling sets and applied to the same test sets for the benchmark.

In QSAR studies, model validation is an important part of the workflow. To make the model validation process more generalizable, a five-fold cross-validation procedure was applied to the data partitioning of modeling sets and test sets (external validation set). Within each modeling set, another five-fold cross-validation procedure was applied for model tuning by further partitioning the modeling set into both training and validation sets (internal test set). To distinguish the two procedures, the five-fold cross-validation procedures associated with model validation and model tuning were denoted as five-fold external cross-validation and five-fold internal cross-validation, respectively.

For all the QSAR studies in this research, five-fold internal cross-validation was applied to estimate the optimal penalty parameters for both SVM and DWD. DWD was tuned without
variable selection and those tuning parameters were used throughout the variable selection process. After the optimal penalty was identified through the five-fold internal cross-validation process, a single model was then built based on the complete modeling set data (using the union of the training set and the validation set). Each complete model was then applied to classify the corresponding test set. To summarize the classification result of the test sets, the prediction outcomes from the 5 test sets were combined. Sensitivity, specificity, and CCR were calculated from this combined set of outcomes.

2.7 QSAR Datasets Description

2.7.1 AmpC β–lactamases Dataset Description (110 Compounds)

The β-lactam ring is an essential structure of several antibiotic families, such as the penicillins, cephalosporins, carbapenems, and monobactams. Due to the commonality of this particular active structural feature among these chemical compounds, they are collectively called as the β-lactam antibiotics. These chemical compounds gain their antibiotics status by inhibiting bacterial cell wall synthesis. The inhibition of cell wall synthesis has a lethal effect on bacteria, especially on the Gram-positive bacteria that are characterized by the high amount of peptidoglycan in the cell wall. However bacteria can become resistant against β-lactam antibiotics by expressing β-lactamase, an enzyme that is produced by Gram-negative organisms and has the ability to break open the β-lactam ring thus deactivating the antibacterial properties. In 1940, AmpC β-lactamase of Escherichia coli was the first bacterial enzyme reported to destroy penicillin. Since the discovery of the β-lactamases and their attributions toward antibiotic resistance, there has been a significant amount of efforts in the scientific community to identify compounds that inhibit β-lactamases to work in conjunction with antibiotics.

A dataset containing AmpC β-lactamases inhibitors and non-inhibitors was published by Shoichet’s group. In this data, competitive binding (K_i in μM) was measured for all the molecules. Molecules with K_i less than 1,000 μM were considered inhibitors and molecules with
K_i greater or equal to 1,000 µM were considered non-inhibitors. The range of inhibition (K_i values) is from 1.0 to 646.0 µM. Molecules with smaller K_i are considered strong inhibitors but for the classification, there is no distinction between strong inhibitors and weak inhibitors. The published data contains 21 inhibitors and 84 non-inhibitors. Additional data were later provided by Shoichet’s group that includes five additional inhibitors of the same chemical series.

2.7.2 5-HT Datasets Description

The serotonin receptors, also known as 5-Hydroxytryptamine (5-HT) receptors, influence various biological and neurological processes such as aggression, anxiety, appetite, cognition, learning, memory, mood, nausea, sleep, and thermoregulation. The serotonin receptors are the target of a variety of pharmaceutical and illicit drugs, including many antidepressants, antipsychotics, anorectics, antiemetics, gastroprokinetic agents, antimigraine agents, hallucinogens, and entactogens [3].

There are 7 different 5-HT receptor families: 5-HT_1, 5-HT_2, 5-HT_3, 5-HT_4, 5-HT_5, 5-HT_6, and 5-HT_7. These 5-HT receptor families can be further characterized in subtypes (shown in Table 2.1). With the exception of the 5-HT_3 receptors, which are ligand-gated ion channel, all other serotonin receptors are G protein-coupled receptors that activate an intracellular second messenger cascade to produce an excitatory or inhibitory response.

A collection of binders and non-binders associated with 5-HT_1B, 5-HT_1D, 5-HT_2B, and 5-HT_6 receptors are obtained from PDSP [112]. In the collected data, there are 91 binders and 79 non-binders associated with 5-HT_1B receptors. As for 5-HT_1D receptors, the numbers are 87 and 81 for
binders and non-binders respectively. Comparison between the binders of 5-HT_{1B} and 5-HT_{1D} receptors indicates 70 overlapping binders that bind to both receptors. However, there is no evidence in the data to suggest that the non-overlapping binders are specific to their corresponding receptors.

2.8 QSAR Modeling Results

2.8.1 AmpC B-lactamase Modeling Result (110 Compounds)

In total 894 two-dimensional (2D) molecular descriptors were generated for these 110 compounds with commercially available software, Dragon^3. As mentioned earlier (Section 2.6), five-fold external cross validation was implemented to perform the QSAR study for this dataset. The classification results of the five test sets were combined into a single set to calculate the

![Combined Result of Test Sets (AmpC Dataset)](image)

**Figure 2.5.** Performance comparison between the methods (AmpC dataset).

The bar graph shows the test set result of the five-fold external cross validation. The predictions for each of the five test sets were combined into a single set to calculate the sensitivity, specificity, and CCR. In the AmpC dataset, both DWD methods achieve better classification outcomes than SVM; however, the classification results between the two DWD methods are quite similar.
sensitivity, specificity, and CCR (shown in Figure 2.5). The three methods performed similarly in classifying the inhibitors vs. non-inhibitors in the test sets. Compared to SVM, DWD with variable selection was able to give correct predictions for one additional inhibitor and two non-inhibitors. The difference in specificity between the two DWD methods was caused by the correct prediction of one non-inhibitor. The small difference in classification performance could be explained by the structure similarities of the inhibitors. The chemical structures of the majority of inhibitors in this dataset belonged to a chemotype that can be characterized by a sulfonamide bridging two aromatic rings, with one aromatic ring containing a carboxylic functional group. Molecular descriptors reflecting the characteristic of this chemotype were highly ranked in both DWD and DWD-VS, thus causing the similarity in classification performance between the two methods. However, DWD-VS did have an advantage over the DWD in model interpretation by identifying a much smaller set of descriptors that are significant to a biological property.

Analyzing the models obtained from DWD-VS indicated that the numbers of descriptors incorporated in each of the five folds were 100, 150, 150, 250, and 100. Cross-checking these descriptors yielded 315 unique descriptor

<table>
<thead>
<tr>
<th>Descriptor Name</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td>Mp</td>
<td>nSO2N</td>
</tr>
<tr>
<td>nS</td>
<td>nThiophenes</td>
</tr>
<tr>
<td>T(O..S)</td>
<td>C-027</td>
</tr>
<tr>
<td>MATS4v</td>
<td>C-029</td>
</tr>
<tr>
<td>MATS3e</td>
<td>C-033</td>
</tr>
<tr>
<td>MATS4e</td>
<td>C-034</td>
</tr>
<tr>
<td>MATS6e</td>
<td>H-047</td>
</tr>
<tr>
<td>MATS4p</td>
<td>H-048</td>
</tr>
<tr>
<td>GATS2m</td>
<td>H-049</td>
</tr>
<tr>
<td>GATS1v</td>
<td>O-057</td>
</tr>
<tr>
<td>GATS2v</td>
<td>O-060</td>
</tr>
<tr>
<td>GATS4v</td>
<td>N-067</td>
</tr>
<tr>
<td>GATS5v</td>
<td>N-075</td>
</tr>
<tr>
<td>GATS6v</td>
<td>Inflammat-80</td>
</tr>
<tr>
<td>GATS5e</td>
<td>Infective-50</td>
</tr>
<tr>
<td>GATS6e</td>
<td>F01[C-N]</td>
</tr>
<tr>
<td>GATS4p</td>
<td>F01[N-S]</td>
</tr>
<tr>
<td>EEig01x</td>
<td>F02[C-S]</td>
</tr>
<tr>
<td>BELm3</td>
<td>F03[C-O]</td>
</tr>
<tr>
<td>BEHe8</td>
<td>F03[N-S]</td>
</tr>
<tr>
<td>BEHp1</td>
<td>F04[N-O]</td>
</tr>
<tr>
<td>BELp3</td>
<td>F04[N-S]</td>
</tr>
<tr>
<td>JGI9</td>
<td>F04[O-S]</td>
</tr>
<tr>
<td>nArCOOH</td>
<td>F05[O-O]</td>
</tr>
<tr>
<td>nArCONHR</td>
<td></td>
</tr>
</tbody>
</table>

Table 2.2. The 49 unique descriptors selected by all 5 models.

The highlighted descriptors reflect the common structure features that distinguished the inhibitors from the non-inhibitors.
names. Among these unique descriptors, 49 were selected by all the five models. These 49 descriptors are shown in Table 2.2.

Although not all the descriptors from Dragon are easily interpretable, some can be easily mapped back to the molecules. For example, nSO2N, nThiophenes, and nArCOOH are the descriptors that reflect the common structure features of the inhibitors. Visual inspection of the inhibitors indicates that the molecular structures usually contain a sulfonamide linking two aromatic substituents (Figure 2.6). One of the aromatic substituents should include a carboxylic acid. These observations match with the descriptors selected by the models. The nArCOOH is a descriptor that indicates the occurrence of an aromatic substituent with carboxylic acid in a molecule. As for nSO2N and nThiophenes, these are the occurrences of sulfomamide and thiophene respectively.

![Figure 2.6. Common structure features of the AmpC inhibitors.](image)

Most of the inhibitors have a sulfonamide (orange) linking two aromatic substituents. One aromatic substituent must contain a carboxylic acid (blue). Thiophene (purple) is considered as an aromatic substituent.
2.8.2 5-HT\textsubscript{1B} Modeling Result

In total 875 2D Dragon descriptors were generated for the 170 structurally diverse compounds. Building global models yielded poor and inconsistent classification performance across the folds. To improve the classification performance, the modeling approach was to partition the data into clusters and to build specific models for each cluster.

Unsupervised clustering analysis of the 170 compounds was performed based on the 875 chemical descriptors. A dendrogram was constructed based on hierarchical clustering with Ward linkage (Figure 2.7). The dendrogram indicates how the data can be subdivided into clusters. Local models were built by clustering the data into either 2 or 3 groups. Building local models for the three clusters yielded better and more consistent results than building local models for the two higher level clusters. In order to obtain some insight regarding the clusters, a principal component analysis (PCA) plot was generated to visualize the distributions of both clusters and different classes of compounds (Figure 2.8).
There are 41 binders and 35 non-binders in the red cluster. The green cluster contains 18 binders and 24 non-binders. As for the blue cluster, the numbers of binders and non-binders are 32 and 20 respectively. Five-fold external cross-validation procedure was applied for each cluster. However, the cluster labels were only applied for the training sets and not the test set.

To predict the compounds in the test sets, a neighborhood search was performed in the full descriptor space. For each compound in the test set, its nearest neighbor in the training set was identified. The decision of model selection for predicting a given test set compound was based on the nearest training set compound in each cluster. For any given data in the test set, its nearest neighbor in the training set will determine the cluster membership. Based on the cluster

**Figure 2.8.** Data distribution of the 5-HT\textsubscript{1B} dataset in Dragon chemical descriptor space.

The different colors (red, blue and green) in the plots indicate the three clusters identified in hierarchical clustering. Binders and non-binders are represented as circles and crosses respectively. In the two clusters scheme, the data in green and red are merged into a single cluster.
membership, the corresponding local model will be applied for prediction. The classification results from the five-fold external cross-validation of the three clusters were combined into a single set to calculate the sensitivity, specificity, and CCR. As indicated by the results in Figure 2.9, all three methods performed well after clustering the 5-HT\textsubscript{1B} data. DWD-VS was able to correctly predict five more binders than SVM and DWD without variable selection. SVM also misclassified five more non-binders than either DWD method.

![Combined Result of Test Sets (5-HT\textsubscript{1B} Dataset)](image)

**Figure 2.9.** Performance comparison through five-fold external cross validation for the 5-HT\textsubscript{1B} dataset.

For both the 5-HT\textsubscript{1B} and the 5-HT\textsubscript{1D} datasets, local models were built based on the clusters suggested by hierarchical clustering. Selection of a model to predict a compound in the test set was based on identifying its most similar compound in the training compounds. All three methods performed well in this 5-HT\textsubscript{1B} dataset.

Analyzing the descriptors incorporated in the optimal models from DWD-VS yielded 169 unique descriptors for the red cluster, 173 for the green cluster, and 119 for the blue cluster.

Among the 169 unique descriptors identified for the red cluster, there are 26 descriptors which
showed up in all models. For the green and blue clusters, the numbers of unique descriptors selected by all models were 33 and 12, respectively. These selected descriptors are listed in Table 2.3 according to the membership of the clusters. The overlap of selected descriptors between clusters was minimal. Between the green and the blue clusters, the two descriptor lists did not overlap. Five descriptors, which are nR12, nRNR2, C-006, N-068, and O-56, were incorporated in all the models for both red and blue clusters. As for the red and green clusters, both incorporated the descriptors, nRCOOH and nPyrroles, in all of their respective models.

**Table 2.3.** The unique descriptors selected by all 5 models from each of the three clusters.
2.8.3 5-HT\textsubscript{1D} Modeling Result

In total 870 2D Dragon descriptors were generated for the 168 structurally diverse compounds. Similar to the outcome for the 5-HT\textsubscript{1B} dataset, local models yielded better classification performance than global models. Hierarchical clustering of the 5-HT\textsubscript{1D} dataset with Ward linkage produced the following dendrogram (Figure 2.10). Due to a significant amount of overlapping compounds between the 5-HT1B and the 5-HT1D datasets, local models were built based on clustering the 5-HT1D dataset into 3 groups in an attempt to compare the results. The PCA plot below shows the data distribution of the 3 clusters in the chemical space (Figure 2.11). The data distribution of 5-HT\textsubscript{1D} in the PCA plot is similar to that of the 5-HT\textsubscript{1B}. The largest cluster (red cluster) contains 61 binders and 41 non-binders while the smallest cluster (green cluster) includes 11 binders and 16 non-binders. As for the blue cluster, the numbers of binders and nonbinders are 15 and 24, respectively.

\begin{figure}[h]
\centering
\includegraphics[width=0.5\textwidth]{dendrogram.png}
\caption{Dendrogram of the 5-HT\textsubscript{1D} dataset.}
\end{figure}

Due to a significant amount of overlapping compounds between the 5-HT\textsubscript{1B} and the 5-HT\textsubscript{1D} datasets, local models were built by clustering the 5-HT\textsubscript{1D} dataset into 3 groups in an attempt to compare the results.
Five-fold external cross-validation procedure was applied to each of the cluster to obtain the corresponding optimized models. Once again, the cluster labels were only applied for the training sets and not the external test set. To predict the compounds in the test sets, a neighborhood search was performed in the full descriptor space. For any given data in the test set, its nearest neighbor in the training set will determine the cluster membership. Based on cluster membership, the corresponding local model will be applied for prediction. The classification results from the five-fold external cross-validation of the three clusters were combined into a single set to calculate the sensitivity, specificity, and CCR. As indicated by the results in Figure 2.11, data distribution of the 5-HT$_{1D}$ dataset in Dragon chemical descriptor space. The different colors (red, green, and blue) in the plots are to indicate the three clusters identified in hierarchical clustering. Binders and non-binders are represented as circles and crosses respectively. In the two clusters scheme, the data in green and red are in a single cluster. The relative position of the three clusters identified for the 5-HT$_{1D}$ dataset is similar to the three clusters associated with 5-HT$_{1B}$.
2.12, all three methods performed well after clustering the 5-HT\textsubscript{1D} data. DWD without variable selection was able to correctly predict five additional binders more than SVM and DWD-VS. SVM also misclassified ten more non-binders than either DWD method.

Analyzing the descriptors incorporated in the optimal models from DWD-VS method yielded 328 unique descriptors for the red cluster, 251 for the green cluster, and 120 for the blue cluster. Among the 328 unique descriptors identified for the red cluster, there are 57 descriptors that showed in all models. For the green and blue clusters, the numbers of unique descriptors selected by all models were 36 and 19, respectively. These selected descriptors are listed in Table 2.3 according to the membership of the clusters.
### Table 2.4
The unique descriptors selected by all 5 models from each of the three clusters.

The highlighted descriptors are selected by the similarly located clusters in both 5-HT<sub>1B</sub> and 5-HT<sub>1D</sub>.

