

Background

In the fields of public health and medicine, many studies are interested in analyzing a count of events over a time at-risk, or a rate. Examples of studies where events over a time period were the outcome variable of interest include bleeding frequencies in dogs with hemophilia B (Nichols 2013), lower respiratory infections in one year old infants (LaVange 1994), exacerbations in chronic obstructive pulmonary disease (COPD) (Keene 2008), and relapses in multiple sclerosis (Kappos 2006). The events are considered as discrete episodes over a time at risk; however, the difficulty in defining frequency of events varies by study. Generally, the risk time periods are variable and differ randomly between subjects. In some studies, the covariates in the study may change over the course of follow-up. This thesis will evaluate the different analysis methods that may be appropriate for data in the form of counts over a period of follow-up time. Considerations of these characteristics will affect the type of statistical methods used to analyze the data and in choosing the most appropriate method.

This honors thesis topic was motivated by Nichols et al. (2013). The intent of Nichols et al. was to design a new study to compare the extent to which a gene-therapy treatment, called tolerization, would reduce the frequency of bleeding events in a dog with hemophilia B over two years (Nichols 2013). Nichols et al. was based on the data collected by a completed study, Russell et al. The original study, Russell et al., was also interested in the extent to which the tolerized treatment would reduce the number of bleeding events in a dog each year (Nichols 2013). The data collected were bleeding frequency of hemophilia A dogs over time segments of 0-4 months, 0-1 year, 1-2 years, 2-3 years, and 3-3.5 years. Nineteen dogs from the same litter were randomly assigned to one of two groups, non-tolerized (control) or tolerized treatment (Nichols 2013). Dogs assigned to the tolerized group were treated prophylactically to a trough level of 1% human FIX using a gene therapy approach. If the dog passed the retention requirements, then the frequency of bleeding events and total time at-risk were recorded. Of the nine dogs assigned to the tolerized group, only five achieved tolerance to the human FIX; hence, only five dogs were followed for the study. For the purposes of this thesis, the bleeding frequency dataset will refer to the first two years of data collected in the original study by Russell et al.

An epidemiological follow-up study of lower respiratory illness (LRI) in children during the first year of life is another application of counted outcomes (LaVange 1994). This study was

interested in whether passive smoking exposure to tobacco smoke had an effect on incidence of LRI and whether this possible difference was due to other covariates. This large study compared two groups of infants: exposed to passive smoke and unexposed to passive smoking. The data were from a community-based cohort study of respiratory illness during the first year of life of 284 infants born in town counties in central North Carolina in 1986 and 1988 (LaVange 1994). A LRI event was defined as the presence of coughing, wheezing, or rattling in the chest as reported by the parents. Covariates included socioeconomic status, seasons, race, breast feeding, crowding in the home, and chronic respiratory symptoms at 12 months. In epidemiologic studies like these, incidence densities are used to compare incidences across subgroups. LaVange et al. (1994) considered the use of the direct sample survey method of ratio estimation to estimate the incidence densities and to model their variation, known as the density ratio method. The density ratio method requires a large sample size such that the sums comprising of the numerators and denominators of the ratios are approximately normal for the risk of groups of interest (LaVange 1994). Thus, the density ratio method is not applicable for evaluation on the bleeding frequency dataset. Instead, the LRI dataset in LaVange et al. (1994) was used to evaluate the density ratio method and model-based methods.

Though not directly used in this paper, COPD exacerbations (Keene 2008) and multiple sclerosis relapses (Kappos 2006) are notable examples of studies that utilized different methods to analyze counts over a period at risk. Specifically, the COPD study applied methods that directly compared rates of each treatment of recurrent events of COPD in a respiratory clinical trial (Keene 2008). The study TRISTAN was a year-long double-blind, randomized study comparing the effects of different treatments on COPD exacerbation rates of 1465 patients (Keene 2008). The data collected were the counted outcomes of acute exacerbations. Exacerbations were defined as worsening of COPD symptoms that required some type of treatment. The original analysis on the exacerbation rates used a Poisson regression model without correction for over-dispersion. However, the assumptions of a Poisson regression were not met for the data of COPD exacerbations (Keene 2008). Thus, a negative binomial model, which assumes a separate Poisson parameter for each subject, was expected to provide a better fit for the data (Keene 2008). The investigators also considered non-parametric methods, since non-parametric methods would also avoid the assumptions of Poisson regression. However, due to the discrete and bounded property of the data, it was determined that a non-parametric method

would not provide estimates of the treatment effects that represent the extent of risk reduction (Keene 2008).

As a final example, Kappos et al. aimed to evaluate the treatment effect of the new oral immunomodulating agent fingolimod (FTY720) on relapsing multiple sclerosis. The study randomly assigned 281 patients to three treatment groups: a daily intake of 1.25 mg of fingolimod, 0.5 mg of fingolimod, or placebo (Kappos 2006). Data collected were magnetic resonance imaging (MRI) and other clinical evaluations over 6 months. Wilcoxon rank sum tests were used to compare the MRI endpoints among the three groups. Poisson regression was also used to compare the annualized relapse rates. Investigators found that fingolimod reduced the number of lesions detected on MRI and clinical disease activity in patients with multiple sclerosis (Kappos 2006). Data from the Sylvia Lawry Centre for Multiple Sclerosis Research were used to calculate power and sample size using a non-parametric bootstrap method with a two-sided Wilcoxon rank-sum test and a 0.05 significance level (Kappos 2006).

This thesis will focus on evaluating four main methods: non-parametric method Wilcoxon rank sum test, model-based methods Poisson regression and negative binomial regression, and an alternative density ratio method. The Statistical Methods section compares general theory and concepts of these methods, as well as the advantages and disadvantages. In the Application section, Wilcoxon rank sum test, Poisson regression, and negative binomial regression will be applied and evaluated using the bleeding frequency dataset (Nichols 2013). Furthermore, the bleeding frequency data will be used to evaluate sample size calculations for the future Nichols et al. (2013) study using the Wilcoxon rank sum test and negative binomial regression with a 0.05 two-sided significance level. The density ratio method, Poisson regression, and negative binomial regression will be evaluated using the LRI dataset (LaVange 1994). The Appendix further elaborates the mathematical expressions of each method.

