NF-κB–Induced Loss of MyoD Messenger RNA: Possible Role in Muscle Decay and Cachexia

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MyoD regulates skeletal muscle differentiation (SMD) and is essential for repair of damaged tissue. The transcription factor nuclear factor kappa B (NF-κB) is activated by the cytokine tumor necrosis factor (TNF), a mediator of skeletal muscle wasting in cachexia. Here, the role of NF-κB in cytokine-induced muscle degeneration was explored. In differentiating C2C12 myocytes, TNF-induced activation of NF-κB inhibited SMD by suppressing MyoD mRNA at the post-transcriptional level. In contrast, in differentiated myotubes, TNF plus interferon-γ (IFN-γ) signaling was required for NF-κB–dependent down-regulation of MyoD and dysfunction of skeletal myofibers. MyoD mRNA was also down-regulated by TNF and IFN-γ expression in mouse muscle in vivo. These data elucidate a possible mechanism that may underlie the skeletal muscle decay in cachexia.

The vertebrate skeletal muscle differentiation (SMD) program is under the strict control of the myogenic bHLH transcription factor family (MyoD, Myf5, myogenin, and MRF4) and of a second class of transcription factors termed myocyte enhancer factor–2 (MEF2A) (1, 2). MyoD and Myf5 are expressed in proliferating undifferentiated myoblasts and, upon growth factor withdrawal, are activated to initiate SMD that ultimately leads to the fusion of myoblasts into multinucleated myotubes (1, 2). Although mice lacking MyoD develop normally (3), MyoD-deficient skeletal muscle is severely impaired in its ability to regenerate after tissue injury, which suggests a specific role for MyoD in the replenishment of lost muscle (4).

NF-κB is a transcription factor expressed in a variety of cell types, including mature muscle (5), and it inhibits SMD by regulating the expression of cyclin D1 (6). The cytokine tumor necrosis factor (TNF) is a potent activator of NF-κB, causing it to translocate to the nucleus and bind to promoters and enhancers of genes involved in inflammatory and proliferative responses (7–9). TNF is an important mediator of skeletal muscle degeneration associated with cachexia, a debilitating syndrome characterized by extreme weight loss and whole-body wasting (10–12). Cachexia is frequently seen in patients afflicted with chronic diseases such as cancer and acquired immunodeficiency syndrome. It is striking that about one-third of cancer mortalities result from cachexia rather than tumor burden (13). Little is known about the molecular etiology of cachexia, and hence few targets have been identified for therapy. Here we investigate the potential role of NF-κB as a downstream effector of TNF–mediated skeletal muscle dysfunction.

TNF was recently shown to inhibit skeletal myogenesis in vitro (6, 14). We therefore investigated the potential role of NF-κB in this regulatory process. The addition of TNF to mouse C2C12 myocytes completely blocked their SMD program, as evidenced by the reduced expression of the differentiation markers myogenin and the cyclin-dependent kinase inhibitor p21, and by the complete absence of the late-stage differentiation marker myosin heavy chain (MHC) (Fig. 1A). TNF also caused a severe reduction in MyoD protein levels (Fig. 1B) but had no effect on the transcription factors Myf5 or MEF2D, which were also expressed but are inactive in undifferentiated myoblasts. Stable heterologous expression of MyoD restored the ability of C2C12 cells to maintain SMD (Fig. 1C) and form myotubes (Fig. 1D) in the presence of TNF, indicating that TNF inhibits myogenesis by down-regulating MyoD. The rapid loss of MyoD protein was preceded by an equally rapid loss of MyoD mRNA (Fig. 1D). In addition, cells devoid of NF-κB activity (through stable expression of the NF-κB inhibitor IκBα [IκBαSR]) retained MyoD mRNA in the presence of TNF (Fig. 1D), indicating that

Fig. 1. TNF-induced activation of NF-κB inhibits SMD through suppression of MyoD mRNA. (A) C2C12 cells were switched from growth medium [GM: Dulbecco’s modified Eagle’s medium–H (DMEM-H) and 20% fetal bovine serum] to differentiation medium [DM: DMEM-H, 2% horse serum, and insulin (5 μg/ml)] and treated with or without TNF (Promega) at 20 ng/ml for a 72-hour period (to sustain NF-κB activity). TNF was also added at 6, 12, and 24 hours after the initial treatment). At indicated times, whole-cell extracts were prepared and probed for SMD markers myogenin and the cyclin-dependent kinase inhibitor p21 (M-225 and C-19, Santa Cruz Biotechnology), MHC (MY-32, Sigma), or α-tubulin (Sigma). (B) TNF regulation is specific to MyoD. Extracts from C2C12 cells in GM treated with or without TNF were probed for MyoD, Myf5 (M-318 and C-20, respectively, Santa Cruz Biotechnology), and MEF2D (Transduction Laboratories). (C) Overexpression of MyoD restores SMD in the presence of TNF. C2C12 cells were infected with pBabe virus containing MyoD cDNA, and selected with puromycin (1 μg/ml). Clones were screened for their expression of MyoD and p21 (15). Vector control cells and three independent clones—pBabeMyoD1, -4, and -7—were treated with TNF in GM, and extracts were prepared for p21 (after 48 hours) and MHC (after 72 hours). (D) TNF regulation of MyoD is dependent on NF-κB activity. Vector control or cells lacking NF-κB activity [IκBαSR] were differentiated and treated with TNF. At the indicated times, MyoD and GAPDH mRNA expression was analyzed.
NF-κB is a downstream TNF effector that regulates MyoD expression.