<table>
<thead>
<tr>
<th>Descriptor Name (Red Cluster)</th>
<th>Descriptor Name (Green Cluster)</th>
<th>Descriptor Name (Blue Cluster)</th>
</tr>
</thead>
<tbody>
<tr>
<td>nO nPyrrolidines</td>
<td>ARR GATS7p</td>
<td>nN C-008</td>
</tr>
<tr>
<td>nX nPyrroles</td>
<td>nN EEig13d</td>
<td>nR09 C-012</td>
</tr>
<tr>
<td>nR09 C-006</td>
<td>nF EEig14d</td>
<td>TI2 C-013</td>
</tr>
<tr>
<td>TI2 C-007</td>
<td>nX nCRX3</td>
<td>D/Dr09 N-067</td>
</tr>
<tr>
<td>D/Dr07 C-027</td>
<td>PW5 nPyrroles</td>
<td>D/Dr10 C-034</td>
</tr>
<tr>
<td>D/Dr09 C-033</td>
<td>piPC10 C-013</td>
<td>T(O..S) C-040</td>
</tr>
<tr>
<td>D/Dr10 C-034</td>
<td>X3A C-033</td>
<td>MATS1p H-049</td>
</tr>
<tr>
<td>T(O..S) C-040</td>
<td>MATS6m O-058</td>
<td>GATS5m H-052</td>
</tr>
<tr>
<td>MATS1p H-049</td>
<td>MATS5v N-073</td>
<td>GATS4v N-068</td>
</tr>
<tr>
<td>GATS5m H-052</td>
<td>MATS5e F-083</td>
<td>GATS5v N-069</td>
</tr>
<tr>
<td>GATS4v N-068</td>
<td>MATS6p GVWA1-80</td>
<td>GATS4e N-075</td>
</tr>
<tr>
<td>GATS5v N-069</td>
<td>GATS6m F01[C-F]</td>
<td>GATS5e F-083</td>
</tr>
<tr>
<td>GATS4e N-075</td>
<td>GATS7m F02[F-F]</td>
<td>GATS6v F03[C-F]</td>
</tr>
<tr>
<td>GATS5e Depressant-80</td>
<td>GATS6v F03[C-F]</td>
<td>GATS7v F04[C-F]</td>
</tr>
<tr>
<td>GATS1p Hypertens-50</td>
<td>GATS6e F05[C-F]</td>
<td>GATS7e F05[N-N]</td>
</tr>
<tr>
<td>EEig01x Hypnotic-50</td>
<td>GATS6p F07[N-F]</td>
<td>GATS6e F07[N-N]</td>
</tr>
<tr>
<td>ESpml2x Neoplastic-80</td>
<td></td>
<td></td>
</tr>
<tr>
<td>ESpml3x Neoplastic-50</td>
<td></td>
<td></td>
</tr>
<tr>
<td>ESpml14x F01[C-N]</td>
<td></td>
<td></td>
</tr>
<tr>
<td>ESpml15x F01[C-O]</td>
<td></td>
<td></td>
</tr>
<tr>
<td>BEHv1 F03[C-S]</td>
<td></td>
<td></td>
</tr>
<tr>
<td>BEHe1 F03[N-O]</td>
<td></td>
<td></td>
</tr>
<tr>
<td>BEHp1 F04[C-S]</td>
<td></td>
<td></td>
</tr>
<tr>
<td>VEA1 F05[S-Cl]</td>
<td></td>
<td></td>
</tr>
<tr>
<td>nCrs F06[N-Cl]</td>
<td></td>
<td></td>
</tr>
<tr>
<td>nRCOOH F07[N-N]</td>
<td></td>
<td></td>
</tr>
<tr>
<td>nCONN F07[N-O]</td>
<td></td>
<td></td>
</tr>
<tr>
<td>nRNHR2 F08[N-S]</td>
<td></td>
<td></td>
</tr>
<tr>
<td>nRNR2 F08[N-S]</td>
<td></td>
<td></td>
</tr>
<tr>
<td>nArX</td>
<td></td>
<td></td>
</tr>
<tr>
<td>C-006</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
Comparing these descriptors with those of the similarly located clusters in 5-HT_{1B} indicated some overlapping descriptors. There were nine descriptors that appeared in both red clusters. The numbers of overlapping descriptors between the two datasets were 12 and 6 for the green and blue clusters, respectively. Understanding the contribution of these overlapping descriptors to the binding of 5-HT_{1B} and 5-HT_{1D} receptors could contribute to designing a safer drug.

### 2.8.4 5-HT_{2B} Modeling Result

In total, 1,030 Dragon 2D descriptors were generated for these 753 compounds. The classification results of the five-fold external cross validation were combined into a single set to calculate the sensitivity, specificity, and CCR (shown in Figure 2.13). The difference in classification results between the three methods is minimal. Both DWD methods had higher

![Figure 2.13](image-url)

**Figure 2.13.** Performance comparison through five-fold external cross validation for the 5-HT_{2B} dataset.

For the 5-HT_{2B} dataset, both DWD methods yielded similar performance in classifying binders vs. non-binders from the test sets. In comparison, the sensitivity associated with SVM did show a drop in performance.
sensitivity and specificity than SVM. Comparison with SVM, DWD-VS was able to provide correct predictions for four additional binders and six non-binders. The differences in sensitivity and specificity between the two DWD methods were caused by the correct predictions of two binders and four non-binders.

Analyzing the models obtained from DWD-VS indicated that the numbers of descriptors incorporated in each of the five folds were 250, 250, 200, 400, and 200. Cross-checking these descriptors yielded 482 unique descriptor names. Among these unique descriptors, 79 were selected by all the 5 models. These 79 descriptors are shown in Table 2.5.

<table>
<thead>
<tr>
<th>Descriptor Name</th>
<th>Ms</th>
<th>BELm6</th>
<th>H-049</th>
</tr>
</thead>
<tbody>
<tr>
<td>nO</td>
<td>BEHv1</td>
<td>N-070</td>
<td></td>
</tr>
<tr>
<td>nR07</td>
<td>BELv6</td>
<td>N-071</td>
<td></td>
</tr>
<tr>
<td>nR10</td>
<td>BELe6</td>
<td>N-074</td>
<td></td>
</tr>
<tr>
<td>nR11</td>
<td>BEHp1</td>
<td>Cl-086</td>
<td></td>
</tr>
<tr>
<td>Lop</td>
<td>JGI2</td>
<td>UJ</td>
<td></td>
</tr>
<tr>
<td>D/Dr11</td>
<td>JG13</td>
<td>Psychotic-50</td>
<td></td>
</tr>
<tr>
<td>T(N..F)</td>
<td>JGI7</td>
<td>F02[C-O]</td>
<td></td>
</tr>
<tr>
<td>PCR</td>
<td>JGI8</td>
<td>F03[N-F]</td>
<td></td>
</tr>
<tr>
<td>X0Av</td>
<td>nCrq</td>
<td>F04[N-Cl]</td>
<td></td>
</tr>
<tr>
<td>IC2</td>
<td>nCb-</td>
<td>F05[N-N]</td>
<td></td>
</tr>
<tr>
<td>SIC2</td>
<td>nR=Cs</td>
<td>F05[N-O]</td>
<td></td>
</tr>
<tr>
<td>CIC2</td>
<td>nR#CH/X</td>
<td>F05[N-Cl]</td>
<td></td>
</tr>
<tr>
<td>BIC2</td>
<td>nRCOOR</td>
<td>F05[S-CI]</td>
<td></td>
</tr>
<tr>
<td>MATS1m</td>
<td>nArCOOR</td>
<td>F07[C-F]</td>
<td></td>
</tr>
<tr>
<td>MATS4m</td>
<td>nArOCON</td>
<td>F08[C-N]</td>
<td></td>
</tr>
<tr>
<td>MATS5m</td>
<td>nC(=N)N2</td>
<td>F08[N-S]</td>
<td></td>
</tr>
<tr>
<td>GATS2m</td>
<td>nArNHR</td>
<td>F09[C-N]</td>
<td></td>
</tr>
<tr>
<td>GATS3m</td>
<td>nArNR2</td>
<td>F09[O-O]</td>
<td></td>
</tr>
<tr>
<td>GATS7v</td>
<td>nRCN</td>
<td></td>
<td></td>
</tr>
<tr>
<td>GATS8v</td>
<td>nOHs</td>
<td></td>
<td></td>
</tr>
<tr>
<td>GATS3e</td>
<td>nArOR</td>
<td></td>
<td></td>
</tr>
<tr>
<td>GATS2p</td>
<td>nSO2N</td>
<td></td>
<td></td>
</tr>
<tr>
<td>GATS3p</td>
<td>nPyrroles</td>
<td></td>
<td></td>
</tr>
<tr>
<td>EEig05x</td>
<td>nPyridines</td>
<td></td>
<td></td>
</tr>
<tr>
<td>EEig06x</td>
<td>C-006</td>
<td></td>
<td></td>
</tr>
<tr>
<td>EEig07x</td>
<td>C-013</td>
<td></td>
<td></td>
</tr>
<tr>
<td>EEig09d</td>
<td>C-021</td>
<td></td>
<td></td>
</tr>
<tr>
<td>EEig11d</td>
<td>C-028</td>
<td></td>
<td></td>
</tr>
<tr>
<td>EEig09r</td>
<td>C-033</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Table 2.5. The 79 unique descriptors selected by all 5 models for the 5-HT$_{2B}$ dataset.
2.8.5 5-HT\textsubscript{6} Modeling Result

In total, 1,233 2D Dragon descriptors were generated for these 176 compounds. The classification results of the five-fold external cross validation were combined into a single set to calculate the sensitivity, specificity, and CCR (shown in Figure 2.14). The classification results of the three methods revealed similar performance in classifying the binders vs. non-binders. DWD-VS was able to predict one additional binder and 4 non-binders more than the other two methods.

![Combined Result of Test Sets (5-HT\textsubscript{6} Dataset)](image)

**Figure 2.14.** Performance comparison through five-fold external cross validation for the 5-HT\textsubscript{6} dataset.

For the 5-HT\textsubscript{6} dataset, DWD with variable selection showed better classification performance than the other two methods.
Analyzing the models obtained from DWD-VS indicated that the numbers of descriptors incorporated in each of the five folds were 400, 300, 250, 200, and 250. Cross-checking these descriptors yielded 505 unique descriptor names. Among these unique descriptors, 87 were selected by all the 5 models. These 87 descriptors are shown in Table 2.2.

<table>
<thead>
<tr>
<th>Descriptor Name</th>
<th>Name</th>
<th>Value</th>
<th>Unit</th>
<th>Code</th>
<th>Code</th>
<th>Code</th>
</tr>
</thead>
<tbody>
<tr>
<td>ARR</td>
<td>nNq</td>
<td>Inflammat-50</td>
<td></td>
<td>B09[O-O]</td>
<td></td>
<td></td>
</tr>
<tr>
<td>nDB</td>
<td>nArOH</td>
<td>Depressant-50</td>
<td></td>
<td>B10[C-C]</td>
<td></td>
<td></td>
</tr>
<tr>
<td>nAB</td>
<td>nPyrroles</td>
<td>Neoplastic-80</td>
<td></td>
<td>B10[C-O]</td>
<td></td>
<td></td>
</tr>
<tr>
<td>nBnz</td>
<td>C-001</td>
<td>Neoplastic-50</td>
<td>F01[C-N]</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>D/DTr09</td>
<td>C-004</td>
<td>Infective-80</td>
<td>F02[C-S]</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>MATS5m</td>
<td>C-005</td>
<td>BLTF96</td>
<td>F03[N-S]</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>MATS1v</td>
<td>C-006</td>
<td>BLTD48</td>
<td>F04[C-S]</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>GATS7v</td>
<td>C-008</td>
<td>BLTA96</td>
<td>F06[C-N]</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>GATS5e</td>
<td>C-013</td>
<td>B02[N-O]</td>
<td>F06[N-Cl]</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>GATS7e</td>
<td>C-025</td>
<td>B03[N-N]</td>
<td>F07[C-N]</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>GATS8p</td>
<td>C-027</td>
<td>B03[N-O]</td>
<td>F07[N-S]</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>EEig03d</td>
<td>C-031</td>
<td>B03[N-S]</td>
<td>F08[C-Cl]</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>JGI9</td>
<td>C-034</td>
<td>B04[N-S]</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>JGI10</td>
<td>C-040</td>
<td>B04[O-O]</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>nCp</td>
<td>H-046</td>
<td>B05[N-O]</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>nCs</td>
<td>H-047</td>
<td>B06[C-N]</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>nCq</td>
<td>H-049</td>
<td>B06[N-Cl]</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>nCrs</td>
<td>O-057</td>
<td>B07[C-S]</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>nCar</td>
<td>O-058</td>
<td>B07[N-S]</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>nCbr</td>
<td>N-073</td>
<td>B07[O-O]</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>nRCOnHR</td>
<td>N-079</td>
<td>B08[C-N]</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>nRCOnR2</td>
<td>Ui</td>
<td>B08[C-O]</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>nN=C-N&lt;</td>
<td>MLOGP</td>
<td>B08[N-N]</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>nArNR2</td>
<td>ALOGP2</td>
<td>B08[O-O]</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>nN+</td>
<td>GVWAI-80</td>
<td>B09[C-F]</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Table 2.6. The 87 unique descriptors selected by all 5 models for the 5-HT\textsubscript{6} dataset.
2.9 Conclusion

The results obtained on simulated data clearly indicated that DWD with variable selection (DWD-VS) could significantly improve the model prediction performance for datasets that are imbalanced and HDLSS. DWD-VS consistently achieved the highest prediction performance, and both DWD methods showed better classification performance than SVM in predicting the test set data. Analysis of DWD-VS models indicated that the high prediction accuracy was consistently achieved by nearly 100% noise removal while retaining the majority of the informative descriptors. In the modeling of real QSAR datasets, DWD-VS was consistently better than SVM and had superior or similar performance to DWD without variable selection.

Analysis of descriptors incorporated in DWD-VS models suggested that the selected descriptors could explain the contribution of a molecular structure feature to the desired target biological property. This is especially evident in the modeling result of the AmpC β-lactamase dataset, where the three descriptors, nArCOOH, nSO2N, and n Thiophenes, reflect common structure features of the inhibitors. Thus, DWD-VS can be used to obtain a better understanding of the underlying process that generated the data. Compared to DWD without variable selection, DWD-VS provided more cost-effective models by identifying predictor variables while achieving high prediction accuracy, i.e. identifying a smaller number of descriptors that are significant to a biological property.
Chapter 3

Novel Three-Dimensional Chirality Atom-Pair Descriptors

3.1 Motivation / Background

Many biologically-active compounds are in fact chiral, and their stereochemistries are believed to directly influence their bioactivities because these compounds (ligands) are recognized differently by their corresponding receptors which are also chiral. A molecule is said to be chiral when it cannot be superimposed on its mirror image.

Chirality can be defined either by the molecular optical activity or by the configuration of the molecule. When optical activity is considered, (+) or (-) notations are used to distinguish the chiral compounds from their mirror images. D/L or R/S notations can alternatively be used to distinguish the enantiomers if chirality of the molecule is defined by its molecular configuration instead (illustrated in Figure 3.1). The assignment of the D/L system is based on the association between chiral molecules and glyceraldehydes. As for the R/S system, it is determined by the Cahn-Ingold-Prelog priority rules. It is interesting to point out

![Figure 3.1](http://en.wikipedia.org/wiki/File:Chirality_with_hands.jpg)

**Figure 3.1.** Two enantiomers of a generic amino acid.

Pairs of enantiomers are often designated as “right-” or “left-handed.” Multiple notations can be used to differentiate one enantiomer from the other.

that there is no fixed relation to the three labeling system. For example, an R isomer can be either dextrorotatory or levorotatory, depending on its exact substituents.

Before the 1980s, the pharmacopoeia was dominated by compounds in the form of racemic mixtures, but a breakthrough in technology enabling pure enantiomers to be generated in significant quantities has not only revolutionized the pharmaceutical industry but also has raised the awareness of and interest in the stereochemistry of drug action. It is frequently the case that one enantiomer is responsible for the activity of interest while its paired enantiomer is found to be inactive but does share some other desirable or undesirable activities of interest.

Benefits such as reduction of the total administered dose, enhancement of the therapeutic window, and a more precise estimation of dose–response relationships have been identified as the advantages of using stereochemically pure drugs. As a result, single enantiomers are preferred by both industry and regulatory authorities, such as the US Food and Drug Administration (FDA). In 1992, the FDA published a formal guideline regarding the development of chiral drugs in a document entitled *Development of New Stereoisomeric Drugs*. As stated in the document, “The stereoisomeric composition of a drug with a chiral center should be known and

![Growing Trend of Chiral Technology](image)

**Figure 3.2** Growing trend of chiral technology.

According to a study by Frost & Sullivan, worldwide revenues related to chiral technology, which amounted to $4.8 billion in 1999, were expected to be $14.9 billion in 2009.
the quantitative isomeric composition of the material used in pharmacologic, toxicologic, and clinical studies known. Specifications for the final product should assure identity, strength, quality, and purity from a stereochemical viewpoint.”

The growing trend for worldwide sales of chiral drugs in single-enantiomer forms is expected to continue. In 1996, the worldwide annual sales of chiral drugs amounted to $74.4 billion, which constituted more than one-fourth of all drug sales; the number exceeded $159 billion in 2002 and the sales figure was projected to reach $200 billion in 2008.

<table>
<thead>
<tr>
<th>Brand Name</th>
<th>Generic Name</th>
<th>Therapeutic Area</th>
<th>Pharmaceutical Company</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lipitor</td>
<td>Atorvastatin calcium</td>
<td>Cardiovascular</td>
<td>Pfizer</td>
</tr>
<tr>
<td>Zocor</td>
<td>Simvastatin</td>
<td>Cardiovascular</td>
<td>Merck</td>
</tr>
<tr>
<td>Pravachol</td>
<td>Pravastatin sodium</td>
<td>Cardiovascular</td>
<td>Bristol-Myers Squibb</td>
</tr>
<tr>
<td>Paxil</td>
<td>Paroxetine hydrochloride</td>
<td>Central Nervous System</td>
<td>GlaxoSmithKline</td>
</tr>
<tr>
<td>Plavix</td>
<td>Clopidogrel bisulfate</td>
<td>Hematology</td>
<td>Sanofi-Synthelabo / Bristol-Myers Squibb</td>
</tr>
<tr>
<td>Zoloft</td>
<td>Sertraline hydrochloride</td>
<td>Central Nervous System</td>
<td>Pfizer</td>
</tr>
<tr>
<td>Advair HFA</td>
<td>Fluticasone propionate and salmeterol xinafoate</td>
<td>Respiratory</td>
<td>GlaxoSmithKline</td>
</tr>
<tr>
<td>Nexium</td>
<td>Esomeprazole magnesium</td>
<td>Gastrointestinal</td>
<td>AstraZeneca</td>
</tr>
<tr>
<td>Augmentin</td>
<td>Amoxicillin and potassium clavulanate</td>
<td>Antibiotic</td>
<td>GlaxoSmithKline</td>
</tr>
<tr>
<td>Diovan</td>
<td>Valsartan</td>
<td>Cardiovascular</td>
<td>Novartis</td>
</tr>
</tbody>
</table>

**Table 3. 1.** List of top ten single enantiomer blocker drugs in ranking order.

Drugs must exceed the amount of $1 billion dollars in annual sales to be qualified as blockbuster drugs. The global combined sales of Lipitor and Zocor almost reached 14 billion in 2002.
According to a study by Frost & Sullivan in 2001, worldwide revenues related to chiral technology (equipment), which amounted to $4.8 billion in 1999, is expected to reach $14.9 billion in 2009 (Figure 3.2). Table 3.1 lists the top ten single enantiomer blockbuster drugs that achieve more than $1 billion in sales per year.