Statistical Methods

Non-parametric methods:

The most commonly used non-parametric method when interested in the difference between two independent random samples of subjects is the Wilcoxon rank sum test. The Wilcoxon rank sum test is used to test for equality of distributions for two groups versus a shift in their location, such as their medians. The application of the Wilcoxon rank sum test assumes

each group comes from one of two populations. Given the sample size of a simple random sample (SRS) from population 1 is n_1 and the sample size of a SRS from population 2 is n_2 , then let N be the sum of two sample sizes from both populations. Every observation in N are ranked as if all observations were from one large sample. The smallest observation is given a rank of 1 and the largest observation is given a rank of N . Ties between observations are given an average rank of the equal observations. The test statistic, W , is the sum of the ranks in the smaller sample n_1 . An exact p-value of W can be determined from an exact distribution, which assumes all possible assignments for ranks to groups are equally likely. If the sample size per group is greater than or equal to 10 subjects, the significance of W can also be approximated by a standard normal approximation for $Z_w = \frac{W - \mu_w}{\sigma_w}$.

Since the Wilcoxon rank sum test uses the ranks of the observations instead of the actual numerical data, no assumptions about the underlying distributions of the data are needed to make inferences. The only needed assumption is that the subjects are independent and identically distributed within groups and independent between groups. Another important advantage is that the Wilcoxon rank sum test uses the data equally from the respective patients. In other words, the data from patients who are enrolled in the study longer will not have more weight in the analysis. Furthermore, the Wilcoxon rank sum test is less sensitive to outliers. However, power in the Wilcoxon rank sum test can be adversely affected by other differences, such as shape and scale, between distributions instead of a shift of location between the two distributions. If the assumptions of a two-sample t-test are met, then the Wilcoxon rank sum test will have a lower power. In addition, measures of the difference between the groups may not be straightforward to obtain or interpret.

The Mann-Whitney estimator is a measure of difference generally calculated when a Wilcoxon rank sum test is used. It estimates the likelihood that the subjects in group 1 have a better outcome than subjects in group 2. In other words, if a_i is the outcome for a randomly selected subject from group 1 and b_j is the outcome for a randomly selected subject from group 2 (with n_1 and n_2 total subjects, respectively), then the Mann-Whitney estimator is calculated as the number of pairs where a_i has a better outcome than b_j divided by $(n_1 * n_2)$, given ties between pairs were randomly broken with probability $\frac{1}{2}$. A Mann-Whitney estimator closer to 1 implies larger ranks for group 1 than group 2 and thus favoring group 1. Alternatively, a Mann-

Whitney estimator closer to 0 favors group 2. If the Mann-Whitney estimator is around $\frac{1}{2}$, it indicates the distributions between the two groups are equal. The Mann-Whitney estimator is related to the ranks of the subjects through the relationship $\frac{1}{N}(\overline{R}_1 - \overline{R}_2)$, where $(\overline{R}_1 - \overline{R}_2)$ is the difference between the mean ranks of group 1 and group 2.

Somer's D is another measure of difference related to the Mann-Whitney estimator that also describes ordinal scaled data. It is related to the Mann-Whitney estimator through the equation: Somer's D = (2*Mann-Whitney) - 1. Consequently, the range of a Somer's D falls between -1 and 1, and thus the Somer's D acts like a correlation coefficient. Accordingly, a Somer's D near 1 implies larger ranks for and favoring group 1, whereas a Somer's D near -1 comparably implies larger ranks for and favoring group 2. If there is no difference between the two groups, then Somer's D will be about 0. The Somer's D is of interest, because it is readily produced in PROC FREQ in SAS, whereas the Mann-Whitney estimator is not.

The Hodges-Lehmann estimate, related to the Mann-Whitney estimator, also attempts to estimate the difference between the medians of two groups. The Hodges-Lehmann point estimate is the median of the difference between all possible pairs of subjects from group 1 with subjects from group 2. Ordering all differences from smallest to largest adjacent to corresponding quantiles of the distribution of the Mann-Whitney statistic identifies the confidence intervals of the Mann-Whitney statistic. The Hodges-Lehmann is of interest because it provides a method of obtaining a confidence interval that corresponds to the Wilcoxon Rank Sum test.

When designing a future study, it is possible to use a transformation of power of a Wilcoxon rank sum test to calculate the needed sample size. Through arithmetic and derivations detailed in the Appendix 1, $\text{Power} = (1 - \beta) = \Phi \left\{ \frac{(\theta - 0.5) - \frac{2}{N^2}}{\sqrt{\frac{N+1}{3N^2}}} - Z_{\frac{\alpha}{2}} \right\}$, where $\theta = E\{\text{Mann-Whitney estimator}\}$, $\text{Var}\{\text{Mann-Whitney estimator} | H_0\} = \frac{N+1}{3N^2}$, the continuity correction = $(\frac{2}{N^2})$, and Φ is cumulative density function of $N(0, 1)$. As detailed in Appendix 1, the sample size under the assumption $n_1 = n_2$ can be estimated by approximating $N+1$ and $(2/N^2)$ as N and 0, respectively. Thus, the sample size per group is approximated by $(Z_{\alpha/2} + Z_{\beta})^2 / 6(\theta - 0.5)^2$, where $Z_{\alpha/2}$ is the z-score given an two-sided alpha level, Z_{β} is the z-score at a desired power level, and θ is the Mann-Whitney statistic. The total sample size N would be two times the determined sample size per group. Also, it is worthy to note that if the calculated sample size is

used to re-calculate the power, the re-calculated power will be less than desired power because the continuity correction was not included in the estimate. Thus, usually an addition of one subject per group (or two subjects to the total sample size) is needed to produce the desired power and appropriate sample size.

Model-based Methods:

The most common and traditional approach to analyzing discrete counted outcome rates is Poisson regression, especially when dealing with rare events. Poisson regression is applied to generate a model fit to predict the count and comparison of groups given a set of predictors (Stokes 2012). It assumes that the counts of event are independent and have a Poisson distribution, in which the expected value and variance are equal. When subjects have different exposure times, the expected value and variance for count of events (y_i) for subject i is $\mu_i = \lambda_i * t_i$. The likelihood function for Poisson regression is $\prod_{i=1}^n \exp(-\lambda_i t_i) \frac{(\lambda_i t_i)^{y_i}}{y_i!}$ and the usual model for λ_i is loglinear. One advantage of Poisson regression is it can be applied to counts that do not have a limit on how large the count can be; for instance, when the variable does not have a known “denominator” or is not a proportion.