3.2 Overview of Current Chirality Descriptors

As summarized by Crippen, current chirality descriptors applied in the field of QSAR can be classified as either qualitative or quantitative descriptors. To characterize chirality quantitatively, three different computational implementations have been applied. The first implementation is to calculate the overlap of the Van Der Waals volume or electrostatic potential by superimposing a pair of enantiomers. Another quantitative implementation is achieved by measuring the degree of distortion, which can be characterized by calculating the distance required to convert the molecule or subsets of its atoms into a structure with a desired symmetry, such as mirror symmetry. The third implementation to quantitatively characterize chirality is achieved by translating and rotating the molecule to a standard position associated with symmetry axes that are based on various atomic properties.

Qualitative chirality descriptors, on the other hand, are typically based on the mathematical concept of oriented volume and with the notational viewpoint of the Cahn-Ingold-Prelog rules. Instead of focus on atomic number, qualitative chirality descriptors could place emphasis on other properties, such as electronegativity, polarizability, resonance stabilization, etc.

The idea of characterizing chirality has sparked a lot of imaginative work by many scientists over the years. Both quantitative and qualitative chirality descriptors have been shown to build successful models in QSAR studies. However, as suggested by Crippen, it has not yet been determined how to best apply these descriptors in QSAR applications.
3.2.1 Prior Implementation of the Chirality Descriptors

Several series of chirality descriptors of chemical organic molecules have been developed earlier in this laboratory \(^{60,61,76}\). These descriptors have been used to build predictive quantitative structure-activity relationship (QSAR) models for several datasets \(^{60-62,74-76}\) and were developed on the basis of conventional topological descriptors of molecular graphs. Chirality descriptors emphasized on electronegativity instead of atomic number were particularly intriguing due to the fact that receptor-ligand interaction is affected by factors such as Van Der Waals interactions, hydrophobic interactions, and electrostatic interactions.

The concept of chirality based on partial charges, which capture the electrostatic interactions at the atomic level, was introduced by Kovatcheva, et al. to study a set of chiral ambergris fragrance compounds \(^{75}\). Similar to the traditional R/S notation, which follows the Cahn-Ingold-Prelog priority rules, the configuration of the molecule was rearranged in such a way that the neighboring atom with the lowest partial-charge was projected toward the back of the plane formed by the remaining 3 neighboring atoms. The three neighboring atoms within the plane can either be in a clockwise or counter-clockwise rotation if moving from the atom with the highest partial charge among the three to the lowest one. The chiral atom would be labeled as \(R^*\) for clockwise rotation and \(S^*\) for counter-clockwise. In a previous study, the

<table>
<thead>
<tr>
<th>Atom Types</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>01</td>
<td>Negative Point Center</td>
</tr>
<tr>
<td>02</td>
<td>Positive Point Center</td>
</tr>
<tr>
<td>03</td>
<td>Hydrogen Bond Acceptor</td>
</tr>
<tr>
<td>04</td>
<td>Hydrogen Bond Donor</td>
</tr>
<tr>
<td>05</td>
<td>Aromatic Ring Center</td>
</tr>
<tr>
<td>06</td>
<td>Nitrogen (All)</td>
</tr>
<tr>
<td>07</td>
<td>Oxygen (All)</td>
</tr>
<tr>
<td>08</td>
<td>Sulfur (All)</td>
</tr>
<tr>
<td>09</td>
<td>Phosphor (All)</td>
</tr>
<tr>
<td>10</td>
<td>Fluorine</td>
</tr>
<tr>
<td>11</td>
<td>Chlorine, Bromine, Iodine</td>
</tr>
<tr>
<td>12</td>
<td>Carbon</td>
</tr>
<tr>
<td>13</td>
<td>Other Elements</td>
</tr>
<tr>
<td>14</td>
<td>Triple Bond Center</td>
</tr>
<tr>
<td>15</td>
<td>Double Bond Center</td>
</tr>
<tr>
<td>16</td>
<td>Chiral Atom R</td>
</tr>
<tr>
<td>17</td>
<td>Chiral Atom S</td>
</tr>
</tbody>
</table>

Table 3.2. The 17 atom types defined in the atom pair descriptors.

Regular atom pair descriptors contain atom types 01 to 15 while chirality atom pair descriptors contained all the atom types in regular atom pair descriptors (with two additional atom types - 16 and 17).
method of choice to calculate partial change in order to define atomic chirality was Gasteiger-Hückel. Additionally, minimum graph distance, which is derived from a 2D molecular structure, was used to define molecular features.

Chiral atom types defined above were incorporated as new atom types for calculation of atom-pair descriptors, based on an approach proposed by Carhart et al. Atom-pair descriptors encoded the molecular graph patterns defined by atom types and topological distance bins. These contained the number of occurrences for each particular pattern. A molecular pattern in atom-pair descriptors was a substructure path separation (or graph distance) between the atoms:

\[ <\text{Atom type I}> <\text{Atom type II}> _{<\text{distance between atom types}>} \]

The distance between any two atom types was defined as the minimum graph distance. The minimum graph distance is the smallest amount of bond distance along the path between any two specified atom types within a molecular structure (2D distance). The value encoded within the atom-pair descriptors contained the number of occurrences associated with a particular pattern as defined by a pair of atom types within a certain distance (binned distance) away. Listing of all 17 atom types implemented in the atom-pair descriptors was shown in Table 3.2.

### 3.2.2 Three Dimensional Chiral Atom-Pair Descriptors

A shortcoming associated with the prior implementation of the atom-pair descriptors was related to the minimum graph distance (2D distance). The molecular patterns defined by the minimum graph distance could not capture the difference between the trans- and cis-configurations of molecules that could have dissimilar biological property. By not capturing the subtle structural differences in the molecular configurations, the descriptors became limited in the QSAR studies.

To address this shortcoming, the distance bin that defined the molecular patterns was changed from 2D distance to three-dimensional (3D) distance, which was calculated by taking the Euclidean distance of the atom coordinates in the physical space (3D space). However, a
limitation associated with 3D molecular descriptors (a set of features describing molecules in 3D) is the fact that the method is conformation dependent. Depending on the structural rigidity of a given molecule, multiple different 3D representations (conformations) can be generated. In order to get a single structural representation for each compound, one common approach is to use the minimum energy conformer, which may not accurately capture the binding conformation of the ligand, especially when the given compounds are very flexible. In fact, there is no a priori reason to exclude higher energy conformers as a source of activity.

Another alternative to the 3D descriptors generated from a single conformation would be deriving the 3D descriptors from multiple conformers of a molecule, which requires a conformation search of all the molecules. A conformation search was achieved through the OMEGA \(^{100}\) software by OpenEye. The search criteria implemented in this study included the generation of a maximum of 50,000 possible conformations for each molecule. For molecules containing less than or equal to 4 rotors, the associated conformers were considered duplicated if the minimum Root Mean Square (RMS) Cartesian distance was less than 0.8. The RMS cutoff value was increased to 1.15 for molecules containing more than 4 rotors. A maximum of 1,000 conformations satisfying the selection criteria were retained. However, the number of resulting conformers could be less than 1,000 if a compound was structurally rigid.

After the completion of a conformation search, 3D chiral atom-pair descriptors were generated for each conformer. Since multiple conformers were associated with a single molecule, each molecule would be characterized by a molecular pattern matrix, where the rows represented the conformers and the molecular patterns associated with each conformer were represented by the columns. To summarize the molecular pattern matrix associated with each molecular entry, taking the Boltzmann average of the molecular pattern matrix could derive a single 3D descriptors vector for a given molecule \(^{80}\). However, using the Boltzmann average placed higher emphasis on the conformers with much lower energy.
Rather than using the Boltzmann average to derive the final 3D descriptor vector for a given molecule, three different values, which were the maximum, the arithmetic mean, and the percentage of zero values (the percentage of conformers that lack the particular feature), were used to characterize the distribution of each descriptor value with the given number of conformers. The choice of an arithmetic mean over the Boltzmann average meant that the possibilities for all the resulting conformers to be in the bioactive form were all equally likely.

In addition to the change of the distance type that defined the molecular patterns, the 3D chiral atom-pair descriptor matrix incorporated the concept of the degree of chirality, which is a threshold used to define the chiral atom types. In order for a carbon atom to be considered as a chiral atom, the minimum difference in the atomic partial charge between any two of the four connecting atoms must be greater than this threshold. By varying this threshold, multiple 3D atom-pair descriptor matrices with different degree of chirality could be generated but only molecular patterns involving atom type 12, 16, and 17 were affected by the threshold. By incorporating the degree of chirality concept, it is possible to identify a subset of chiral carbon atoms and their association with the target properties.

In the earlier study, a carbon atom would be considered as a chiral atom type if all four of its connecting atoms were different and the method of choice to calculate partial charge was Gasteiger-Hückel. The partial charge calculation in this study was accomplished by the QUACpac software from OpenEye, which suggested AM1-BCC to be the model of choice for calculating partial charge due to the better performance on predicting protein-ligand binding calculations and virtual screening through docking methods. The calculation scheme for AM1-BCC was to calculate the initial partial charges derived from the AM1 semi-empirical method, followed by bond-charge corrections (BCC) to generate the final atomic partial charges.

To avoid incorporating the non-chiral atoms as defined by the International Union of Pure and Applied Chemistry into the chiral atom types in the 3D chiral atom-pair descriptors, the minimum threshold to define chiral atoms was set to 0.0010. Additional thresholds were
evaluated by setting the maximum value to 0.0290 with a step size of 0.0020. Thus a total of 15 different thresholds were evaluated. With higher threshold value, the number of chiral atoms in the new definition will be smaller.

3.3 Dataset Description

3.3.1 Peptide Transporter 1 (PEPT1) Dataset

Peptide transporter 1 (PEPT1), localized to the brush border membrane of the intestinal epithelium, is a solute carrier for oligopeptides and transports nutritional di- and triptides across the luminal membrane into small intestinal cells. In addition to the oligopeptides, other peptidomimetics, which are molecules designed to mimic peptides, are also transported by PEPT1. The ability of PEPT1 to transport peptidomimetic drugs, such as β-lactam antibiotics, valacyclovir, δ-amino-levulinic acid, angiotensin-converting enzyme inhibitors, and bestatin, allows the oral application of these drugs for the therapy of several diseases.

Targeting the membrane transport protein PEPT1 to enhance the oral bioavailability of drugs is a promising strategy; however, very little is known about the substrate binding pocket of PEPT1. To gain some insights regarding the structural requirements for the PEPT1 binding substrates, cheminformatic analysis, such as QSAR, can be applied to identify key molecular structure features that are contributing to the binding of PEPT1.

A dataset published by Biegel et al., which contained 122 compounds (substrates) and their corresponding binding affinity values (K<sub>i</sub> values) to PEPT1, was retrieved from the ChEMBL database. Among the 122 PEPT1 binding substrates, there were 31 β–lactam antibiotics, 32 tripeptides, and 59 dipeptides. For tripeptides and dipeptides, different amino acid sequences were measured experimentally for their binding affinity. The experimentally measured K<sub>i</sub> values also included polypeptides (tripeptides and dipeptides) with amino acids that were strategically substituted with D-enantiomers. Tripeptides in LLL configuration display a higher affinity to PEPT1 (with K<sub>i</sub> values ranging from 0.1 to 0.5 mM) compared to tripeptides in DLL, LDL, and
LLD configurations. This stereochemical preference is also observed in dipeptides, which preferred LL more than DL, LD, or DD. However the binding affinity value for the tripeptide, D-Met-Met-Met, also indicated high affinity to PEPT1 (with $K_i$ value of 0.52 mM). In contrast, the LLL configuration of the tripeptides achieved the $K_i$ value of 0.10 mM.

The $K_i$ values for the 122 compounds ranged from 0.01 mM to values greater than 30 mM. The locations of the $K_i$ values were used to determine the labels of the corresponding compounds, which can be labeled as either high affinity or low affinity groups. Based on the 122 logarithmic transformed $K_i$ values, Gaussian kernel density estimation was constructed to estimate the distribution of the experimental binding affinity values (Figure 3.3). According to the constructed density estimation, the tripeptides, D-Met-Met-Met, are closer to the high affinity group than to the low affinity group. By incorporating D-Met-Met-Met into the high affinity group, the upper bound of $K_i$ values for the high affinity group would increase to 0.52 mM. As for the low affinity group, the new lower bound was shifted to 0.86 mM. By taking the average of these two boundary values, a hypothetical threshold of 0.69 was proposed to

![Figure 3.3](image-url) Activity Distribution of the 122 PEPT1 Binding Substrates

The plot shows the distribution of the $\log_{10}(K_i)$ values for the 122 compounds (black circles). The vertical positions of the black circles were based on the entry order, which was sorted in an ascending order according to the $K_i$ values. The $K_i$ values for the 122 compounds ranged from 0.01 mM to values greater than 30 mM. Compounds with $K_i$ value less than 0.69 (green vertical line) are considered as high affinity. Low affinity compounds have $K_i$ value greater than 0.69.
determine the labels of compounds. Compounds with $K_i$ value less than 0.69 mM were considered as high affinity. Low affinity compounds had $K_i$ value greater than 0.69 mM. As a result, the dataset contained 52 and 70 compounds in the high affinity and low affinity group respectively.

To ensure the proper stereochemistry of the retrieved molecules is represented as in the original literature, visual inspection was performed by comparing the retrieved structures with the data available in both the original literature and SciFinder. The resulting comparison indicated that there were 28 structure entries obtained from the ChEMBL database with either incorrect structures or misannotated stereochemistries.

3.3.2 AmpC β–lactamases Dataset (149 Compounds)

As previously described in Section 2.7.1, a dataset containing AmpC β-lactamases inhibitors and non-inhibitors was obtained from Shoichet’s group [17]. In addition to the 110 compounds previously modeled, 39 more compounds with a different scaffold were added. The new scaffold resembles the isoindole (Figure 3.4) and is structurally different from the sulfonamide scaffold.

As with the compounds with the sulfonamide scaffold, the 39 additional compounds were experimentally tested with the same assay to obtain the $K_i$ values. The same $K_i$ threshold criteria applied to the previous data for determining inhibitors and non-inhibitors was also applied to the 39 additional molecules. Molecules with $K_i$ less than 1,000 µM were considered as inhibitors whereas molecules with $K_i$ greater or equal to

![Figure 3.4. Isoindole-like scaffold (top) vs. isoindole (bottom).](image)

The 39 additional compounds have a scaffold resembling isoindole, which is structurally different from the sulfonamide scaffold.
1,000 µM were considered as non-inhibitors. With these 39 additional molecules, the dataset now contained 64 inhibitors and 85 non-inhibitors.

Given the criteria to determine inhibitors and non-inhibitors, it was observed that stereochemistry did have some effect on the 39 additional molecules. There were two pairs of enantiomers observed. The first pair of enantiomers, with both considered as inhibitors, involved entries 111 and 112. Entries 134 and 135 were the other pair of enantiomers, with entry 135 as an inhibitor and entry 134 as a non-inhibitor. The chemical structures of these four molecules were shown in Figure 3.5.

![Figure 3.5](image-url)  
**Figure 3.5.** The two enantiomeric pairs observed in the additional AmpC dataset.

There were two enantiomeric pairs observed in the 39 additional compounds. The first pair of enantiomers, which were both inhibitors, involved entries 111 and 112. Entries 134 and 135 were the other enantiomeric pair with different labels. Entry 134 was considered as a non-inhibitor while entry 135 was an inhibitor.
3.3.3 Artemisinin Dataset

Artemisinin and its derivatives have become the focus of anti-malarial treatment in recent years due to their effectiveness in treating multi-drug resistant P. falciparum and their excellent safety records. However their mechanism of action is not well understood. The presence of a peroxide bridge (Figure 3.14) in artemisinin and its analogues is hypothesized to be an essential feature for its ability to form a bond with a high valence non-heme iron molecule, resulting in a generation of free radicals. These free radicals cause lethal damage to the parasites. However, there is another interesting feature of artemisinin and its derivatives, an abundance of chiral atoms located near the peroxide bridge.