However, the Poisson method has many assumptions about the underlying distribution of the data which limits its applicability. The methodology assumes that the outcome variable has a Poisson distribution with the same Poisson parameter λ for all subjects. A common rate assumption for all subjects means the distribution of rates for each subject has the same mean for the same follow-up time. The biggest disadvantage of this assumption is the variability between patients is not satisfactorily accounted for (Kotz-Johnson 1986). It also does not account for correlation of events within a subject as a potential source of over-dispersion. Since a Poisson analysis weighs the unit of time equally, the model will not appropriately account for situations where subjects withdraw from the study earlier, assuming subjects who leave the study have a higher frequency of events than subjects who continue to the end of the study (Keene 2008). If the data do not follow these assumptions, the Poisson regression tends to under-estimate the underlying true rate (Stokes 2012).

When the observed variance is larger than the nominal variance for the assumed distribution, over-dispersion occurs (Stokes 2012). Over-dispersion is common in analysis of proportions or discrete counts because variances for binomial and Poisson distributions are fixed

by the mean, a single parameter (Stokes 2012). In regression, the ratio of the goodness-of-fit chi-square statistic versus degrees of freedom can indicate the presence of over-dispersion by exceeding 1. An over-dispersion correction attempts to account for variability between patients not explained by the Poisson regression by increasing the standard errors of the estimates. There are three main over-dispersion corrections; however, there is no universal consensus or method of determining which correction to use (Keene 2008). The simplest correction is multiplying the variance by a scaling factor. The scaling factor ϕ is the χ^2 statistic divided by its degrees of freedom (Stokes 2012). The covariance matrix will be pre-multiplied by the scaling factor, and the scaled deviance and log likelihoods will be divided by the scaling factor. The Poisson regression is then performed as usual with the scaled values. In the scaled Poisson regression, ϕ is equal to 1. Another over-dispersion correction is using generalized estimating equations (GEE) as described in Chapter 15 of Stokes et al. (2012). The GEE estimation comes from a variance estimation process that uses subject-to-subject measures instead of the model-based variance and involves aggregates at the subject level. Therefore, the GEE estimation method is more robust to misspecification of the covariance structure and can be applied by using PROC GENMOD in SAS (Stokes 2012). However, generally, the corrected Poisson regression is still not applicable because the variance of the dataset is much larger than assumed.

Negative binomial regression is a more appealing correction method, because it corrects for over-dispersion with a better model. The negative binomial regression model combines the idea behind Poisson regression with a model error that follows a gamma distribution (Keene 2008). The negative binomial regression assumes that each subject conditionally has their own underlying rate λ_i , which allows for a more flexible variance. Given an underlying rate (λ_i) and time at risk (t_i) for each subject i , the outcome variable y_i has a Poisson distribution with an expected value of $\lambda_i t_i$, where λ_i has a gamma distribution with parameters (α_i, ξ_i) . The expected value of λ_i is $\alpha_i \xi_i$ and the variance is $\alpha_i \xi_i^2$. Through explanations given in Appendix 2, the expected value of the outcome variable for every subject is μ_i and the variance is $\mu_i + k\mu_i^2$, where k is the negative binomial dispersion parameter (Stokes 2012). When $k=0$, the negative binomial distribution equals the Poisson distribution. Negative binomial regression more effectively accounts for increased rates among subjects who withdraw earlier, which is one of Poisson regression's biggest disadvantages. If over-dispersion is present, a negative binomial regression will produce more precise confidence intervals compared to a Poisson regression.

However, since the negative binomial regression still assumes that each subject has an underlying conditional Poisson distribution, it is not applicable to cases where the underlying distribution is not known.

For prospective studies, sample size can be determined based on the results in a negative binomial regression. The sample size per group can be determined through a relationship between the means for group 1 and group 2 (μ_1 and μ_2 , respectively) and the negative binomial dispersion value k (Keene 2007). The sample size per group is calculated as

$$n = \frac{(Z_\alpha + Z_\beta)^2 \left\{ \frac{1}{\mu_1} + \frac{1}{\mu_2} + 2k \right\}}{\{\log_e(\frac{\mu_1}{\mu_2})\}^2}$$
, where Z_α is the z-score given a two-sided alpha and Z_β is the z-score at a desired power level (Keene 2007). Similar to the variance parameter, if a sample size calculation based on a Poisson distribution was desired, then this equation can be used with $k=0$.

Density Ratio Method:

An alternative method that can be used to analyze counts over a risk time period is the density ratio method outlined in LaVange et al. (1994). An incidence density is defined as the number of new cases divided by the time at risk. The density ratio method is interested in estimating $\lambda = \mu_y/\mu_x$, the ratio of the two mean parameters for the variables of interest in the target population, by R with a random sample (LaVange 1994). Measures \bar{x} and \bar{y} estimate the population parameters μ_x and μ_y , and $R = (\bar{y}/\bar{x})$. The variance of R can be approximated by the variance of z through the relationship $\sum_{i=1}^n \frac{(z_i - \bar{z})^2}{n-1} = \sum_{i=1}^n \frac{(y_i - Rx_i)^2}{n(n-1)\bar{x}^2}$, where $z_i = \frac{(y_i - Rx_i)}{\bar{x}}$. Thus, the 95% confidence interval of R can be determined by $\exp\{\log_e R \pm 1.96\sqrt{(\vartheta_{\log_e R})}\}$, where $\vartheta_{\log_e R} = \text{Var}\{\log_e R\} = \frac{\text{Var}(z)}{R^2}$. The density ratio method provides a direct approach to analyzing disease incidence while adjusting for confounding. Like non-parametric methods, the density ratio method has minimal assumptions on the underlying distribution of the variables of interest. This method also provides a more convenient way to construct confidence intervals for estimated incidence densities and for comparison across groups (LaVange 1994). Unfortunately, the ratio method is only applicable when there is a large sample size, or that the sums for numerators and denominators of ratios are approximately normal for the groups of interest (LaVange 1994). This method is also less flexible in the number and types of covariates or confounders that can be accommodated (LaVange 1994).

Application of Methods

Bleeding Frequency dataset (Nichols 2013):

A rate of bleeds per year over the first two years was computed for each dog by dividing the total number of bleeds in the first two years by the total years of follow-up time. The total years of follow-up time was not the same for all dogs, because some were euthanized or died before the end of two years. Table 1 and Table 2 summarize the data used to design the new study (Nichols et al.) and descriptive statistics of the data respectively.

Table 1: Bleeding frequency raw dataset (Nichols et al.)