The dataset of 122 artemisinin analogs was obtained from Avery’s lab. Rather than using the experimentally derived IC50 value (in ng/mL), the relative activity (RA) was associated with each molecule. RA, suggested by Avery et al., was first calculated by taking the ratio of the experimentally derived IC50 values between artemisinin and the analog, followed by the correction ratio of the molecular weight between the analog and artemisinin. This value was proposed to minimize the intraday and interlaboratory measurement variation in the IC50 of artemisinin. Based on logarithmically transformed RA values (Equation 1), the molecules can be categorized into 2 classes: molecules with activities better than artemisinin (positive class) and molecules with activities equal to or less than artemisinin (negative class). This particular classification was implemented because our collaborators are interested only in the molecules that

Figure 3.6. Structure of artemisinin.
The peroxide bridge (highlighted in red) is hypothesized to be the key attribute of the anti-malarial property.
have activities better than artemisinin. As a result, the data contained 71 and 51 entries associated with positive and negative classes, respectively.

\[
\log RA = \log \left( \frac{IC_{50} \text{ of artemisinin}}{IC_{50} \text{ of the analog}} \right) \times \log \left( \frac{MW \text{ of the analog}}{MW \text{ of artemisinin}} \right) \tag{Equation 1}
\]

### 3.4 Evaluation of Descriptor Significance and Generalizability

Two different procedures were performed to evaluate the performance of the 3D chiral atom-pair (cAP) descriptors. The first procedure was to evaluate the significances of different cAP descriptor matrices, which were generated by varying the threshold for chirality. By varying the chirality threshold, a different degree of chirality can be assigned to the corresponding cAP descriptor matrix. This procedure allowed for the identification of a subset of chiral carbon atoms that are more likely to attribute to a given biological target property by optimizing the chirality threshold to achieve the highest model significance. The calculation of model significance described in Section 2.3.1 was applied to the full dataset without data partitioning to identify the optimal chirality threshold. By varying the chirality threshold from 0.001 to 0.029 with an increment of 0.002, a total of fifteen cAP descriptor matrices were generated for each of the three datasets. A regular atom-pair descriptor matrix (APair) without chiral atom types was generated as a control group in order to compare the significance of the chiral atom types.

To evaluate the generalizability of the cAP descriptors, the five-fold external cross-validation procedure described in Section 2.6 was applied to generate five different sets of descriptor matrices for the performance benchmark. The benchmark was designed not only to compare the performance between Dragon, APair, and cAP descriptors but also to evaluate the potential for prediction accuracy by combining Dragon descriptors with chiral atom-pair descriptors. Regular atom-pair descriptors were combined with Dragon descriptors to serve as a control group in order to compare the generalizability of the chiral atom types in predicting
unseen data (data in the external test sets). All data entries in training and test sets remained the same for the five different descriptor matrices.

3.5 Modeling Results

3.5.1 PEPT1 Dataset

Compared to all the fifteen cAP descriptor matrices, the model built with APair achieved a much lower significance. As illustrated in Figure 3.7, all fifteen optimal models (models with the highest z-scores) associated with the corresponding cAP descriptor matrices achieved much

![Figure 3.7. Degrees of chirality and the corresponding significance for the PEPT1 dataset.](image)

The model significance, which is determined by the z-scores, indicated that models built with the chiral atom types (atom types 16 and 17) were more significant than the model built without (APair; black line). The most significant model was achieved with the threshold of 0.007 (cyan).
higher z-score values than the model with APair descriptors. This outcome indicated that the chiral atom types do capture the effect of stereochemistry in this highly chirality sensitive dataset. Additional comparison among the optimal models built with different cAP descriptors indicated that different levels of significance were achieved by varying the chirality threshold (degree of chirality). The observation of lower z-score values associated with thresholds ranging from 0.001 to 0.005 suggested the corresponding descriptor matrices contained some chiral atoms that might be trivial to the target property. By considering those trivial chiral atoms as non-chiral atoms, higher z-score values were obtained from the modified descriptor matrices, which were generated by increasing the chirality threshold. Given the ranges of chirality threshold evaluated in this study, the highest z-score value was achieved by the optimal model obtained from the cAP descriptor matrix with a chirality threshold of 0.007.

After establishing the association between the target property and the degree of chirality, the next goal was to evaluate the generalizability of the 3D chiral atom-pair descriptors through a five-fold external cross-validation procedure. To summarize the classification results for the five-fold external cross-validation procedure, sensitivity, specificity, and CCR were calculated from the single combined set of the five test sets associated with each descriptor matrix. The cAP descriptor matrix being evaluated was generated with a threshold of 0.007. As illustrated in Figure 3.8, the models obtained using cAP descriptors achieved higher prediction accuracy than models obtained from either APair or Dragon descriptors alone. By combining Dragon and cAP descriptors, the resulting optimal models achieved higher sensitivity, specificity, and CCR than the models obtained from the combination of Dragon and APair descriptors. Furthermore, the addition of either APair or cAP descriptors to Dragon descriptors improved the models prediction accuracy more than the Dragon descriptors alone.
The benchmarking results of the five-fold external cross-validation indicated cAP descriptors can be used to build predictive models, especially for a chirality sensitivity dataset. In this PEPT1 dataset, models with cAP descriptors achieved more significance and higher prediction accuracy than models with APair descriptors. Both cAP and APair descriptors encoded valuable information associated with PEPT1 binding that is not captured by 2D descriptors such as the Dragon descriptors.

**Figure 3.8.** Five-fold external cross-validation results of the PEPT1 dataset.

Models built with descriptors containing only 2D molecular structure information (Dragon descriptors) performed worse than models built with descriptors containing 3D molecular structure information. Among the descriptors containing 3D molecular structure information, descriptor matrices containing chiral atom pair descriptors yielded much better performances in classifying the test sets.
3.5.2 AmpC β–lactamase Dataset (149 Compounds)

When compared to the APair descriptors, only the two cAP descriptor matrices generated with the thresholds 0.011 and 0.013 achieved higher values in z-scores during the evaluation of significance. As illustrated in Figure 3.9, most of the optimal models (models with the highest z-score) associated with the corresponding cAP descriptor matrices achieved similar if not lower z-score values than the model with APair descriptors. This outcome matched the fact that this AmpC β-lactamase dataset is fairly insensitive to stereochemistry since there only existed a pair of enantiomers with different property labels. However, the outcome of higher z-scores achieved

![Figure 3.9](image.png)

**Figure 3.9.** Degrees of chirality and the corresponding significance for AmpC β-lactamase dataset (149 compounds).

The semi-log plot indicated that only the chiral atom pair descriptor matrices generated with a threshold of 0.011 and 0.013 were more significant than the regular atom pair descriptor matrix (APair).
by setting the chirality threshold to 0.011 and 0.013 suggested that the targeted biological property could be influenced by a subset of chiral atoms within the molecular entries. These potentially influential chiral atoms and their associated 3D molecular patterns became less distinctive as the chirality threshold further increased, thus resulting in a drop in z-score values.

A five-fold external cross-validation procedure was applied to evaluate the generalizability of the cAP descriptors. To summarize the classification results for the five-fold external cross-validation procedure, sensitivity, specificity, and CCR were calculated from the single combined set of the five test sets associated with each descriptor matrix. The cAP

![Classification Performance](image)

**Figure 3.10.** Five-fold external cross-validation result of AmpC β-lactamase dataset (149 compounds).

For this AmpC β-lactamase dataset, the models built with chiral atom pair descriptors achieved better specificity than models built with other descriptors. The outcome was interesting, especially when comparing the model performance between descriptor matrices containing chiral atom pair descriptors only and the combined descriptor matrix containing both Dragon and chiral atom pair descriptors.
descriptor matrix being evaluated was generated with a threshold of 0.011. As illustrated in Figure 3.10, the models obtained using cAP descriptors achieved higher prediction accuracy (higher specificity) than models obtained from either APair or Dragon descriptors alone. However, the difference in performance only existed when predicting the non-inhibitors. The combination of Dragon descriptors with either APair or cAP descriptors showed minor performance gain in specificity more than the Dragon descriptors alone. However, combining Dragon and cAP descriptors achieved similar if not equivalent results to the combination of Dragon with APair descriptors.

The benchmarking results of the five-fold external cross-validation indicated cAP descriptors can be used to build predictive models that are comparable to the models built with Dragon descriptors. For this AmpC β-lactamase dataset, which is relative insensitive to stereochemistry, models built with cAP descriptors achieved higher prediction accuracy than models with other descriptors. Given the higher prediction accuracy associated with models built with cAP and APair descriptors, there could be molecular patterns encoded in these descriptor matrices that might not be captured by 2D descriptors such as the Dragon descriptors.

### 3.5.3 Artemisinin Dataset

Similar to the PEPT1 dataset, the model built with APair achieved a much lower significance when comparing all fifteen cAP descriptor matrices. As illustrated in Figure 3.11, all fifteen optimal models (models with the highest z-scores) associated with the corresponding cAP descriptor matrices achieved much higher z-score values than the model with APair descriptors. This outcome indicated that the chiral atom types do capture the effect of stereochemistry in this chirality sensitive dataset. Additional comparison among the optimal models built with different cAP descriptors indicated that different levels of significance were achieved by varying the chirality threshold (degree of chirality). Given the ranges of chirality threshold evaluated in this
study, the highest z-score value was achieved by the optimal model obtained from the cAP descriptor matrix with a chirality threshold of 0.015.

![Artemisinin: Degrees of Chirality and Corresponding Significance](image)

**Figure 3.11.** Degrees of chirality and corresponding significance for the artemisinin dataset.

The model significance, which is determined by the z-scores, indicated that models built with the chiral atom types (atom types 16 and 17) were more significant than the model built without (APair; black line). The most significant model was achieved with the threshold of 0.015(red).

Five-fold external cross-validation was applied to evaluating the generalizability of the 3D chiral atom-pair descriptors. To summarize the classification results from the five-fold external cross-validation procedure, average values for sensitivity, specificity, and CCR were reported for each descriptor matrix. The cAP descriptor matrix being evaluated was generated with a threshold of 0.015. As illustrated in Figure 3.12, the models obtained using cAP
descriptors achieved higher CCR than models obtained from either APair or Dragon descriptors alone. By combining Dragon and cAP descriptors, the resulting optimal models achieved higher specificity than the models obtained from the combination of Dragon and APair descriptors. With the addition of either APair or cAP descriptors combined with Dragon descriptors, the prediction accuracy of the models improved compared to models obtained from Dragon descriptors alone.

The benchmarking results of the five-fold external cross-validation indicated that cAP descriptors can be used to build predictive models for datasets containing chiral centers but not necessarily including enantiomeric pairs. In this artemisinin dataset, models with cAP descriptors
achieved more significance and higher prediction accuracy than models with APair descriptors. Both cAP and APair descriptors encoded valuable information associated with antimalarial property that is not captured by the 2D descriptors, such as the Dragon descriptors.

3.6 Conclusion

Chirality is an important concept in medicinal chemistry, and many biochemical reactions and processes are stereospecific, such as the recognition of ligands by their corresponding receptors. In order to identify key stereochemistry associated with a given biological property, novel 3D chiral atom-pair descriptors were developed. By varying the chirality threshold, different subsets of chiral atoms and their associated molecular patterns were encoded in a descriptor matrix. Three datasets containing stereochemistry information were selected for benchmarking the developed descriptors. Results indicated that the models based on developed 3D chiral atom-pair descriptors were more predictive than models obtained from non-chiral 3D atom-pair or 2D Dragon descriptors alone. The difference in descriptor performance directly correlated to the amount of stereochemistry information available in the dataset.

Due to the varying amount of chirality data available in the datasets, both 3D chiral and 3D non-chiral atom-pair descriptors were independently compared in combination with 2D Dragon descriptors. The resulting models indicated better prediction accuracy than models obtained from Dragon descriptors alone, suggesting that the 3D atom-pair descriptors used in this study could be complementary to Dragon descriptors. However, lower prediction accuracy associated with the models obtained from both sets of combined descriptors (Dragon + 3D atom-pairs) when compared to models obtained from 3D atom-pair descriptors alone raised two concerns: 1) a problem of simple merge for descriptor matrices and 2) a limitation of DWD with variable selection. Thus, simple merging of descriptor matrices can result in the united descriptor matrix containing highly correlated descriptors, which could affect the descriptor loadings assigned in DWD and decrease the predictivity of the resulting model. To address this problem
and to improve the performance of the variable selection DWD, correlation between descriptors will have to be considered as part of the variable selection procedure. One possible method would be to cluster the descriptors first and evaluate the significance of the descriptors as a group. This solution will likely retain all instances when important descriptors are highly correlated with one another.
4.1 Growth of Protein Structure Data

Proteins, whether from bacteria or the most complex forms of life, are constructed from the same ubiquitous set of twenty amino acids (building blocks). Cells can produce proteins with strikingly different properties and activities by joining these building blocks in many different combinations and varying the length of sequences, creating a great variety of enzymes with specific biological activities. Enzymes are usually very specific to the reactions they catalyze and the substrates involved. Impressive levels of stereo-, region- and chemo-specificity have been demonstrated in enzymes. This specificity is achieved by the complementarity in characteristics such as shape, charge and pattern of hydrophilic/hydrophobic interactions between enzymes and substrates.

Enzymes are generally globular proteins made of long, linear chains of amino acids that fold to three-dimensional structures with their sizes ranging from 62 amino acid residues, e.g., the monomer of 4-oxalocrotonate tautomerase, to over 2,500 residues, e.g., the fatty acid synthase found in animals. Each unique amino acid sequence produces a specific structure, which has unique properties. Individual protein chains may sometimes group together to form a protein complex. Most enzymes are much larger than the substrates they act on, but only a small portion of the enzyme (up to 7 amino acids) is believed to be directly involved in catalysis. This region, which is responsible for binding the substrate and carries out the reaction, is known as the active site. Enzymes can also contain sites (allosteric sites) that bind cofactors (for catalysis) and/or small molecules that directly or indirectly interact with products or substrates of the reaction.
catalyzed. The later can contribute to the activity of an enzyme through feedback regulation. The activities of enzymes are believed to be determined by their three-dimensional structure. Although structure does determine function, predicting a novel enzyme’s activity base purely on its structure information remains a challenging problem.

However, due to recent technology advances in the field of genomics, there are a significant amount of protein structures made available. Figure 4.1 shows the nearly exponential growth in the number of structures added each year to the Protein Data Bank (PDB), a public database for biological macromolecular structures. In 1976, there were only 13 protein structures in the PDB. Starting from 1990, the annual growth of protein structure entries exceeded 3 digits.

Figure 4.1. Annual growth of structures in Protein Data Bank (PDB).

The figure shows the nearly exponential growth of protein structure entries in the PDB. The red line represents the cumulative total structure entries available per year (log10 scale) while the blue bars represent the protein structure entries added in each year.
Beginning with 1993, the pace at which protein structures are being discovered has grown at a much faster rate, with the PDB exceeding 23,000 structures in 2003, and 78,000 in 2011.

With the increase in protein structure entries, the number of protein structures with unknown function also increases. There were only 5 protein structures with unknown function in 2000, but within a decade, that number had quickly grown to 2,422. Figure 4.2 illustrates the number of protein structures obtained by querying with the keyword, “Unknown Function,” grouped by the year that the structure was released. Identifying the function of these protein structures remains a challenging task.

![Annual Growth of Protein Structures with Unknown Function](image)

**Figure 4.2.** Annual growth of protein structures with unknown function.

A recent revision of the PDB allows users easier retrieval of protein structure entries based on a wide variety of information associated with the entries. Possible query types include, but are not limited to, structure features, sequence features, chemical components, biology, and method. Under the biology query types, users can retrieve protein structure entries with similar
functions by querying Enzyme Commission or Classification (E.C.) number, which is a numerical classification scheme for enzymes that is based on the chemical reactions they catalyze. The E.C. number is based on a hierarchical order as denoted by the four numbers separated by periods, with the first number ranging from one to six to represent the six major classes of enzyme: oxidoreductases (E.C. 1), transferases (E.C. 2), hydrolases (E.C. 3), lyases (E.C. 4), isomerases (E.C. 5), and ligases (E.C. 6). As the notations increase in length, a progressively finer classification of the enzyme can be obtained. The first version of the enzyme classification number, which contains 712 enzymes, was published in 1961. The latest version, published in 1992, contains 3,196 different enzymes.

4.2 Overview of Current Methods for Protein Function Annotation

Function-prediction methods existing today are based on the sequence-function and structure-function relationships of proteins. The challenge for protein function annotation is to decipher the connection between the structural or sequence similarities and the actual level of functional relatedness. In this section, we describe the progress in the automated prediction of protein function based on protein sequence and structure. We will first focus on methods that attempt to extract functional information from protein sequences, which generally utilize sequence alignment and clustering, and then discuss methods that use protein structure information.

4.2.1 Sequence-Based Methods

Sequence-based methods annotate protein structures with possible function by either grouping proteins into families or comparing a target sequence with pre-compiled databases of families, which often rely on a sequence similarity search through BLAST, or Basic Local Alignment Search Tool. BLAST calculates similarity of a query sequence against a database of sequences and retrieves sequences that resemble the query sequence above a certain threshold. Sequences retrieved from BLAST are typically close homologues to the query sequence.
To identify distant homologues, Position-Specific Iterative BLAST, or PSI-BLAST $^9$, is utilized. PSI-BLAST first retrieves a list of all closely related proteins and generates a sequence profile based on multiple sequence alignment (MSA). This sequence profile, which summarizes significant features presented in these closely related sequences, is served as query for searching the database. Examples of these sequence profiles can be found at Conserved Domains Database $^{90}$ (CDD), ProDom $^{27}$, and PROSITE $^{68}$. Sequence profiles can also be obtained through hidden Markov models (HMM), which assume the system being modeled is a memoryless stochastic process with unobserved states. Examples of sequence profiles generated from HMM are provided by Pfam $^{54}$, EVEREST $^{106}$, and SMART $^{85}$. All profile-based methods vary significantly in their level of automation, manual curation, and reliance on complementary resources in the annotation of protein functions $^{88}$. By integrating all the different methods, a more comprehensive coverage can be achieved, such as shown in InterPro $^{95}$, an integrated database of protein families that combines at least twelve member databases.

4.2.2 Structure-Based Methods

Observations have been made that evolution retains the protein folding pattern long after sequence similarity becomes undetectable $^{37,143}$. For example, the catalytic triad located in the active site of the enzyme is structurally preserved in all serine protease enzymes. The triad is a coordinated structure consisting of three essential amino acids: histidine (His), aspartic acid (Asp), and serine (Ser). Mapping the triad back to the sequence indicates that the His, Asp, and Ser are located at sequence positions of 57, 102, and 195 for bacterial serine protease. For mammalian serine protease, the positions are 64, 32 and 221 respectively (Figure 4.3). Neither the order nor the length between these three amino acids is conserved at the sequence level $^{103}$. Consequently, a benchmark study showed that structural similarities can be more reliable than sequence similarities for grouping proteins with a common biological function $^{88,131}$.
Structure similarity can be calculated by using global structure comparison, which compares the structure of a queried protein to domains in the structure databases. Global structure-comparison methods, such as DALI ⁶⁵, MSDFold ⁷⁹, VAST ⁸⁹, CE ¹¹⁹, STRUCTAL ⁷₃, and FATCAT ¹⁴⁷, identify structural neighbors in the Protein Data Bank through pairwise structural alignment and differ in alignment methods; however, they do not discriminate between conservation of the overall fold and functionally relevant regions of the protein ⁸⁸. By focusing structure similarity comparison on more localized regions, such as clefts, pockets and surfaces, the identification and comparison of such regions can suggest similarity in protein function, because the ligand-binding pocket or active site is commonly situated in the largest cleft in the protein ⁸³.

Multiple methods have been developed to define these local regions within a protein structure. SURFNET ⁵⁸,⁸² attempts to identify clefts by fitting spheres of a range of sizes between atoms of a given protein. Further enhancement can be made by coupling SURFNET with ConSurf ⁸¹ to isolate the clefts that are close in proximity to the evolutionarily conserved residues which are defined by the ConSurfHSSP database ⁵⁹.

A surface-comparison method that identifies similar surface patterns based on geometrically defined pocket and void surfaces of amino acid residues on proteins is called pvSOAR ²¹,²² (pocket and void Surfaces Of Amino acid Residues). This method first establishes a residue correspondence between surfaces by aligning sub-sequences of surface residues. These residues are then superimposed and the resulting root-mean-square deviation (RMSD) is evaluated for statistical significance. The pvSOAR and CASTp ⁵² databases are the two essential components for the Global Protein Surface Survey (GPSS) ²³, which contains three-dimensional libraries of functionally annotated surfaces from ligand, deoxyribonucleic acid (DNA), metal and peptide binding surfaces.

Other methods such as Catalytic Site Atlas ¹⁰⁵, PDBSiteScan ⁶⁹, PINTS ¹²⁴ (Patterns In Non-homogous Tertiary Structures), and ET ⁸⁷ (Evolutionary Trace) are template-based methods
to identify active-site residues. These methods utilize a variety of template-based scans to identify active sites and putative ligand-binding pockets. The representation of a template varies by method and can be derived either manually or automatically. For Catalytic Site Atlas, the templates can either reflect the backbone orientation (a set of Cα and Cβ atoms) or the orientation of the ends of the residue sidechains (three functional atoms for each sidechain). These templates from Catalytic Site Atlas are derived manually by mining the literature. The templates from PDBScan that contain a set of residues and their corresponding atoms are automatically generated based on the information within the SITE records of PDB files and protein-protein interaction data. PINTS defines its templates by detecting the largest common 3D arrangement of residues between any two structures. The representation of the templates in PINTS can be atoms or points defined by other criteria (e.g., sidechain centroids). As for ET, it uses phylogenetic trees to rank residues in a protein sequence by their evolutionary importance and maps these residues onto the structure.

There are also other pocket-centric methods, such as FEATURE, SiteEngine, and SURF’S UP, that describe protein-ligand interactions and active-site chemistry by the physicochemical properties of the local environments in the pockets and surfaces. For example, FEATURE represents the local microenvironment using various physical and chemical properties ranging from simple atom-based characteristics such as charge to polypeptide-based characteristics such as type of secondary structure. These methods are based on the assumption that protein surface regions with similar physicochemical properties and shapes may perform similar functions and bind similar binding partners.

Integrating multiple resources from both sequence-based and structure-based methods provides a consensus view, which increases the likelihood of accurate predictions of function. Examples of such integrated metaservers include ProFunc and ProKnow. However, most methods described above either rely on global structural alignment or local region comparison (template based). Methods based on global structural alignment find proteins with a similar fold,
but will not be successful when either proteins adopt a new fold (i.e., does not resemble any known structure) or proteins adopt very common folds that perform many different functions. Template-based methods usually perform better in scenarios where methods based on global structural alignment fail; however, the performance of the template-based methods depends on the identification of the active sites.

4.3 Motivation

With the large amount of protein structure information available, identifying the common structure features, or structure motifs associated with a group of enzymes sharing similar functions, has become feasible. Once identified, the presence of these structure motifs, which could be portions of an active site, allosteric site, or structure conserved region, is used to associate (or infer) the protein structures with unknown functions to a enzyme class with known function. Function inference from structure is facilitated by the use of patterns of residues (3D motifs), normally identified by expert knowledge, that correlate with function. As an alternative to often limited expert knowledge, we use statistical learning techniques and novel protein descriptors to automatically identify residues that contribute to protein functions.

4.4 Novel Protein Descriptors

The goal for designing novel protein descriptors is to study the structure-function relationship of proteins through QSAR-like strategies. By combining variable selection DWD with the novel protein descriptors, it is possible to identify important amino acids that are common in proteins with similar functions.

The novel protein descriptors implemented in this research are designed to characterize the three-dimensional protein structures, with emphasis on the local geometric properties of proteins. The rationale is that the 3D arrangement of enzyme active-site residues is often more conserved than the overall fold. For enzymes, or proteins with similar functions, it is not the similarity of their global structures that define their functions but rather the catalytic sites (local
region). Enzymes can have dissimilar structures globally but perform the same catalytic reaction. The goal of this set of descriptors is to capture the information within the local regions of protein structures.

For protein function annotation, multiple studies have successfully identified family motifs by applying Delaunay Tessellation to the protein crystal structures. By representing amino acid residues in protein chains by Cα atoms or side-chain centroids, a protein is represented as a set of points in three-dimensional space. Cα atoms, which are located on a protein backbone, were chosen to represent the amino acid residues in the protein descriptors for their relative stability over the side-chain centroid. Delaunay Tessellation naturally partitions the space occupied by the protein into tetrahedra with Cα atoms or side-chain centroids at their vertices. Implementing Delaunay tessellation on this set of points generated an aggregate of space-filling irregular tetrahedra, or Delaunay simplices. The vertices of each simplex (tetrahedron) define objectively four nearest neighbor Cα atoms, which correspond to four nearest-neighbor residues; thus, these tetrahedra can be categorized by the amino acid residues occupied at their vertices. By combining the different categories of tetrahedra and the corresponding geometric properties, a protein structure can be encoded as a feature vector.

4.4.1 Classification Scheme of Amino Acids (Ten Classes) and Geometric Properties

Each tetrahedron is categorized by the various amino acids occupied at its four vertices, and the number of possible tetrahedral categories, if determined by all the twenty amino acids, is 8,855. This number is tripled when geometric features, such as volume, exposed surface area, and hidden surface area, are incorporated into each tetrahedral category as features to describe protein structures. This rapid increase in dimensions puts constraints on the number of possible geometric properties that can be incorporated into each tetrahedral category. By reducing the twenty amino acids labels into ten classes, the total number of tetrahedral categories can be drastically reduced (a reduction that exceeds tenfold reduction), and Table 4.1 showed the categorization of possible
tetrahedral associated with twenty amino acids and ten amino acid classes, which are 8,855 and 715 respectively. A recent study by Li et al. compared the ability to detect distantly related protein folds with various reduced alphabets of amino acids and suggested that ten classes of amino acids may be the degree of freedom for characterizing the complexity in proteins.

We assigned the vertices of the tetrahedra using a modified Delvin’s amino acid classification (Table 4.2) that classifies twenty amino acids into ten classes.

<table>
<thead>
<tr>
<th>Tetrahedral Categories</th>
<th>Qty 20 amino acids</th>
<th>Qty 10 Classes</th>
</tr>
</thead>
<tbody>
<tr>
<td>AAAA</td>
<td>20</td>
<td>10</td>
</tr>
<tr>
<td>AAAB</td>
<td>380</td>
<td>90</td>
</tr>
<tr>
<td>AABB</td>
<td>190</td>
<td>45</td>
</tr>
<tr>
<td>AABC</td>
<td>3420</td>
<td>360</td>
</tr>
<tr>
<td>ABCD</td>
<td>4845</td>
<td>210</td>
</tr>
<tr>
<td><strong>Total</strong></td>
<td><strong>8855</strong></td>
<td><strong>715</strong></td>
</tr>
</tbody>
</table>

*Table 4.1. Possible categories of tetrahedra.*

The amino acids corresponding to the vertices of each tetrahedron can be classified into 10 classes. By placing the twenty amino acids into ten classes, there is a reduction of the possible tetrahedron types from 8,855 to 715. The A, B, C, and D, which are listed in the tetrahedral categories, represent different labels or classes of amino acids.

After establishing the tetrahedral categories with ten amino acid classes, we calculated various geometric properties, such as volume and two different surface area measures (i.e., hidden and exposed surfaces between tetrahedra), for a set of tetrahedra to describe a protein structure. The volume of a tetrahedron encodes the proximity of the four amino acids at its vertices situated in space. The hidden surface area is the surface area that a tetrahedron shared with its nearest neighboring tetrahedron, and the exposed surface area is the unshared surface area of a tetrahedra. By recording different surface area measures, the location of the tetrahedron with respect to other tetrahedra was encoded in the descriptors. For instance, a tetrahedron surrounded by 4 other tetrahedra would have zero exposed surface area; thus it is buried inside the protein structure.
Our implemented protein descriptors characterize a protein structure with a vector containing 2,145 features, which include the total volume, total hidden surface area, and total exposed surface area for each of the 715 tetrahedral categories. As illustrated in Figure 4.3, when comparing two hypothetical proteins containing the same amount of tetrahedra of the same category but different sizes, one can easily distinguish the two proteins by using the total volume.

<table>
<thead>
<tr>
<th>Class</th>
<th>Full Name</th>
<th>3 Letter</th>
<th>1 Letter</th>
<th>Superstructure</th>
<th>structure</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>Glycine</td>
<td>Gly</td>
<td>G</td>
<td>Monoamino, monocarboxylic</td>
<td>Small</td>
</tr>
<tr>
<td></td>
<td>Alanine</td>
<td>Ala</td>
<td>A</td>
<td></td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>Valine</td>
<td>Val</td>
<td>V</td>
<td>Monoamino, monocarboxylic</td>
<td>Unsubstituted</td>
</tr>
<tr>
<td></td>
<td>Leucine</td>
<td>Leu</td>
<td>L</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Isoleucine</td>
<td>Ile</td>
<td>I</td>
<td></td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>Proline</td>
<td>Pro</td>
<td>P</td>
<td>Monoamino, monocarboxylic</td>
<td>Heterocyclic</td>
</tr>
<tr>
<td>3</td>
<td>Tyrosine</td>
<td>Tyr</td>
<td>Y</td>
<td>Monoamino, monocarboxylic</td>
<td>Aromatic</td>
</tr>
<tr>
<td></td>
<td>Phenylalanine</td>
<td>Phe</td>
<td>F</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Tryptophan</td>
<td>Trp</td>
<td>W</td>
<td></td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>Methionine</td>
<td>Met</td>
<td>M</td>
<td>Monoamino, monocarboxylic</td>
<td>Thioether</td>
</tr>
<tr>
<td>5</td>
<td>Serine</td>
<td>Ser</td>
<td>S</td>
<td>Monoamino, monocarboxylic</td>
<td>Hydroxy</td>
</tr>
<tr>
<td></td>
<td>Threonine</td>
<td>Thr</td>
<td>T</td>
<td></td>
<td></td>
</tr>
<tr>
<td>6</td>
<td>Cysteine</td>
<td>Cys</td>
<td>C</td>
<td>Monoamino, monocarboxylic</td>
<td>Mercapto</td>
</tr>
<tr>
<td>7</td>
<td>Asparagine</td>
<td>Asn</td>
<td>N</td>
<td>Monoamino, monocarboxylic</td>
<td>Carboxamide</td>
</tr>
<tr>
<td></td>
<td>Glutamine</td>
<td>Gln</td>
<td>Q</td>
<td></td>
<td></td>
</tr>
<tr>
<td>8</td>
<td>Aspartate</td>
<td>Asp</td>
<td>D</td>
<td>Monoamino, dicarboxylic</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Glutamate</td>
<td>Glu</td>
<td>E</td>
<td></td>
<td></td>
</tr>
<tr>
<td>9</td>
<td>Lysine</td>
<td>Lys</td>
<td>K</td>
<td>Diamino, monocarboxylic</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Arginine</td>
<td>Arg</td>
<td>R</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Histidine</td>
<td>His</td>
<td>H</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

**Table 4.2.** Classification of amino acids.

The twenty amino acids were partitioned into 10 classes based on Devlin. We split Class 0 and Class 1, which Devlin puts in the same class.
4.4.2 Geometric Property of α–helix vs. β sheet

In proteins, α–helix and β-sheet are the two common forms of secondary structure, or highly regular local sub-structures. To evaluate the ability of different geometric properties in capturing the difference between α–helix and β-sheet, we applied Delaunay Tessellation to Cα atoms of two PDB entries that represent α–helix (PDB ID: 2GOF) and β-sheet (PDB ID: 2JNI). We then removed tetrahedra containing an edge greater or equal to 12 Å in order to focus on the local neighborhood relationships of amino acids within a protein structure. Setting the threshold to 12 Å was empirical, and this threshold value was determined based on tessellating a random set of PDB entries to avoid orphan Cα atoms that were not associated with any tetrahedron.

Visual comparison between the two sets of Delaunay simplices (tetrahedra) suggested that
geometric properties alone (without information regarding the composition of amino acids) can
distinguish α–helix from β sheet. Figure 4.4 indicates that the packing of tetrahedra between α–
helix (PDB ID: 2GOF) and β-sheet (PDB ID: 2JNI) appears different. The tetrahedra in α–helix
visually appear more uniform than those in β sheet.

![Figure 4.4. α–helix vs. β–sheet.](image)

The packing of the tetrahedra in blue and red corresponds to the α–helix (PDB ID: 2GOF)
and the β-sheet (PDB ID: 2JNI) respectively. The tetrahedra in α–helix are more uniform
in size than those in β-sheet.

By comparing three geometric properties of tetrahedra, i.e., volume, hidden surface area,
and exposed surface area, between α–helix and β-sheet, we can make two observations. First, the
tetrahedra associated with α–helix have less variation in both volume and hidden surface area
(shared surface area). The tetrahedra with volume greater than 15 units or with exposed surface
area greater than 60 units were likely to belong to β sheet. This result confirmed the uniformity of
tetrahedra associated with α–helix that were previously observed through visual inspection.

Second, the exposed surface areas for tetrahedra associated with both α–helix and β-sheet
have a similar longtail distribution with zero as the dominate value (bottom plot of Figure 4.5).
This finding suggested that using the exposed surface area of a tetrahedron as a feature by itself is
insufficient to distinguish an α-helix from a β sheet.
Figure 4. 5. Difference in geometric properties of tetrahedra between α-helix and β sheet. Tetrahedra associated with α-helix are blue; those with β-sheet are red. The tetrahedra associated with α-helix are more uniform than those from β-sheet as shown in volume (top) and hidden surface area (middle). It is less distinctive in exposed surface area (below).
4.5 Selection of Enzyme Diversity Set

We selected a subset representing each of the six major types of enzyme as the enzyme diversity set. This enzyme diversity set served as the negative class to contrast with the enzyme class of interest (positive class) in order for the classifier described in Chapter 2 to extract tetrahera unique to the positive class. The selection criteria for PDB entries associated with each major enzyme type were as follows:

- Must be a biological unit
- Must match the enzyme classification number (top level)
- Must have X-ray resolution better than 1.59 Å
- Must have sequence identity less than 50%

**Figure 4.6.** Distribution of the six major enzyme groups in the enzyme diversity set.

There are 1,005 enzymes in the enzyme diversity set, consisting of hydrolases (EC 3) 45%, transferases (EC 2) 21%, and oxidoreductases (EC 1) 18%. Lyases (EC 4), isomerases (EC 5), and ligases (EC 6) made up the remaining data, each less than 10%.
The resulting enzyme diversity set contained 1,005 PDB entries, consisting of hydrolases (EC 3) 45%, Transferases (EC 2) 21%, and oxidoreductases (EC 1) 18%. Lyases (EC 4), isomerases (EC 5), and ligases (EC 6) made up the remaining data, with each less than 10%. Figure 4.6 shows the distribution of enzyme families in the enzyme diversity set. The number of entries in the enzyme diversity set was further reduced by removing entries with identical function as the enzyme class of interest. Thus the size of the enzyme diversity set depends on the enzyme class of interest.

The distribution of the enzyme families within the enzyme diversity set showed the limitation for selecting the enzyme class of interest. With the data size of isomerases and ligases both under 50, it was not feasible to study the enzymes in these two groups with more specific function.

4.6 Selection of Enzyme Homologs

The level of function specificity for a given enzyme class can be determined by its EC number. The length of the EC numbers (up to four digits) is designed to reflect the level of specificity in enzyme function. As the enzyme functions become more specific, their corresponding EC number becomes longer. The selection criteria for the enzyme homologs were similar to that of the enzyme diversity set. However, the sequence identity criterion had to be modified in order to obtain a sufficient number of structure entries that are functional homologs with low sequence similarity. The selection criteria for the enzyme homologs were as follows:

- Must be a biological unit
- Must match exactly the enzyme classification number
- Must have X-ray resolution better than 1.59 Å
- Must have sequence identity less than 70%

The PDB partitions the sequence identity into intervals and the jump from 50% to 70% is the smallest step possible. Six different functional classes of enzyme homologs were selected according to these criteria: superoxide dismutase (EC1.15.1.1), cellulase (EC 3.2.1.4), β-
lactamase (EC 3.5.2.6), methyltransferase (EC 2.1.1), protein-tyrosine kinase (EC 2.7.10), and protein-serine/threonine kinases (EC 2.7.11).