ID	Group	Sex	Bleeds in Year 0-1	Bleeds in Year 1-2	Total bleeds (Year 0-2)	Total years of follow-up	Rate of bleeds per year (over total follow-up time)
X02	Non-tolerized	M	3	9	12	2	6
X06	Non-tolerized	M	8	21	29	2	14.5
Y21	Non-tolerized	M	3	8	11	2	5.5
E19	Non-tolerized	F	13	9	22	1.38	15.9
Z100	Non-tolerized	F	7	2	9	2	4.5
Z58	Non-tolerized	F	4	2	6	1.27	4.7
Y24	Non-tolerized	F	7	1	8	1.27	6.3
Z91	Non-tolerized	M	3	4	7	2	3.5
E30	Non-tolerized	M	3	0	3	2	1.5
C20	Non-tolerized	M	4	2	6	2	3.0
C22	Tolerized	M	2	0	2	2	1.0
C23	Tolerized	M	2	0	2	2	1.0
C25	Tolerized	F	2	1	3	2	1.5
C26	Tolerized	F	2	5	7	2	3.5
X02	Tolerized	F	1	2	3	2	1.5

Table 2: Descriptive statistics of outcome variables total bleeds in first year and rate of bleeds per year over total follow-up time of bleeding frequency dataset (Nichols 2013)

Group	Frequency of bleeds in first year (YR 0-1)	Rate of bleeds per year (over total follow-up time)
Non-tolerized		
Mean	5.50	6.55
Standard dev.	3.27	4.80
Tolerized		
Mean	1.80	1.70
Standard dev.	0.45	1.04

Non-parametric Methods

The small sample size in the original study and new study lends itself well to non-parametric methods, such as the Wilcoxon rank sum test. The Wilcoxon rank sum test was applied to the outcome variables of frequency of bleeds in the first year and rate of bleeds per year over total follow-up time. Measures of differences produced for the two outcome variables included Somer's D, Mann-Whitney Estimator, and Hodges-Lehmann estimate. The Somer's D was determined through the MEASURES option of PROC FREQ in SAS 9.3 and the Mann-Whitney statistic was manually calculated using the relationship $\frac{\text{Somer's } D + 1}{2}$ from the SAS produced Somer's D. Similarly, the standard error of the Mann-Whitney statistic was calculated as the standard error of the Somer's D value divided by 2. Due to the small sample size of bleeding frequency dataset, a normal approximation of the significance of W in the Wilcoxon rank sum test may not be appropriate. Therefore, an exact Wilcoxon rank sum test was also computed for bleeding frequency dataset. The non-parametric results are summarized in Table 3.

Table 3. Non-parametric results for outcome variables frequency of bleeds in Year 0-1 and rate of bleeds per year (over total follow-up time) in bleeding frequency dataset (Nichols 2013)

Outcome variable	Frequency of bleeds in Year 0-1	Rate of bleeds per year (over two years)
Wilcoxon statistic	15.00	18.50
P-value (approx.)	0.0085	0.0216
P-value (exact)	< 0.0003	0.0060
Hodges-Lehmann	-2.0	-3.5
95%Confidence Interval	(-6.0, -1.0)	(-11.0, -1.5)
Somer's D (SE)	- 1.00 (0.00)	- 0.86 (0.12)
Mann-Whitney Estimator (SE)	0.00 (0.00)	0.07 (0.06)

For both outcome variables, the Wilcoxon rank sum test produced a significant p-value at the 0.05 significance level, indicating that the two distributions between the non-tolerized group and tolerized group were not equal. In other words, the two distributions are shifted in location. Furthermore, for both outcome variables, the Somer's D was near -1, the Mann-Whitney estimator was near 0, and the Hodges-Lehmann confidence interval did not contain 0, thus indicating the difference strongly favors the tolerized group with less bleeds. In other words, a dog in a non-tolerized group has a higher likelihood of having more bleeds than dog in tolerized group. Conversely, a tolerized treated dog will have a higher likelihood of having less bleeds

than a non-tolerized dog. This supports the hypothesis that tolerized treatment can reduce the number of bleeds a hemophilia B dog has.

As mentioned earlier, Nichols et al. (2013) intended to design a new study based on the first two years of the data collected by Russell et al. Thus, sample size was calculated for the future Nichols et al. (2013) study based on the Wilcoxon rank sum test results of the bleeding frequency dataset. Assuming that the number of subjects in each group are equal, the sample size needed for 0.90 power was calculated for different Mann-Whitney estimators around 0.07 (the Mann-Whitney statistic for bleeds per year over total follow-up time). Table 4 summarizes the sample size needed for Mann-Whitney estimators around 0.07 for 0.90 power and a two-sided alpha 0.05. In order to match the data for the formulas and methods described in Appendix 1, the reverse ranks were implicitly used. This is feasible as ranks are symmetric; consequently, a theta of 0.07 corresponds to a theta of 0.93 in the power calculation. The power of the test was also calculated using the previously calculated sample sizes in Table 4 to re-confirm 0.90 power . As expected, the power was lower than the intended 0.90. One subject was added to each group and the power calculation was repeated to produce powers above 0.90. Table 5 displays the power calculations with the corrected sample size of an additional subject per group at a two-sided alpha of 0.05.

Table 4. Calculated Sample Size via Wilcoxon rank sum test for 0.90 power and two-sided alpha of 0.05

$\theta(1 - \theta)$	$n_1 = n_2$	N	Power
0.05 (0.95)	9	18	0.888
0.075 (0.925)	10	20	0.888
0.1 (0.90)	11	22	0.882
0.125 (0.875)	13	26	0.897

Table 5. Re-calculated Power for the Wilcoxon rank sum test after adding one subject per group.

$\theta(1 - \theta)$	N	Power
0.05 (0.95)	20	0.920
0.075 (0.925)	22	0.917
0.10 (0.90)	24	0.909
0.125 (0.875)	28	0.918

Model-based Methods

For comparison to the non-parametric results, model-based methods were also applied to the bleeding frequency dataset. Poisson regression, and over-dispersion corrections scaled Poisson regression and negative binomial regression were computed on the outcome variables total bleeds in the first year of observation and rate of bleeds per year (over total follow-up time). Table 6 and Table 7 display the results from these three model-based methods for each of the two outcome variables. The x -variable is the treatment group the dog was assigned to and y is the outcome variable.

Table 6. Model-based method results for outcome variable frequency of bleeds in first year

Test	Poisson	Scaled Poisson	Negative binomial
$\phi[Q_p/(DF = 13)]$	2.22	1.00	1.19
Dispersion	----	----	0.16
LR Statistic	15.28	6.89	8.50
p-value	<0.0001	0.0087	0.0036
Model	$\log_e(y) = -1.23x + 1.13$	$\log_e(y) = -1.23x + 1.13$	$\log_e(y) = -1.27x + 1.16$
95%CI for μ_T/μ_N	(0.14, 0.59)	(0.10, 0.84)	(0.12, 0.64)

Table 7. Model-based results for outcome variable rate of bleeds per year (over two years)

Test	Poisson	Scaled Poisson	Negative binomial
$\phi[Q_p/(DF = 13)]$	4.74	1.00	1.24
Dispersion	----	---	0.27
LR Statistic	34.29	7.24	9.15
p-value	< 0.0001	0.0071	0.0025
Model	$\log_e(y) = -1.31x + 1.84$	$\log_e(y) = -1.31x + 1.84$	$\log_e(y) = -1.34x + 1.87$
95%CI for μ_T/μ_N	(0.16, 0.45)	(0.09, 0.82)	(0.12, 0.56)

When Poisson regression was applied, $\phi = Q_p/(DF = 13)$ was 2.22 and 4.74 for the two outcome variables respectively. This strongly indicates the presence of over-dispersion and therefore, an unadjusted Poisson regression is not an appropriate test to perform. After scaling the Poisson regression, ϕ was equal to 1 which indicated correction for over-dispersion. For both outcome variables, the 95% confidence interval for μ_T/μ_N was narrower in the Poisson regression than in the scaled Poisson, as expected. The negative binomial regression had ϕ of 1.19 and 1.24 for total bleeds in the first year and rate of bleeds per year (over two years),

respectively, also indicating correction for over-dispersion. The 95% confidence interval for the negative binomial is narrower than the scaled Poisson, indicating that it is a more appropriate model for the data.

Sample size via the negative binomial model can also be calculated using the estimated means of group 1 and group 2. Since the observed mean rate of bleeds per year (over total follow-up time) for the non-tolerized dogs was 6.55, the observed mean rate of bleeds per year (over total follow-up time) for the tolerized dogs was 1.70, and the dispersion factor was 0.27, conservative estimates of 6.0, 2.0, and 0.33 were used. The calculated sample size per group for Nichols et al. (2013) via the negative binomial model was 12 per group.

LRI dataset (LaVange 1994) :

For this paper, explanatory variable passive smoking (one or more smokers in the household) and covariate crowding in the household were of interest. The time at-risk was converted from weeks to years for the analysis. Crowding was defined as present if there was greater than or equal to 0.5 persons per room in a household; conversely, if there was less than 0.5 persons per room then there was no crowding. The main effect of interest was the effect of passive smoking adjusted for crowding. Additional analysis of the effect of passive smoking at both levels of crowding (present and absent) and the unadjusted effect of passive smoking on the count of LRI events were also of interest.

Model-based Methods

Model based methods, such as Poisson regression and negative binomial regression, were performed on all outcomes. When Poisson regression was applied in the four models, the deviance divided by degrees of freedom and the Pearson Chi-Square statistic divided by degrees of freedom were much greater than one, which indicated over-dispersion in the data. Consequently, negative binomial regression was used to correct the over-dispersion. For all models, the likelihood ratio (LR) statistics for Type 3 analysis showed passive smoking, and crowding when applicable, were statistically significant. The estimate of the parameter corresponding to passive smoking will be used to compare the incidence density ratios calculated below. Tables 8 to 11 summarize the Poisson regression and negative binomial regression models for all four outcomes. The parameter estimate for passive smoking is of interest for comparison between methods.

Table 8a. Poisson regression model of counts of LRI events adjusted for effect of passive smoking which was adjusted for crowding

Analysis Of Maximum Likelihood Parameter Estimates							
Parameter	DF	Estimate	Standard Error	Wald 95% Confidence Limits		Wald Chi-Square	Pr > ChiSq
Intercept	1	-0.5936	0.1418	-0.8716	-0.3157	17.52	<.0001
passive	1	0.5407	0.1533	0.2403	0.8411	12.44	0.0004
crowding	1	0.6214	0.1489	0.3295	0.9133	17.41	<.0001

Table 8b. Negative binomial regression model of counts of LRI events adjusted for effect of passive smoking which was adjusted for crowding

Analysis Of Maximum Likelihood Parameter Estimates							
Parameter	DF	Estimate	Standard Error	Wald 0.95onfidence Limits		Wald Chi-Square	Pr > ChiSq
Intercept	1	-0.562	0.176	-0.907	-0.216	10.15	0.0014
passive	1	0.570	0.200	0.178	0.963	8.10	0.0044
crowding	1	0.608	0.197	0.223	0.994	9.55	0.0020
Dispersion	1	1.035	0.268	0.623	1.718		

Table 9a. Poisson regression model of counts of LRI events adjusted for effect of passive smoking where crowding is not present (i.e. crowding=0)

Analysis Of Maximum Likelihood Parameter Estimates							
Parameter	DF	Estimate	Standard Error	Wald 95% Confidence Limits		Wald Chi-Square	Pr > ChiSq
Intercept	1	-0.641	0.174	-0.982	-0.300	13.55	0.0002
passive	1	0.625	0.230	0.173	1.076	7.36	0.0067

Table 9b. Negative binomial regression model of counts of LRI events adjusted for effect of passive smoking where crowding is not present (i.e. crowding=0)

Analysis Of Maximum Likelihood Parameter Estimates							
Parameter	DF	Estimate	Standard Error	Wald 95% Confidence Limits		Wald Chi-Square	Pr > ChiSq
Intercept	1	-0.581	0.230	-1.031	-0.132	6.42	0.0113
passive	1	0.656	0.324	0.022	1.290	4.11	0.0427
Dispersion	1	1.864	0.634	0.957	3.631		

Table 10a. Poisson regression model of counts of LRI events adjusted for effect of passive smoking where crowding is present (i.e. crowding=1)

Analysis Of Maximum Likelihood Parameter Estimates							
Parameter	DF	Estimate	Standard Error	Wald 95% Confidence Limits		Wald Chi-Square	Pr > ChiSq
Intercept	1	0.076	0.172	-0.260	0.412	0.20	0.6597
passive	1	0.474	0.203	0.076	0.872	5.44	0.0207

Table 10b. Negative binomial regression model of counts of LRI events adjusted for effect of passive smoking where crowding is present (i.e. crowding=1)