These six functional classes of enzyme homologs were selected because the numbers of entries associated with them are sufficient after applying the selection criteria. More than ten PDB entries, which are functional homologs, were retrieved for all of the six enzyme classes. These six enzyme classes could be grouped into two types: enzymes classes with more specific function or enzymes classes with more general functions. The goal to include enzyme classes with two different levels of function specificity was to evaluate the capability of applying the protein descriptors in order to study enzyme classes with more generalized function.

4.6.1 Superoxide Dismutase (EC 1.15.1.1)

EC 1.15.1.1 is the enzyme classification number for superoxide dismutases (SOD). The enzymes that belong to this class are responsible to catalyze the dismutation of superoxide into oxygen and hydrogen peroxide. They serve as an important antioxidant defense in nearly all cells exposed to oxygen. Three major families of superoxide dismutase are classified based on the binding metal cofactor: SODs that bind to nickel, SODs that bind to either iron or manganese, and SODs that bind to copper/zinc.

Abnormalities in the copper- and zinc-
dependent superoxide dismutase gene may contribute to the development of Amyotrophic Lateral Sclerosis, a fatal disease that causes deterioration of motor nerve cells in the brain and spinal cord. Table 4.3 shows the fourteen PDB entries associated with SOD that met the selection criteria.

4.6.2 Cellulase (EC 3.2.1.4)

EC 3.2.1.4 is the enzyme classification number for cellulase, which is a class of enzymes produced by fungi, bacteria, and protozoans to catalyze cellulolysis. Other types of organisms, including some plants and animals, also produce cellulases. Several different kinds of cellulases are known, which differ structurally and mechanistically, and their applications exist in both commercial and pharmaceutical settings. Commercial applications of cellulases include laundry detergents; food processing, e.g., hydrolysis of cellulose during drying of coffee beans; and renewable energy, e.g., hydrolysis of cellulose in biomass to glucose in the fermentation stage.

Since cellulase is a digestive enzyme and an anti-cholinergic agent, it is used to help digest protein, starch, and fat. As a result, there are also pharmaceutical applications associated with cellulases, including treatments for bowel spasms and Phytobezoars, a form of cellulose bezoar found in the human stomach. Table 4.4 shows the eighteen retrieved PDB entries associated with cellulase based on the selection criteria.

<table>
<thead>
<tr>
<th>PDB Entries</th>
<th>Types</th>
</tr>
</thead>
<tbody>
<tr>
<td>1H1N</td>
<td>ENDO TYPE CELLULASE ENGI</td>
</tr>
<tr>
<td>1KS8</td>
<td>ENDO-BETA-1,4-GLUCANASE</td>
</tr>
<tr>
<td>1KWF</td>
<td>ENDOGLUCANASE A</td>
</tr>
<tr>
<td>1OA2</td>
<td>ENDO-BETA-1,4-GLUCANASE</td>
</tr>
<tr>
<td>1OJJ</td>
<td>ENDOGLUCANASE I</td>
</tr>
<tr>
<td>1OLR</td>
<td>ENDO-BETA-1,4-GLUCANASE</td>
</tr>
<tr>
<td>1TVN</td>
<td>CELLULASE</td>
</tr>
<tr>
<td>1UWW</td>
<td>ENDOGLUCANASE</td>
</tr>
<tr>
<td>1WC2</td>
<td>ENDOGLUCANASE</td>
</tr>
<tr>
<td>2BOG</td>
<td>ENDOGLUCANASE E-2</td>
</tr>
<tr>
<td>2BW8</td>
<td>ENDOGLUCANASE</td>
</tr>
<tr>
<td>2E4T</td>
<td>ENDOGLUCANASE</td>
</tr>
<tr>
<td>2ENG</td>
<td>ENDOGLUCANASE V</td>
</tr>
<tr>
<td>2JEN</td>
<td>ENDO-BETA-1,4-GLUCANASE</td>
</tr>
<tr>
<td>2NLR</td>
<td>ENDOGLUCANASE</td>
</tr>
<tr>
<td>2V3G</td>
<td>ENDOGLUCANASE H</td>
</tr>
<tr>
<td>3ACH</td>
<td>ENDO-BETA-1,4-GLUCANASE</td>
</tr>
<tr>
<td>7A3H</td>
<td>ENDOGLUCANASE</td>
</tr>
</tbody>
</table>

**Table 4.4.** Eighteen PDB entries retrieved based on the selection criteria for EC 3.2.1.4.
4.6.3 β-lactamase (EC 3.5.2.6)

EC 3.5.2.6 is the enzyme classification number for β-lactamase, which is secreted by Gram-negative bacteria to hydrolyze the β-lactam ring of penicillins and cephalosporins. Based on the specificity of the β-lactamase, individual enzymes may be called penicillinase or cephalosporinase. There are four groups of β-lactamase that are roughly determined by inhibition of clavulanic acid. Enzymes in group 1 are cephalosporinases not inhibited by clavulanic acid while enzymes in group 4 are penicillinases that are also not inhibited by clavulanic acid. Group 2 contains penicillinases, cephalosporinases, or both that are inhibited by clavulanic acid. Group 3 holds the zinc based or metallo β-lactamases. Due to the increased size in group 2, it can further be divided into 8 subgroups based on the specificity of the enzymes. Table 4.5 showed the twelve retrieved PDB entries associated with β-lactamase based on the selection criteria.

<table>
<thead>
<tr>
<th>PDB Entries</th>
<th>Types</th>
</tr>
</thead>
<tbody>
<tr>
<td>1K38</td>
<td>BETA-LACTAMASE OXA-2</td>
</tr>
<tr>
<td>1K55</td>
<td>BETA-LACTAMASE OXA-10</td>
</tr>
<tr>
<td>1M2X</td>
<td>CLASS B CARBAPENEMASE BLAB-1</td>
</tr>
<tr>
<td>1M6K</td>
<td>BETA-LACTAMASE OXA-1</td>
</tr>
<tr>
<td>1MQO</td>
<td>BETA-LACTAMASE II</td>
</tr>
<tr>
<td>1NYM</td>
<td>BETA-LACTAMASE TEM</td>
</tr>
<tr>
<td>1O7E</td>
<td>L2 BETA-LACTAMASE</td>
</tr>
<tr>
<td>1ONG</td>
<td>BETA-LACTAMASE SHV-1</td>
</tr>
<tr>
<td>1ZKJ</td>
<td>EXTENDED-SPECTRUM BETA-LACTAMASE</td>
</tr>
<tr>
<td>2GMN</td>
<td>METALLO-BETA-LACTAMASE</td>
</tr>
<tr>
<td>2HDS</td>
<td>BETA-LACTAMASE</td>
</tr>
<tr>
<td>3G35</td>
<td>BETA-LACTAMASE CTX-M9A</td>
</tr>
</tbody>
</table>

Table 4.5. Twelve PDB entries retrieved based on the selection criteria for EC 3.5.2.6.
4.6.4 Methyltransferases (EC 2.1.1)

A methyltransferase (EC 2.1.1) is a type of transferase enzyme that transfers a methyl group from a donor to an acceptor and can be further classified into 195 subfamilies. Methylation often occurs on amino acids in protein structures or nucleic bases in DNA. DNA methylation is often utilized to silence and regulate genes without changing the original DNA sequence. While DNA methylation is an important regulator of gene transcription, its role in carcinogenesis has recently generated considerable interest. Compared with normal cells, the malignant cells show major disruptions in their DNA methylation patterns.

Methylation of amino acids in the formation of proteins leads to more diversity of possible amino acids and, therefore, more diversity of function. The methylation reactions occurring on nitrogen atoms in N-terminal and side-chain positions are generally irreversible. Table 4.6 shows the thirty-two retrieved PDB entries associated with methyltransferase based on the selection criteria.

<table>
<thead>
<tr>
<th>PDB Entries</th>
<th>E.C. Numbers</th>
</tr>
</thead>
<tbody>
<tr>
<td>1EJ0</td>
<td>2.1.1.-</td>
</tr>
<tr>
<td>1I1N</td>
<td>2.1.1.77</td>
</tr>
<tr>
<td>1JG1</td>
<td>2.1.1.77</td>
</tr>
<tr>
<td>1NTH</td>
<td>2.1.1.-</td>
</tr>
<tr>
<td>1V2X</td>
<td>2.1.1.34</td>
</tr>
<tr>
<td>2BLN</td>
<td>2.1.1.2</td>
</tr>
<tr>
<td>2F69</td>
<td>2.1.1.43</td>
</tr>
<tr>
<td>2G8O</td>
<td>2.1.1.45</td>
</tr>
<tr>
<td>2GB4</td>
<td>2.1.1.67</td>
</tr>
<tr>
<td>2NX1</td>
<td>2.1.1.-</td>
</tr>
<tr>
<td>2VK1</td>
<td>2.1.1.-</td>
</tr>
<tr>
<td>2Z6R</td>
<td>2.1.1.98</td>
</tr>
<tr>
<td>3AJD</td>
<td>2.1.1.-</td>
</tr>
<tr>
<td>3BO5</td>
<td>2.1.1.43</td>
</tr>
<tr>
<td>3C3Y</td>
<td>2.1.1.104</td>
</tr>
<tr>
<td>3CJS</td>
<td>2.1.1.-</td>
</tr>
<tr>
<td>3CKK</td>
<td>2.1.1.33</td>
</tr>
<tr>
<td>3DMG</td>
<td>2.1.1.-</td>
</tr>
<tr>
<td>3DOU</td>
<td>2.1.1.-</td>
</tr>
<tr>
<td>3DUW</td>
<td>2.1.1.-</td>
</tr>
<tr>
<td>3DXY</td>
<td>2.1.1.33</td>
</tr>
<tr>
<td>3EVF</td>
<td>2.1.1.56;2.7.7.48</td>
</tr>
<tr>
<td>3F9X</td>
<td>2.1.1.43</td>
</tr>
<tr>
<td>3FRH</td>
<td>2.1.1.-</td>
</tr>
<tr>
<td>3FTD</td>
<td>2.1.1.-</td>
</tr>
<tr>
<td>3FUT</td>
<td>2.1.1.-</td>
</tr>
<tr>
<td>3GS5</td>
<td>2.1.1.74</td>
</tr>
<tr>
<td>3G5T</td>
<td>2.1.1.145</td>
</tr>
<tr>
<td>3G89</td>
<td>2.1.1.-</td>
</tr>
<tr>
<td>3HNA</td>
<td>2.1.1.43</td>
</tr>
<tr>
<td>3HVI</td>
<td>2.1.1.6</td>
</tr>
<tr>
<td>3M6W</td>
<td>2.1.1.-</td>
</tr>
</tbody>
</table>

Table 4.6. Thirty-two PDB entries retrieved based on the selection criteria for EC 2.1.1.
4.6.5 Protein-Tyrosine Kinases (EC 2.7.10)

Tyrosine kinase is an enzyme that regulates many cellular functions through the transfer of a phosphate group from adenosine-5’-triphosphate (ATP) to the amino acid tyrosine on the protein. Phosphorylation of proteins by kinases is an important mechanism in cellular signal transduction and cellular activity regulation in response to external and internal stimuli. The cellular activities regulated by tyrosine kinases include apoptosis, cell cycle progression, cytoskeletal rearrangement, differentiation, development, the immune response, nervous system function, and transcription \(^{111}\). Dysregulation of protein kinases caused by the mutation or the fusion of tyrosine kinases with a partner protein has been implicated with cancer \(^{77;111}\).

Protein tyrosine kinases are divided into two main classes: receptor tyrosine kinases (EC 2.7.10.1) and nonreceptor tyrosine kinases (EC 2.7.10.2). Both receptor and nonreceptor tyrosine kinases have emerged as clinically useful drug target molecules for treating certain types of cancer \(^{77;141}\). Table 4.7 shows the eleven retrieved PDB entries associated with EC 2.7.10 based on the selection criteria.

<table>
<thead>
<tr>
<th>PDB Entries</th>
<th>E.C. Numbers</th>
</tr>
</thead>
<tbody>
<tr>
<td>2QOL</td>
<td>2.7.10.1</td>
</tr>
<tr>
<td>2REI</td>
<td>2.7.10.1</td>
</tr>
<tr>
<td>3CC6</td>
<td>2.7.10.2</td>
</tr>
<tr>
<td>3CQT</td>
<td>2.7.10.2</td>
</tr>
<tr>
<td>3EAZ</td>
<td>2.7.10.2</td>
</tr>
<tr>
<td>3EG3</td>
<td>2.7.10.2</td>
</tr>
<tr>
<td>3EWH</td>
<td>2.7.10.1</td>
</tr>
<tr>
<td>3F66</td>
<td>2.7.10.1</td>
</tr>
<tr>
<td>3G0E</td>
<td>2.7.10.1</td>
</tr>
<tr>
<td>3GEN</td>
<td>2.7.10.2</td>
</tr>
<tr>
<td>3KFA</td>
<td>2.7.10.2</td>
</tr>
</tbody>
</table>

Table 4.7. Eleven PDB entries retrieved based on the selection criteria for EC 2.7.10.
4.6.6 Protein-Serine/Threonine Kinases (EC 2.7.11)

Protein serine/threonine kinases (EC 2.7.11) differ from protein tyrosine kinases by phosphorylating the OH group of serine or threonine instead of tyrosine. EC 2.7.11 can be further divided into 31 subfamilies (Table 4.8). The activity of serine/threonine protease can be triggered

<table>
<thead>
<tr>
<th>Subfamilies</th>
<th>Descriptions</th>
</tr>
</thead>
<tbody>
<tr>
<td>2.7.11.1</td>
<td>Non-specific serine/threonine protein kinase.</td>
</tr>
<tr>
<td>2.7.11.2</td>
<td>Pyruvate dehydrogenase (acetyl-transferring) kinase.</td>
</tr>
<tr>
<td>2.7.11.3</td>
<td>Dephospho-[reductase kinase] kinase.</td>
</tr>
<tr>
<td>2.7.11.4</td>
<td>3-methyl-2-oxobutanoate dehydrogenase (acetyl-transferring) kinase.</td>
</tr>
<tr>
<td>2.7.11.5</td>
<td>Isocitrate dehydrogenase (NADP(+)) kinase.</td>
</tr>
<tr>
<td>2.7.11.6</td>
<td>Tyrosine 3-monoxygenase kinase.</td>
</tr>
<tr>
<td>2.7.11.7</td>
<td>Myosin heavy-chain kinase.</td>
</tr>
<tr>
<td>2.7.11.8</td>
<td>Fas-activated serine/threonine kinase.</td>
</tr>
<tr>
<td>2.7.11.9</td>
<td>Goodpasture-antigen-binding protein kinase.</td>
</tr>
<tr>
<td>2.7.11.10</td>
<td>I-kappa-B kinase.</td>
</tr>
<tr>
<td>2.7.11.11</td>
<td>eAMP-dependent protein kinase.</td>
</tr>
<tr>
<td>2.7.11.12</td>
<td>cGMP-dependent protein kinase.</td>
</tr>
<tr>
<td>2.7.11.13</td>
<td>Protein kinase C.</td>
</tr>
<tr>
<td>2.7.11.14</td>
<td>Rhodopsin kinase.</td>
</tr>
<tr>
<td>2.7.11.15</td>
<td>Beta-adrenergic-receptor kinase.</td>
</tr>
<tr>
<td>2.7.11.16</td>
<td>G-protein-coupled receptor kinase.</td>
</tr>
<tr>
<td>2.7.11.17</td>
<td>Calcium/calmodulin-dependent protein kinase.</td>
</tr>
<tr>
<td>2.7.11.18</td>
<td>Myosin light-chain kinase.</td>
</tr>
<tr>
<td>2.7.11.19</td>
<td>Phosphorylase kinase.</td>
</tr>
<tr>
<td>2.7.11.20</td>
<td>Elongation factor 2 kinase.</td>
</tr>
<tr>
<td>2.7.11.21</td>
<td>Polo kinase.</td>
</tr>
<tr>
<td>2.7.11.22</td>
<td>Cyclin-dependent kinase.</td>
</tr>
<tr>
<td>2.7.11.23</td>
<td>RNA-polymerase subunit kinase.</td>
</tr>
<tr>
<td>2.7.11.24</td>
<td>Mitogen-activated protein kinase.</td>
</tr>
<tr>
<td>2.7.11.25</td>
<td>Mitogen-activated protein kinase kinase.</td>
</tr>
<tr>
<td>2.7.11.26</td>
<td>Tau protein kinase.</td>
</tr>
<tr>
<td>2.7.11.27</td>
<td>Acetyl-CoA carboxylase kinase.</td>
</tr>
<tr>
<td>2.7.11.28</td>
<td>Tropomyosin kinase.</td>
</tr>
<tr>
<td>2.7.11.29</td>
<td>Low-density-lipoprotein receptor kinase.</td>
</tr>
<tr>
<td>2.7.11.30</td>
<td>Receptor protein serine/threonine kinase.</td>
</tr>
<tr>
<td>2.7.11.31</td>
<td>Hydroxymethylglutaryl-CoA reductase (NADPH) kinase.</td>
</tr>
</tbody>
</table>

Table 4.8. List of subfamilies associated with EC 2.7.11.

EC 2.7.11 can be further classified into 31 subfamilies.
by DNA damage and chemical signals, such as

cAMP/cGMP, diacylglycerol, and Ca2+/calmodulin.