Analysis Of Maximum Likelihood Parameter Estimates							
Parameter	DF	Estimate	Standard Error	Wald 95% Confidence Limits		Wald Chi-Square	Pr > ChiSq
Intercept	1	0.090	0.212	-0.325	0.505	0.18	0.6713
passive	1	0.494	0.257	-0.010	0.997	3.69	0.0546
Dispersion	1	0.678	0.257	0.322	1.425		

Table 11a. Poisson regression model of counts of LRI events adjusted for effect of passive smoking

Analysis Of Maximum Likelihood Parameter Estimates							
Parameter	DF	Estimate	Standard Error	Wald 95% Confidence Limits		Wald Chi-Square	Pr > ChiSq
Intercept	1	-0.340	0.122	-0.580	-0.101	7.76	0.0054
passive	1	0.658	0.150	0.362	0.953	19.02	<.0001

Table 11b. Negative binomial regression model of counts of LRI events adjusted for effect of passive smoking

Analysis Of Maximum Likelihood Parameter Estimates							
Parameter	DF	Estimate	Standard Error	Wald 95% Confidence Limits		Wald Chi-Square	Pr > ChiSq
Intercept	1	-0.302	0.156	-0.608	0.004	3.75	0.0527
passive	1	0.680	0.202	0.284	1.075	11.32	0.0008
Dispersion	1	1.194	0.288	0.745	1.914		

Density Ratio Method

The incidence density ratio method described in LaVange et al. (1994) was used to determine the density ratios for the four different outcomes. The average count (\bar{y}) and average time at-risk (\bar{x}) were determined through PROC MEANS in SAS to produce an estimate of R for each model by exposure to passive smoking and no exposure to passive smoking. The variable $z_i = \frac{(y_i - Rx_i)}{\bar{x}}$ was computed, and the relationship between the variance of $\log_e R$ and the variance of z was calculated to determine the 95% confidence intervals for λ in both the exposed group and the non-exposed group. The incidence density ratios, on the natural log scale, were then calculated by $R_{exposed}/R_{nonexposed}$. The 95% confidence intervals for the incidence density ratios were calculated by

$$\exp \left\{ \log_e (R_{exposed}/R_{nonexposed}) \pm 1.96 \sqrt{\vartheta_{\log_e R_{exposed}} + \vartheta_{\log_e R_{non-exposed}}} \right\}.$$

Table 12 summarizes the results obtained from the density ratio method and the model-based methods. All results in Table 12 have been converted from the natural log (\log_e) scale.

Table 12. Comparison between results from density ratio method and model-based methods (Poisson regression and Negative binomial regression)

Method	Effect of passive smoking, adjusted for crowding	Effect of Passive smoking	Effect of Passive smoking, where crowding = 0	Effect of Passive smoking, where crowding = 1
Density Ratio				
Estimate	1.70	1.93	1.87	1.61
95%CI	(1.17, 2.47)	(1.33, 2.81)	(1.01, 3.44)	(1.00, 2.58)
p-value	0.0054	0.0006	0.0454	0.0496
Poisson (unadjusted)				
Estimate	1.72	1.93	1.87	1.61
95%CI	(1.27, 2.32)	(1.44, 2.59)	(1.19, 2.93)	(1.08, 2.39)
p-value	<0.0001	< 0.0001	0.0063	0.0163
Negative Binomial				
Estimate	1.77	1.97	1.93	1.64
95%CI	(1.19, 2.62)	(1.33, 2.93)	(1.02, 3.63)	(0.99, 2.71)
p-value	0.0046	0.0008	0.0438	0.0545

Discussion

This thesis evaluated different statistical methods for comparing counts of events for at-risk time periods across two groups, such as the Wilcoxon rank sum test, Poisson regression, negative binomial regression, and the density ratio method. The bleeding frequency dataset (Nichols 2013) was used to evaluate the first three methods. The primary outcome of interest was the rate of bleeds per year (over total follow-up time) and the secondary outcome of interest was the frequency of bleeds in the first year. The mean and the standard deviations of the mean bleeds for both outcome variables suggested that tolerized treatment reduced the frequency of bleeds in dogs with hemophilia B. Due to the small sample size of the bleeding frequency dataset, applying the Wilcoxon rank sum test was of particular interest to test the inference suggested by the means and standard deviations, because it does not have any underlying distribution assumptions. For both outcome variables, the Wilcoxon rank sum test had a significant two-sided p-value (0.0003 and 0.0060, respectively). Thus, there is sufficient evidence to suggest that there is a difference between the distribution of bleeds between the non-tolerized dogs and tolerized

dogs for both outcomes. The Somer's D, Mann-Whitney estimator, and Hodges-Lehmann confidence interval were also in agreement with the Wilcoxon rank sum test results. For both outcome variables, Somer's D was close to -1, the Mann-Whitney estimator was close to 0, and the Hodges-Lehmann confidence interval did not contain 0, indicating the comparisons between randomly selected subjects in the non-tolerized group and randomly selected subjects in the tolerized group strongly favor the tolerized group. In other words, the dogs in the non-tolerized group had a higher likelihood of having more bleeds than dogs in the tolerized group. This supports the notion the tolerized treatment reduces the likelihood of having a bleed in hemophilia B dogs.

Generally, when data are in the form of counts over a period of time at-risk, the model-based method of Poisson regression is used because it is easily applied to this type of data. However, the Poisson regression model assumes that there is a common rate for all subjects, and thus it will under-estimate the variance of the parameter if this assumption is not met. To correct for over-dispersion in the bleeding frequency dataset (Nichols 2013), scaled Poisson regression and negative binomial regression were applied.

These three model-based methods produce similar models. As expected, the Poisson regression model incorrectly had the narrowest 95% confidence interval since over-dispersion was a factor in the bleeding frequency dataset. The 95% confidence interval for the negative binomial regression model was narrower than the scaled Poisson regression model for both outcome variables (respectively, (0.10, 0.84) versus (0.12, 0.64) for frequency of bleeds in first year, and (0.09, 0.82) versus (0.12, 0.56) for rate of bleeds per year over two years of follow-up). This indicates that the scaled Poisson model, which intended to correct for over-dispersion, over-estimated the variance. Consequently, the negative binomial regression model is a more appealing option than other corrections because it corrects for over-dispersion in a Poisson regression with a better model. However, since it still assumes that each subject has an underlying conditional Poisson distribution, it is not appropriate to use in the case of a small sample size (less than or equal to 10 subjects per group) or any time that assumption is not met.