Compared to tyrosine kinases, serine/threonine
kinases received comparatively less attention in cancer
studies. However, a recent study has found frequent
alterations in the expression of serine/threonine kinases
in human cancers\textsuperscript{31}. Table 4.9 shows the twenty-two
retrieved PDB entries associated with EC 2.7.11 based
on the selection criteria.

<table>
<thead>
<tr>
<th>PDB Entries</th>
<th>E.C. Numbers</th>
</tr>
</thead>
<tbody>
<tr>
<td>1UNR</td>
<td>2.7.11.1</td>
</tr>
<tr>
<td>2HLR</td>
<td>2.7.11.30</td>
</tr>
<tr>
<td>2IZR</td>
<td>2.7.11.1</td>
</tr>
<tr>
<td>2IZX</td>
<td>2.7.11.11</td>
</tr>
<tr>
<td>2J0I</td>
<td>2.7.11.1</td>
</tr>
<tr>
<td>2R3I</td>
<td>2.7.11.22</td>
</tr>
<tr>
<td>2RIK</td>
<td>2.7.11.1</td>
</tr>
<tr>
<td>2W5A</td>
<td>2.7.11.1</td>
</tr>
<tr>
<td>3A99</td>
<td>2.7.11.1</td>
</tr>
<tr>
<td>3BHLY</td>
<td>2.7.11.1</td>
</tr>
<tr>
<td>3CCD</td>
<td>2.7.11.-</td>
</tr>
<tr>
<td>3F6Q</td>
<td>2.7.11.1</td>
</tr>
<tr>
<td>3FJQ</td>
<td>2.7.11.11</td>
</tr>
<tr>
<td>3FVH</td>
<td>2.7.11.21</td>
</tr>
<tr>
<td>3GP2</td>
<td>2.7.11.17</td>
</tr>
<tr>
<td>3K21</td>
<td>2.7.11.1</td>
</tr>
<tr>
<td>3KHF</td>
<td>2.7.11.1</td>
</tr>
<tr>
<td>3KNB</td>
<td>2.7.11.1</td>
</tr>
<tr>
<td>3LKM</td>
<td>2.7.11.7</td>
</tr>
<tr>
<td>3NSZ</td>
<td>2.7.11.1</td>
</tr>
<tr>
<td>3OEF</td>
<td>2.6.11.24</td>
</tr>
<tr>
<td>3PA3</td>
<td>2.6.11.1</td>
</tr>
</tbody>
</table>

Table 4.9. Twenty-two PDB entries retrieved based on the
selection criteria for EC 2.7.11.
4.7 Modeling Results

4.7.1 Superoxide Dismutase (EC 1.15.1.1)

After applying the classification method described in Chapter 2 to separate the EC 1.15.1.1 from the enzyme diversity set, nine different models with corresponding z-score, sensitivity, and specificity values were obtained. As illustrated in Figure 4.7, the z-score was the highest, thus most significant, when only the top five ranked features were considered. This particular plot was to help guiding the selection of features when multiple z-scores are close to one another in values. Since the z-score associated with the top five ranked features is distinct from the rest of the z-scores, sensitivity and specificity will not be considered for selecting the important features of superoxide dismutase. With the top 5 ranked features, it is possible to achieve sensitivity of 1.0 and specificity above 0.90.

Figure 4.7. Identifying the most significant model through z-scores for the EC1.15.1.1 dataset.

This semi-log plot indicates that the z-score associated with the top five features is the highest. The y-axis on the left (Percentage) is for sensitivity and specificity while the y-axis on the right is for the z-score.
Figure 4.8 shows the top five ranked features associated with EC 1.15.1.1. The descriptor names of these top five ranked features indicated that four tetrahedral categories were identified: 6600, 9997, 9999, and 9995. For tetrahedral category 6600, a larger value in total volume was preferred for EC 1.15.1.1. For both 9997 and 9999 tetrahedral categories, a larger value in total hidden surface area was preferable. For tetrahedral category 9995, both total volume and total hidden surface area were listed in the top five features, which implied that a tetrahedron with large volume but small hidden surface area is likely to associate with EC 1.15.1.1.

Mapping these four categories of tetrahedra on to the 14 structures showed interesting results, as they tend to cluster around the binding ligands (Figure 4.9 and Figure 4.10). Among the fourteen structure entries, tetrahedra with category 6600 appeared only on SODs that bind to copper/zinc (Cu/Zn). In addition, tetrahedra with category 9999 did not appear on all the Cu/Zn binding SOD. The remaining three categories of tetrahedra appeared to cluster around the binding ligands, with the exception of PDB entry 1T6U, which was the only nickel binding SOD among the 14 structure entries. Although the tetrahedra in PDB entry 1T6U did not cluster around the ligand, nickel ion, these tetrahedra did cluster at six local regions. Incidentally, there are also six nickel ions in this PDB entry.
Figure 4.9. Mapping the four identified tetrahedral categories onto structures associated with SOD binding to Fe$^{3+}$ and Mn$^{2+}$.

The four categories of tetrahedra clustered around the binding ligands. Among the 14 structure entries, the tetrahedron with category 6600 did not occur in SODs that bind to either iron or manganese. The remaining three categories of tetrahedra appeared to cluster around the binding ligands. Both PDB entries (1BSM and 2P4K) belong to the same SOD family.
When mapping the four categories of tetrahedra onto the fourteen structure entries, tetrahedron with category 6600 appeared only on SODs that bind to Cu\(^{2+}\)/Zn\(^{2+}\). The additional molecule in the PDB entry 3CE1 is an acetate ion. For the PDB entry 1T6U, the identified tetrahedra did not cluster around the binding ligand; however, it was the only nickel binding SOD among the fourteen structure entries.

**Figure 4.10.** Mapping the four identified tetrahedral categories onto structures associated with SOD binding to Cu\(^{2+}\)/Zn\(^{2+}\) and Ni\(^{2+}\).
For PDB entry 1TO4, which is a copper and zinc binding SOD with two identical chains illustrated in Figure 4.11, only two of the four tetrahedral categories appear: 6600 and 9997. Tetrahedral category 9997 was identified to locate near the zinc ion, but it only appeared on one of the two chains.

Figure 4.11. Mapping the four identified tetrahedral categories onto PDB entry, 1TO4.

The PDB entry, 1TO4, contains two identical chains, but the tetrahedral category 9997, which was found to be near the zinc ion, only appears on one chain and not the other. None of the four identified tetrahedral categories were found near the copper (II) ions. The coordinate of Cu$^{2+}$ is imprecise as shown in the figure.
4.7.2 Cellulase (EC 3.2.1.4)

Nine different models with corresponding z-score, sensitivity, and specificity values were obtained after applying the classification method described in Chapter 2 to separate the EC 3.2.1.4 from the enzyme diversity set. As illustrated in Figure 4.12, there were a few models with z-scores in proximity of one other. The model built with the top 20 ranked features achieved a z-score of 10.95 while the z-score associated with the model with the top ten ranked features was 10.67. Since the z-scores of these two models were above 10.5, we used both sensitivity and specificity to guide the final selection of features. The set of the 20 top ranked features had higher z-score (10.95), sensitivity (0.94) and specificity (0.90) than the set of the 10 top ranked features, so we selected the top 20 ranked features as the important features associated with cellulase.

![Figure 4.12. Identifying the most significant model through z-score for the EC 3.2.1.4 dataset.](image)

This semi-log plot indicates that the z-scores associated with the top ten and top twenty features are both similar in values. A model built with top twenty features is selected for the classification performance in sensitivity and specificity. The y-axis on the left (Percentage) is for sensitivity and specificity while the y-axis on the right is for z-score.
Figure 4.13 shows the top twenty ranked features associated with the EC 3.2.1.4. The descriptor names of these top twenty ranked features indicated that eighteen tetrahedral categories were identified as significant to cellulase. For tetrahedral categories 5330 and 8870, both total volume and total hidden surface area were listed in the top twenty features, which implied that a tetrahedron with large volume and hidden surface area is likely to associate with EC 3.2.1.4. In addition, the lower value in total exposed surface area for type 9751 and type 9811 were also preferable for cellulase.

Mapping these 18 categories of tetrahedra onto structures indicated that not all 18 categories of tetrahedra were available in each structure entry (Figure 4.14). Each structure entry contained different sets of the 18 categories of tetrahedra. Although not all the tetrahedra were located in proximity of the binding ligands, some do cluster around the ligands. In the example of PDB entry 2BOG, there are three structurally similar ligands located in the same binding pocket. For PDB entry 1OA2, the binding pockets for the two different ligands are far apart.
Figure 4. 14. Mapping the 18 identified tetrahedral categories on to the structures associated with cellulases.

The selected structure entries of cellulases did not contain all 18 categories of tetrahedra. Although not all the tetrahedra were located in proximity of the binding ligands, some do cluster around the ligands. There are three structurally similar ligands located in the same binding pocket for PDB entry 2BOG. For PDB entry 1OA2, the binding pockets for the two different ligands are far apart.
4.7.3 β-lactamase (EC 3.5.2.6)

Nine different models with corresponding z-score, sensitivity, and specificity values were obtained after applying the aforementioned classification method to separate EC 3.5.2.6 from the enzyme diversity set. As illustrated in Figure 4.15, the model built with the top ten ranked features achieved a much higher z-score than the other models. With the top ten ranked features, it was possible to achieve sensitivity of 0.92 and specificity above 0.88. Figure 4.16 shows the top ten ranked features associated with EC 3.5.2.6. The descriptor names from these top ten ranked features indicated that there were six tetrahedral categories that were significant to β-lactamase: 3322, 5442, 5543, 7332, 9555 and 9965. The loading associated with the top ten ranked features were all positive, which indicating higher values were preferable. For tetrahedra with categories 5442, 5543, 9555, and 9965, both total volume and total hidden surface area were indicated as important features, which implied that a tetrahedron with large volume and hidden surface area is likely to associate with EC 3.5.2.6. In addition, the exposed surface area associated with tetrahedral category 3322 received the largest weight.

![Figure 4.15. Identifying the most significant model through z-score for the EC 3.5.2.6 dataset.](image-url)

This semi-log plot indicates the z-score associated with the top ten features is the highest. The y-axis on the left (Percentage) is for sensitivity and specificity while the y-axis on the right is for z-score.
Mapping these six categories of tetrahedra onto the structures showed that not all six categories of tetrahedra appeared in the selected structure entries (Figure 4.17 and Figure 4.18). Each structure entry contained different sets of the six categories of tetrahedra, with tetrahedral category 9555 being the most frequent. Among the twelve structure entries, tetrahedra with category 9555 was typically found next to a binding ligand that is not a metal ion, except PDB entry 1M2X. For that structure entry, the tetrahedron next to the binding ligand was category 9965, which was one of the six identified tetrahedral categories.

Mapping the tetrahedra on to the structures also showed mixed results for structure entries that contained multiple binding pockets. In some cases, the six identified categories of tetrahedra might not be observed in all the binding pockets. In the example of PDB entry 3G35, there are two binding pockets for the same ligand; however, the identified tetrahedral categories were found only in one of the two pockets. The two binding pockets are different in amino acid composition and, the original paper 36 focused on the binding site that contains the amino acids belonging to the tetrahedra identified.

![Figure 4.16. The ten most significant features associated with β-lactamase.](image)

The bar graph shows the ten most significant features selected by DWD and their corresponding weights. All features have positive weights.
Figure 4. 17. Mapping the six identified tetrahedral categories on to the 1M2X and 1NYM structure entries (EC 3.5.2.6 family).

Mapping the six categories of tetrahedra on to the structures showed the selected structure entries contained only a subset of the six categories of tetrahedron. Tetrahedral category 9555, being the most frequent, was typically found next to a binding ligand that is not a metal ion, except PDB entry 1M2X. For PDB entry 1M2X, the tetrahedron next to the binding ligand was category 9965, which was one of the six identified tetrahedral categories.
Figure 4. 18. Mapping the six identified tetrahedral categories on to the 3G35 structure entry (EC 3.5.2.6 family).

Mixed results for structure entries contained multiple binding pockets that were observed when mapping the tetrahedra on to the structures. In some cases, the six identified categories of tetrahedra might not be observed in all the binding pockets. For example, PDB entry 3G35 contains two binding pockets for the same ligand; however, the identified tetrahedral categories were found only in one of the two pockets.
4.7.4 Methyltransferases (EC 2.1.1)

Nine different models with corresponding z-score, sensitivity, and specificity values were obtained after applying the classification method described in Chapter 2 to separate the EC 2.1.1 from the enzyme diversity set. EC 2.1.1 contains enzymes with more generalized function as hinted by the three-digit Enzyme Classification number and can be further classified into 195 subfamilies. As illustrated in Figure 4.19, the most significant model (with the highest z-score) was built with the top 500 ranked features. Although the model was capable to perfectly separate the structure entries associated with EC 2.1.1 from the enzyme diversity set (both sensitivity and specificity equal to one), it was not feasible to analyze or interpret the model due to the large number of features incorporated.

Figure 4.19. Identifying the most significant model through z-score for the EC 2.1.1 dataset.

This semi-log plot indicates the z-score associated with the top 500 features is the highest. The y-axis on the left (Percentage) is for sensitivity and specificity while the y-axis on the right is for z-score.
### 4.7.5 Protein-Tyrosine Kinases (EC 2.7.10)

Nine different models with corresponding z-score, sensitivity, and specificity values were obtained after applying the classification method described in Chapter 2 to separate the EC 2.7.10 from the enzyme diversity set. EC 2.7.10 contains enzymes with more generalized function and can be further classified into two additional subfamilies. As illustrated in Figure 4.20, both models built with top 50 and top 100 ranked features achieved similar values in z-scores, which are 7.09 and 7.06 respectively. Since the z-scores of these two models were above 7.0, we used both sensitivity and specificity to guide the final selection of features. Based on the classification performance (both sensitivity and specificity equal to 1.0), we selected the top 100 ranked features as the important features associated with protein-tyrosine kinases. Similarly to the results for EC 2.1.1, it was not feasible to analyze or interpret the model due to the large number of features incorporated.

![Figure 4.20](image)

**Figure 4.20.** Identifying the most significant model through z-score for the EC 2.7.10 dataset.

This semi-log plot indicates that the z-scores associated with the top 50 and top 100 features are both similar in values. A model built with top 100 features is selected for the performance in sensitivity and specificity. The y-axis on the left (Percentage) is for sensitivity and specificity while the y-axis on the right is for z-score.
4.7.6 Protein-Serine/Threonine Kinase (EC 2.7.11)

Nine different models with corresponding z-score, sensitivity, and specificity values were obtained after applying the classification method described in Chapter 2 to separate the EC 2.7.11 from the enzyme diversity set. EC 2.7.11 contains enzymes with more generalized function and can be further classified into 31 subfamilies. As illustrated in Figure 4.21, the model built with the top 1,000 ranked features achieved the highest z-score, sensitivity (1.0), and specificity (1.0). As noted in other results obtained from enzymes with more generalized function, it was not feasible to analyze or interpret the model due to the large number of features incorporated.

Figure 4.21. Identifying the most significant model through z-scores for the EC 2.7.11 dataset.

This semi-log plot indicates the z-score associated with the top 1000 features is the highest. The y-axis on the left (Percentage) is for sensitivity and specificity while the y-axis on the right is for z-score.
4.8 Evaluating the Stability of the Delaunay Simplices

The definition of the Delaunay tessellation depends on the precise coordinate values given to its points. In the scenario that point coordinates are known only imprecisely, the stability and robustness of the resulting Delaunay simplices under changes to the input coordinates could be questioned. The results in enzyme families EC 1.15.1.1 and EC 3.5.2.6, where the Delaunay simplices captured only one binding pocket of the two available, might be explained by the imprecision of the input coordinates. In PDB entry, 3G35, there are two binding pockets for the same ligand but the amino acid compositions for these two sites are completely different. However, in the case of PDB entry, 1TO4, there are two identical binding pockets within the structure, and the tetrahedral categories identified as significant to the protein function only appeared in one of the two binding pockets.

To evaluate the stability and robustness of the Delaunay simplices, a Gaussian sphere with a radius of approximately 0.5 Å was applied to each of the original Cα atom coordinates in PDB entry, 1TO4, to permit the shifting of the points. The shifting of each Cα atom is independent of one another, and 30 structures were generated with the new Cα coordinates. Delaunay tessellation was then applied to these 30 modified structures, and tetrahedra with edges longer than 12 Å were removed. Within each of the 30 tessellation outcomes, a search for the four tetrahedral categories, i.e., 6600, 9995, 9997, and 9999, was performed. Figure 4.22 shows all the tetrahedra associated with the four tetrahedral types that are found in the search result.

The three tetrahedra associated with category 6600 that were found in the original structure also appeared in all the 30 modified structures. These tetrahedra were formed by the amino acids Gly (9), Gly (55), Cys (56), Ala (144), and Cys (145). While the tetrahedron formed by Gly (9), Gly(55), Cys (56), and Cys (145) was found in only one of the two chains, the tetrahedron formed by Gly (9), Cys (56), Ala (144), and Cys (145) appeared in both chains.
For tetrahedral categories 9997, new tetrahedra were found in addition to the one tetrahedron appearing in the original structure. The tetrahedron formed by amino acids His (62), Asn (64), His (70), and His (79) that appeared in the original structure was also found on the modified structures in 21 out of 30 instances. The same four amino acids on the other chain also formed a tetrahedron in 9 out of 30 instances of modified structures. Additionally, the tetrahedron formed by Lys (38), Lys (41), His (42), and Asn (121) that did not appear in the original structure was found in four out of the 30 instances.