Sample size for a future study based on the results of a Wilcoxon rank sum test and a negative binomial regression were calculated. Assuming that there will be an equal number of subjects in the two groups and 0.90 power is desired, the sample size via the negative binomial regression model was 12 per group. Using a conservative Mann-Whitney statistic, the sample

size via the Wilcoxon rank sum method was approximately 11 subjects per group. Since the sample size approximation assumed the continuity correction was about 0, the adjusted sample size per group to achieve the desired power produced a sample size of 12 subjects per group. The sample sizes calculated from both methods agree with each other.

The density ratio method provides an additional method that, like the Wilcoxon rank sum method, has minimal assumptions for the underlying distributions. Furthermore, it provides a direct way to analyze disease incidence. Unfortunately, the density ratio method requires large sample size; therefore, it is not applicable to the bleeding frequency dataset. Instead, the dataset of lower respiratory incidence (LRI) in infants during their first year from LaVange et al. (1994) was used. The main effect of interest was passive smoking adjusted for crowding, though the effect of passive smoking on LRI and the effect of passive smoking by crowding status (i.e., the presence of crowding and no crowding) were also analyzed.

Poisson regression and negative binomial regression were also applied as model-based methods. Over-dispersion was present in the LRI dataset, and thus negative binomial regression was more appropriate than Poisson regression. As expected, for all four models, the 95% confidence interval from Poisson regression was inappropriately narrower than from the negative binomial model. The estimated standard errors for the parameters in all models were higher in the negative binomial regression than in Poisson regression.

When the density ratio method was applied to estimate the four effects, the estimates were more similar to the Poisson regression models. However, the 95% confidence intervals were wider than the Poisson models. Instead, they were more similar to the negative binomial 95% confidence intervals. A narrower 95% confidence interval than the negative binomial regression model indicates that the density ratio method adequately adjusts for the over-dispersion, but is a more precise method than negative binomial regression.

From these statistical considerations of different methods that deal with data of counts over a time at-risk, it can be concluded that even though Poisson regression is most readily applied to this type of data, it may not be the most appropriate model. When its distributional assumptions are not met, Poisson regressions will inappropriately under-estimate the variance. In the case of over-dispersion, the correction scaled Poisson regression may over-correct and thus over-estimate the variance. Negative binomial regression provides a more appealing method for correcting over-dispersion in a Poisson regression and produces more precise estimates.

However, in datasets where the assumption that every subject has an underlying Poisson regression is not met, the negative binomial model is also not the most appropriate method.

In cases where assumptions about the underlying distribution are not met, the non-parametric Wilcoxon rank sum test and alternative density ratio method are more appropriate to apply than the model-based methods mentioned above. When the sample size is small, the non-parametric Wilcoxon rank sum test is most commonly used. However, the power of a Wilcoxon rank sum test can be adversely affected by other differences, such as shape and scale, instead of just location. Furthermore, measures of differences for groups, such as Hodges-Lehmann and Mann-Whitney estimators, may not be straightforward to obtain or interpret. The density ratio method, on the other hand, cannot be used with small datasets and it is not as flexible in regard to the number and types of covariates allowed in the model. However, with large sample sizes, the density ratio method provides a direct method to evaluate incidence densities and produces more precise estimates than the model-based methods. These considerations and other aspects are important to evaluate because it will inform use of the most appropriate method to produce valid estimates and inferences from the data.

References

- LaVange L., Keyes L., Koch G., and Margolis P. Application of Sample Survey Methods for Modelling Ratios to incidence Densities. *Statistics in Medicine*, 1994; 13: 343-355.
- LaVange L., and Koch G. Rank Score Tests. *Circulation*, 2006; 114(23):2528-33.
- Kappos L., Antel J., Comi G., Montalban X., O'Connor P., Polman C., Haas T., Korn A., Karlsson G., and Radue E., Oral Fingolimod (FTY720) for Relapsing Multiple Sclerosis. *The New England Journal of Medicine*, 2006; 355: 1124-1140
- Keene O., Jones M., Lane P., and Anderson J. Analysis of exacerbation rates in asthma and chronic obstructive pulmonary disease: example from TRISTAN study. *Pharmaceutical Statistics*, 2007; 6: 89-97.
- Keene O., Calverley P., Jones P., Vesbo J., and Anderson J. Statistical analysis of exacerbation rates in COPD: TRISTAN and ISOLDE revisited. *European Respiratory Journal*, 2008; 32: 17-24

- Koch G. and Stokes M. Stokes. Some Nonparametric and Regression Model Strategies for Counts of Primary Events from Randomized Studies. *International Journal of Statistical Sciences*, 2006; 5
- Kotz-Johnson. Poisson Regression. *Encyclopedia of Statistical Sciences*, 1986; 7
- Nichols T., Bellinger D., and Raymer R. Version 4: Effect of AAV8-mediated expression of canine FVIIIa on bleeding frequency of hemophilia A dogs. 2013
- Saville B., LaVange L., and Koch G. Estimating Covariate-Adjusted Incidence Density Ratios for Multiple Time Intervals in Clinical Trials Using Nonparametric Randomization-based ANCOVA. *Statistics in Biopharmaceutical Research*, 2011; 3 (2).
- Stokes M., Davis C., and Koch G. Categorical Data Analysis Using SAS Third Edition. : 2012, Chapter 5, 12, 15.

Appendix 1: Sample Size via Wilcoxon rank sum method

We will use a transformation of the power definition to calculate the needed sample size. The following calculations are for the normal approximation of the Wilcoxon rank sum test.

For the Wilcoxon rank sum test, $d_R = (\bar{R}_1 - \bar{R}_2)$ is the difference between mean ranks for group 1 and group 2.

The null hypothesis, H_0 , is the distributions for Group 1 and Group 2 are equal. Let $N = (n_1 + n_2)$, then

$$\begin{aligned} E\{(\bar{R}_1 - \bar{R}_2)|H_0\} &= 0 \text{ and} \\ \text{Var}\{(\bar{R}_1 - \bar{R}_2)|H_0, \text{no ties}\} &= \left(\frac{1}{n_1} + \frac{1}{n_2}\right) \frac{\sum_{k=1}^N (k - (N+1)/2)^2}{(N-1)} \\ &= \frac{N N(N+1)}{n_1 n_2 12} \end{aligned}$$

If $n_1 = n_2 = \left(\frac{N}{2}\right)$, then $\text{Var}\{(\bar{R}_1 - \bar{R}_2)|H_0\} = \frac{N+1}{3}$.

We know that $\left\{\left(\frac{U}{n_1 n_2}\right) - 0.5\right\} = \frac{\bar{R}_1 - \bar{R}_2}{N}$, where $\frac{U}{n_1 n_2}$ is the Mann-Whitney Statistic.