The outcome from this evaluation indicated that the Delaunay simplices calculated solely from the original X-ray crystal structures did not fully capture the comprehensive local neighborhood relationship of the amino acids within a protein structure. By treating the atom
coordinates of a protein crystal structure as flexible points rather than fixed ones, the resulting Delaunay simplices, which were obtained from a set of thirty structures with perturbed atom coordinates, could capture a more comprehensive local neighborhood relationship of the amino acids within the protein structure. Since our protocol for studying the protein structure-function relationship was designed to utilize the atom coordinates as fixed points, this finding raised concerns regarding the performance of our model in protein function prediction.

There are two possible methods to predict a protein function based on this developed QSAR-like strategy: direct and indirect. The direction prediction, which is a straightforward utilization of a model, would be most likely to suffer from a higher type II error (false negative) caused by a set of simplices that did not fully capture the local packing order of amino acids in protein structures. The same cause for a higher type II error would also likely affect indirect prediction but with less impact, because an additional search for common simplices with similar geometric properties was performed in the tetrahedral categories identified by the model. The presence of these resulting common simplices would then be used to predict the function of other protein structures that are not part of the training set. However, grouping simplices based on similar geometric properties required additional analysis to critically assess the appropriate binning of each geometric property, which was not part of the scope of this research; thus, indirect prediction was not implemented in this research.
4.9 Comparison with PROSITE

PROSITE is a protein database consisting of manually curated amino acid profiles and patterns that describe protein domains, families and functional sites. Applications of these profiles and patterns include both identifying possible functions for newly discovered proteins and analyzing known proteins for previously undetermined activities. These patterns were utilized in this comparison study.

To compare the performance between our method and PROSITE, we used ScanPROSITE, a web-based tool for detecting protein sequences that match PROSITE patterns and/or profiles. Additionally, a separate test set containing both positive and negative classes was selected to benchmark both methods. Standard criteria for evaluating the performance of information retrieval systems, such as precision, recall (sensitivity), and true negative rate (specificity), were calculated to compare the two methods. For PROSITE evaluation, a PDB entry was considered to be a positive hit if its sequence matched at least one of the evaluated PROSITE profiles or patterns. In total, nine PROSITE patterns were utilized in this benchmark study – four patterns for E.C. 1.15.1.1 and five patterns for E.C. 3.5.2.6.

4.9.1 Test Set Selection Criteria for Both E.C. 1.15.1.1 and E.C. 3.5.2.6

For the positive class, a set of PDB entries from family members belonging to either E.C. 1.15.1.1 or E.C. 3.5.2.6 was created, while the negative class contained PDB entries from an enzyme diversity set, excluding E.C. 1.15.1.1 and E.C. 3.5.2.6. The test set for each enzyme set, i.e., E.C. 1.15.11 and E.C. 3.5.2.6, was created by combing the corresponding positive class with the negative class. The selection criteria for the positive and negative classes were similar to that of the enzyme class of interest and enzyme diversity set, respectively. This benchmark study excluded E.C. 3.2.1.4 because none of the PROSITE patterns was specific for this particular enzyme family.
Several criteria were used during the PDB entry selection to ensure the quality and size of the test set. First, the X-ray resolution for the positive class of the test set was relaxed to 2.00 Å to include a greater number of crystal structures. Structure entries previously included in our modeling procedure were removed from the test sets to allow for fair comparison. Similarly, all entries selected were released within the past three years, i.e., between 01/01/2008 and 12/31/2011, to ensure that the resulting structure entries were not used in generating the PROSITE signatures applied to this study, which were last updated in 2006. The resulting test sets, based on these selection criteria, contained 421 structure entries for the negative class, 15 structure entries for E.C. 1.15.1.1 and 22 for E.C. 3.5.2.6. The PDB entries for the two positive classes are listed in Table 4.10.

### Table 4.10

<table>
<thead>
<tr>
<th>PDB Entries</th>
<th>E.C. Numbers</th>
<th>PDB Entries</th>
<th>E.C. Numbers</th>
</tr>
</thead>
<tbody>
<tr>
<td>2JLP</td>
<td>1.15.1.1</td>
<td>2V1Z</td>
<td>3.5.2.6</td>
</tr>
<tr>
<td>2RCV</td>
<td>1.15.1.1</td>
<td>2WHG</td>
<td>3.5.2.6</td>
</tr>
<tr>
<td>2WYT</td>
<td>1.15.1.1</td>
<td>2WK0</td>
<td>3.5.2.6</td>
</tr>
<tr>
<td>3AK2</td>
<td>1.15.1.1</td>
<td>2X02</td>
<td>3.5.2.6</td>
</tr>
<tr>
<td>3DC5</td>
<td>1.15.1.1</td>
<td>2ZD8</td>
<td>3.5.2.6</td>
</tr>
<tr>
<td>3G4Z</td>
<td>1.15.1.1</td>
<td>2ZQ7</td>
<td>3.5.2.6</td>
</tr>
<tr>
<td>3H1S</td>
<td>1.15.1.1</td>
<td>3E2L</td>
<td>3.5.2.6</td>
</tr>
<tr>
<td>3JS4</td>
<td>1.15.1.1</td>
<td>3FKW</td>
<td>3.5.2.6</td>
</tr>
<tr>
<td>3K9S</td>
<td>1.15.1.1</td>
<td>3G4P</td>
<td>3.5.2.6</td>
</tr>
<tr>
<td>3KBE</td>
<td>1.15.1.1</td>
<td>3HBR</td>
<td>3.5.2.6</td>
</tr>
<tr>
<td>3KKY</td>
<td>1.15.1.1</td>
<td>3I11</td>
<td>3.5.2.6</td>
</tr>
<tr>
<td>3L9Y</td>
<td>1.15.1.1</td>
<td>3IOF</td>
<td>3.5.2.6</td>
</tr>
<tr>
<td>3LO</td>
<td>1.15.1.1</td>
<td>3ISG</td>
<td>3.5.2.6</td>
</tr>
<tr>
<td>3LSU</td>
<td>1.15.1.1</td>
<td>3L6N</td>
<td>3.5.2.6</td>
</tr>
<tr>
<td>3PU7</td>
<td>1.15.1.1</td>
<td>3LEZ</td>
<td>3.5.2.6</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3M8T</td>
<td>3.5.2.6</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3MZF</td>
<td>3.5.2.6</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3NY4</td>
<td>3.5.2.6</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3P09</td>
<td>3.5.2.6</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3Q6V</td>
<td>3.5.2.6</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3Q6X</td>
<td>3.5.2.6</td>
</tr>
<tr>
<td></td>
<td></td>
<td>3S1Y</td>
<td>3.5.2.6</td>
</tr>
</tbody>
</table>

### 4.9.2 Comparison Results

Both methods were able to retrieve the positive class for both E.C. 1.15.1.1 and E.C. 3.5.2.6 comparably well. For E.C. 1.15.1.1, our method achieved a recall (sensitivity) value of 0.87 by retrieving thirteen out of the fifteen PDB entries associated with the positive class while ScanPROSITE was able to retrieve one addition entry and achieved a recall value of 0.93.
However, when comparing the two lists of retrieval entries, only twelve entries were correctly identified by both methods. ScanPROSITE failed to identify 3G4Z, while our model missed 3JS4 and 3KBE. The failure to identify 3G4Z by PROSITE was likely due to the lack of manually curated patterns for the nickel-binding SOD, which belonged to E.C. 1.15.1.1. A similar outcome was also observed in the test set for E.C. 3.5.2.6. Both methods were able to retrieve twelve out of the twenty-two PDB entries from the positive class, yielding a recall value of 0.55. Comparison of each of the twelve entries identified indicated only four common entries between the two methods. Used together, 100% of the positive class was correctly identified for E.C. 1.15.1.1, and 91% of the positive class was identified for E.C. 3.5.2.6, suggesting that the two protocols are potentially complementary. Figure 4.23 shows the benchmarking results.

![Figure 4.23. Benchmarking results.](image)

Both methods performed similarly in sensitivity and specificity. When used together, we observe an increase in sensitivity for both E.C. 1.15.1.1 and E.C. 3.5.2.6, suggesting that the two protocols are potentially complementary.
However, ScanPROSITE identified no false positives for either test set, while our method incorrectly labeled 33 negatives (0.93 in true negative rate or specificity) and 50 (0.87 in true negative rate or specificity) in the E.C. 1.15.1.1 and E.C. 3.5.2.6 datasets, respectively. Due to the number of false positives (or type I error) in our model, the precision of our models for predicting the test sets for E.C. 1.15.1.1 and E.C. 3.5.2.6 was 0.28 and 0.19, respectively.

To improve the number of true positives and false positives predicted by our model, two possible enhancements could be made to our protein descriptors that would enable DWD-VS to identify a more specific set of Delaunay simplices that contributed to a given enzyme function. Our protein descriptors, which utilized Delaunay simplices to characterize protein structures, could be enhanced by considering protein structures obtained from experimental X-ray crystallography as flexible entities rather than rigid ones. By calculating the protein descriptors from a set of movable points instead of fixed ones, the Delaunay simplices could then be able to capture a more comprehensive local neighborhood relationship between Cα atoms, as was observed in our Delaunay simplices stability evaluation (Section 4.7).

Additionally, we could further distinguish Delaunay simplices with additional geometric characteristics, such as chirality and volume. Especially the chirality of a Delaunay simplex, which could be defined by the four amino acid vertices and the Cahn-Ingold-Prelog priority rules, would provide the relative spatial orientation of the four amino acids to distinguish between Delaunay simplices with the same composition.

**4.10 Conclusion**

The goal for implementing the protein descriptors was to study the structure-function relationship of proteins. By combining variable selection DWD with the novel protein descriptors, our intention was to identify important amino acids that are common in proteins with similar functions. The results of our study indicated that our QSAR-like strategy helped to identify groups of amino acid residues located near the corresponding binding ligand, which indicated that
the identified amino acids could be part of the binding pocket. Additionally, categorized Delaunay simplices and their geometric properties were found to encode valuable structural information and enabled DWD-VS to identify a specific set of Delaunay simplices that attributed to a given enzyme function, especially when enzyme function was more specific, such as observed in the studies of EC 1.15.1.1, EC 3.2.1.4, and EC 3.5.2.6. Mapping identified Delaunay simplices onto their corresponding protein structures provided insights and led to generation of hypotheses regarding the important binding residues.

The results of the comparison study for E.C. 1.15.1.1 and E.C. 3.5.2.6 against ScanPROSITE indicated that when combining the protein descriptors with variable selection DWD could provide a potentially complementary method to PROSITE, which utilizes expert knowledge to annotate protein functions. Currently, this automated strategy is based on objective structural data and proof of the concept for use of a QSAR-like analysis for protein function annotation. Additional enhancements to the protein descriptors, such as flexible structure sampling and geometric categorization of Delaunay simplices, would likely improve the precision and recall for this QSAR-like strategy.

For applicability in future studies of protein families and for predicting structures with unknown functions, the prediction procedure will also have to improve. One possible improvement could result from the coupling of direct and indirect prediction using our method. Specifically, direct prediction involves using a model built for a specific protein family, while indirect prediction (discussed in Section 4.7) refers to a similarity search for tetrahedra within the training set that display similar geometric characteristics, i.e., the use similar tetrahedra to imply similar function.

Overall, our automated method showed comparable sensitivity to that of ScanPROSITE, which showed considerable improvement and complementarity when the two methods were combined. However, our method suffers from poor specificity, a problem that could likely be resolved with the addition of the geometric descriptors mentioned above and by the use of both
direct and indirect prediction. Despite this drawback, the complementary results suggest that our automated method is able to correctly identify functionally similar proteins that PROSITE patterns, which were curated using expert knowledge, completely miss. The nature of these missed proteins suggests that our method is better at identifying protein functional homologs with a more distant similarity. Thus, our method could prove useful for more difficult protein annotation, especially with further improvement and refinement of the protein descriptors.
Chapter 5

Summary

The increasing availability of biological and chemical data has led to a critical need for cheminformatics and bioinformatics tools to analyze the data. One of the major challenges involved in this data analysis is HDLSS. To overcome the statistical difficulties inherent in HDLSS data, DWD was improved by adding variable selection. In a simulation using imbalanced, HDLSS data, DWD with variable selection (DWD-VS) significantly improved model prediction performance compared to SVM and DWD without variable selection. Analysis of models indicated that DWD-VS consistently achieved high-prediction accuracy by removing greater than 99.9% of the noise while retaining up to 70% of the signal through informative descriptors. These simulation results suggested that DWD-VS could be used to obtain a better understanding of the underlying biological activities; thus, DWD-VS could provide faster and more cost-effective models by identifying predictor variables to achieve high prediction accuracy. Similar results were also observed in QSAR studies; however, the differences between SVM, DWD, and DWD-VS were all too small to claim one method is much better than the others based solely on the prediction accuracy.

While the strategies used to build predictive models are important for identifying the structure-activity relationship of biomolecules, it is also essential to employ descriptors that encode molecular characteristics associated with a target property. The 3D chiral atom-pair descriptors were developed to evaluate the effect that the degree of chirality contributes to various target properties. For the selected stereochemistry-dependent datasets, the 3D chiral atom-pair descriptors showed better classification performance than 3D non-chiral atom-pair descriptors (without chiral atom types), and as expected, both sets of 3D atom-pair descriptors performed
better than 2D Dragon descriptors. Not unexpectedly, for datasets with minimal chirality information, classification performance of developed chirality-sensitive descriptors is similar to the performance with either 2D Dragon or 3D chirality-insensitive atom-pair descriptors.

The QSAR studies of the stereochemistry dependent datasets also suggested that the variable selection procedure implemented in the DWD-VS was likely to miss groups of descriptors that contributed to biological activity but were highly correlated to one another. For two datasets where the effect of stereochemistry on biological activities was not apparent, lower prediction accuracy was observed in the DWD-VS models obtained from the combined descriptors (2D Dragon + either 3D chiral atom-pair or 3D non-chiral atom-pair descriptors) than DWD-VS models obtained from each of the 3D atom-pair descriptors alone. By using a simple merge to combine different descriptor matrices, more instances of highly correlated descriptors are likely to occur, especially if 3D atom-pair descriptors encode redundant information as the 2D Dragon descriptors. These highly correlated descriptors affect the weighting assigned by DWD internal algorithms, causing the descriptors that should be selected to be dropped instead. To improve the performance of the variable selection DWD, correlation between descriptors will have to be considered as part of the variable selection procedure.

To study the structure-function relationship of proteins, combining novel protein descriptors with DWD-VS provided a potential complement to ScanPROSITE, which utilized expert knowledge to annotate protein function. This QSAR-like strategy helped to identify groups of amino acid residues that were a part of the binding pocket, as observed in the studies of EC 1.15.1.1, EC 3.2.1.4, and EC 3.5.2.6. Mapping identified tetrahedra onto the corresponding protein structures provided insights and hypotheses on the important binding residues; however, it also indicated two drawbacks of the protein descriptors. First, Delaunay simplices in protein structures are sensitive to the imprecision of the input coordinates -- some examples capture one binding pocket when there are multiple binding pockets available. Second, if multiple tetrahedra from the same category are identified as significant, the current implementation of tetrahedral
categories does not allow users to further narrow down these tetrahedra to a smaller set that is critical to binding. To address the first shortcoming, one solution is to apply a Gaussian sphere around each coordinate to sample other likely locations of the atoms and then generate a descriptor matrix that summarizes all the resulting Delaunay simplices and their associated geometric properties. This solution is likely to address only the scenario where multiple binding pockets within a protein structure have the same amino acid composition, e.g. PDB entry 1TO4. In the case of multiple binding pockets that are different in amino acid composition, e.g. PDB entry 3G35, the binding pocket that was not identified in our result may not be the primary site since the original literature only focused on the binding pocket identified in this research. As for the second shortcoming, a possible solution would be to further categorize the tetrahedron based on values of geometric properties.

In summary, the outcomes of this research provide cheminformatics and bioinformatics tools for modeling and analyzing the structure-activity relationship within biomolecular data through novel molecular descriptors and a variable selection based statistical machine learning method. Specifically, the technologies set forth in this dissertation address the HDLSS imbalanced categorical characteristics present in many biomolecular datasets. The data evaluated in this research were embedded in dimension that ranged from 2 to 42 times the sample size, and multiple imbalanced categorical datasets were evaluated, including datasets with the positive class contributing less than 5% of the total data. Our results indicated that DWD-VS gave models with high external prediction power and the estimated intrinsic dimension that is usually lower than the sample size and contained predictive descriptors that characterize the target biological property. We also showed that the developed chirality-sensitive descriptors increased the predictive power of QSAR models obtained for stereochemistry dependent datasets. In addition we demonstrated that our method is better at identifying protein functional homologs with a more distant similarity and could prove useful for more difficult protein annotation, especially with further improvement and refinement of the protein descriptors. Overall, these developed
descriptors and DWD-VS provide not only tools for modeling and analyzing the structure-activity relationship of biomolecular data but also direction for future advancements in chemical compound and protein classification.
Reference List


47. Ding Z. Diversified Ensemble Classifiers for Highly Imbalanced Data Learning and their Application in Bioinformatics. 1-116. 5-7-2011. Georgia State University.


55. Food and Drug Administration. FDA's policy statement for the development of new stereoisomeric drugs. 5-1-1992. Ref Type: Bill/Resolution


99. OpenEye. QUACPAC. [1.3.1]. 7-3-2008. Ref Type: Computer Program

100. OpenEye. OMEGA. [2.4.2]. 2010. Ref Type: Computer Program


112. Roth, B. L. and Driscoll, B. S. PDSP. 2011.

Ref Type: Computer Program


120. Shoichet Laboratory. AmpC Dataset. 2010. Ref Type: Data File


134. Thomsen, A. E.; Friedrichsen, G. M.; Sorensen, A. H.; Andersen, R.; Nielsen, C. U.; Brodin, B.; Begtrup, M.; Frokjaer, S.; and Steffansen, B. Prodrugs of purine and


Ref Type: Unpublished Work