If $n_1 = n_2 = \frac{N}{2}$, then $\text{Var}\left\{\left(\frac{U}{n_1 n_2}\right) \middle| H_0\right\} = \frac{N+1}{3N^2}$

The alternative hypothesis, H_A , is that the two distributions are not equal. In other words,

$$H_A: E\left\{\left(\frac{U}{n_1 n_2}\right)\right\} = \theta, \text{ where } \theta > 0.5.$$

$$Power = (1 - \beta) = Pr \left\{ \frac{\frac{(R_1 - R_2)}{N} - \frac{1}{2} \left(\frac{1}{n_1} + \frac{1}{n_2} \right)}{\sqrt{\frac{N+1}{3N^2}}} > Z_{\alpha} \middle| H_A \right\}, \text{ where } \frac{1}{2} \left(\frac{1}{n_1} + \frac{1}{n_2} \right) \text{ is the continuity}$$

correction.

$$\text{If } n_1 = n_2 = \frac{N}{2}, \text{ then } Power = 1 - \beta \equiv Pr \left\{ \frac{U - \theta}{\sqrt{(N+1)/3N^2}} > Z_{\frac{\alpha}{2}} - \frac{(\theta - 0.5) - \frac{2}{N^2}}{\sqrt{(N+1)/3N^2}} \middle| H_A \right\}$$

Φ is cumulative density function of $N(0,1)$, then

$$1 - \beta = \Phi \left\{ \frac{(\theta - 0.5) - \frac{2}{N^2}}{\sqrt{\frac{N+1}{3N^2}}} - Z_{\alpha/2} \right\} \quad (1)$$

where $\theta = E\{\text{Mann-Whitney estimator}\}$ and $\left(\frac{2}{N^2}\right)$ is the continuity correction.

$$(Z_{\alpha/2} + Z_{\beta})^2 \left(\frac{N+1}{3N^2} \right) = \left\{ (\theta - 0.5) - \frac{2}{N^2} \right\}^2$$

Since $N + 1 \cong N$ and $\left(\frac{2}{N^2}\right) \cong 0$, $(Z_{\alpha/2} + Z_{\beta})^2 \left(\frac{1}{3N}\right) = (\theta - 0.5)^2$

$$\rightarrow N = n_1 + n_2 = \frac{\left(\frac{Z_{\alpha} + Z_{\beta}}{2} \right)^2}{3(\theta - 0.5)^2}$$

$$\rightarrow n_1 = n_2 = \frac{\left(\frac{Z_{\alpha} + Z_{\beta}}{2} \right)^2}{6(\theta - 0.5)^2}$$

To determine a sample size needed for 0.90 power at two-sided $\alpha = 0.05$, $Z_{\alpha/2} = 1.96$, $Z_{\beta} = 1.282$.

The power for a given calculated sample size N can be verified with (1). It is expected that the power calculated with calculated N will be less than the power desired so one subject per group is added.

Appendix 2: Poisson Regression and Negative Binomial Regression

Subjects are assumed to represent a population comparable, in a sense, to a random sample. Let y denote the count of events and t denote time at-risk.

Let $k = 1, 2$ denote two groups of subjects and $i = 1, 2, \dots, n_i$ denote subjects in the k^{th} group.

Poisson regression:

For $i = 1, 2, \dots, n$, let y_i be the count of events for subject i and t_i be the exposure (time at-risk) for subject i

Poisson regression assumes that each subject's respective y_i has an independent Poisson distribution. Thus,

$$E(y_i) = \text{Var}(y_i) = \mu_i = \lambda_i * t_i.$$

The likelihood function is $\prod_{i=1}^n \exp(-\lambda_i t_i) \frac{(\lambda_i t_i)^{y_i}}{y_i!}$.

The usual model for the λ_i is loglinear with $\lambda_i = \exp(x_i' * \beta)$ with x_i' as the vector of explanatory variables and β as the vector of unknown parameters. β is estimated by the maximum likelihood.

When $\lambda_i = \lambda$ for all i , $\hat{\lambda} = \sum y_i / \sum t_i$, thus

$$\begin{aligned} \text{Var}(\hat{\lambda}) &= \left(\lambda / \sum t_i \right) \\ &\cong \left(\hat{\lambda} / \sum t_i \right) \\ &= \sum y_i / (\sum t_i)^2 = \vartheta_{\hat{\lambda}} \end{aligned}$$

Negative Binomial:

A negative binomial model combines Poisson regression with a model error that follows a gamma distribution.

For $i = 1, 2, \dots, n$, y_i has the Poisson distribution with a given λ_i and t_i , where t_i is the fixed duration of follow-up and λ_i has gamma distribution with parameters (α_i, ξ_i) . Thus,

$$E(y_i) = \lambda_i t_i,$$

Furthermore,

$$E(\lambda_i) = \alpha_i \xi_i \text{ and } \text{Var}(\lambda_i) = \alpha_i \xi_i^2.$$

The usual model for negative binomial regression is $\exp(\mu_i) = x_i' * \beta$ with offset $\log_e(t_i)$, where x_i' is the vector of explanatory variables and β is the vector of unknown parameters estimated by maximum likelihood.

Assume $\alpha_i = \alpha$ for all patients, then λ_i has density function

$$f(\lambda_i) = \lambda_i^{\alpha-1} \frac{\exp\left(-\frac{\lambda_i}{\xi_i}\right)}{\xi_i^\alpha \Gamma(\alpha)} \quad \text{for } \lambda_i > 0.$$

Then,

$$\begin{aligned} E(y_i|\lambda_i) &= \lambda_i t_i \text{ and } E(y_i) = E(E(y_i|\lambda_i)) \\ &= E(\lambda_i t_i) \\ &= \alpha \xi_i t_i = \mu_i. \end{aligned}$$

and

$$\begin{aligned} \text{Var}(y_i) &= \text{Var}\{E(y_i|\lambda_i)\} + E\{\text{Var}(y_i|\lambda_i)\} \\ &= \text{Var}\{\lambda_i t_i\} + E\{\lambda_i t_i\} \\ &= \alpha \xi_i^2 t_i^2 + \alpha \xi_i t_i \\ &= \frac{\mu_i^2}{\alpha} + \mu_i \\ &= \mu_i + k \mu_i^2, \end{aligned}$$

where k is the negative binomial dispersion factor.

For fixed λ , as α gets large (or as k approaches 0), the negative binomial distribution converges to a Poisson distribution