To examine which NF-κB subunits were involved in the regulation of MyoD, reporter assays were performed in mouse 10T1/2 fibroblasts. Coexpression of p50 and p65 subunits, or of the p65 subunit alone, strongly blocked MyoD activity, and this effect was reversed with equivalent expression of the IκBαSR plasmid (16). Examination of whole-cell lysates from these transfections demonstrated that p65 alone was sufficient to dramatically reduce MyoD protein levels (Fig. 2A), and this inhibition was again lost after the expression of IκBαSR. In addition, similar to what was observed with TNF treatment of C2C12 cells, overexpression of p65 reduced MyoD mRNA levels (Fig. 2B). These results demonstrate that TNF regulation of MyoD in C2C12 cells is dependent on NF-κB activity and that the p65 subunit is sufficient to induce the down-regulation of MyoD mRNA.

These transfection results showed that p65 regulated MyoD transcripts produced from a heterologous promoter. This suggested that NF-κB was ultimately targeting the MyoD mRNA itself for down-regulation. We therefore attempted to map the MyoD sequences that rendered the mRNA susceptible to p65-mediated down-regulation. Deletion analysis revealed that nucleotides 539 through 914 in the MyoD mRNA were required for this response (Fig. 3A). Introduction of this MyoD fragment into the coding region of the GADPH gene resulted in a p65-dependent reduction in GADPH mRNA levels (Fig. 3B), confirming that p65 targets this MyoD sequence. We next examined whether NF-κB inhibition of MyoD mRNA was direct (that is, requiring NF-κB-dependent transcription). MyoD mRNA decay rates, determined in experiments in which transcription was blocked with actinomycin D, were similar whether C2C12 myocytes were treated or not treated with TNF (16), which suggests that regulation of MyoD is dependent on an NF-κB–responsive gene. To test the requirement for the transcription function of p65 in the regulation of MyoD, we performed reporter assays in mouse 10T1/2 fibroblasts with deletion mutants lacking regions of the p65 transactivation domain. Deletion of the entire transactivation domain of p65 [Fp65(313)] restored MyoD function (Fig. 3C). These data are consistent with those in Fig. 2A, where the p50 subunit of NF-κB, which lacks a transactivation domain, was also unable to inhibit MyoD expression. Taken together, these data indicate that MyoD mRNA is inhibited at the posttranscriptional level in a manner requiring NF-κB–dependent transcription.

TNF is an important mediator in the skeletal muscle degeneration associated with cancer-induced cachexia (10–12). Direct inhibition of NF-κB also blocks cachexia in an animal model (17), suggesting a link between NF-κB and TNF in this disease. Although delivery of TNF is sufficient to induce weight loss in animals, in vitro studies showing that skeletal muscle explants incubated with TNF do not undergo degeneration (18) support the notion that at least one other factor is required to induce muscle wasting. Consistent with these findings, we found that in contrast to the effect of TNF on differentiating myoblasts, differentiated myotubes were completely refractory to this cytokine with respect to MyoD and MHC expression (Fig. 4A). Because other inflammatory cytokines...

**Fig. 2.** MyoD is regulated by the p65 subunit of NF-κB. (A) p65 suppresses MyoD expression. 10T1/2 fibroblasts were transfected with MyoD, NF-κB, and IκBαSR expression plasmids (23). Whole-cell lysates were prepared and probed for MyoD and α-tubulin. (B) p65 suppresses MyoD mRNA expression. Transfections were repeated in 100-mm culture dishes with 0.5 μg of vector or of MyoD expression plasmid, with or without 1.0 μg of p65 expression plasmid. Cells were either kept in GM or differentiated in DM, and at indicated times after transfection, RNA was prepared and probed for MyoD and GAPDH mRNA expression.

**Fig. 3.** Identification of the MyoD sequences regulated by p65-dependent transcription. (A) p65 inhibits MyoD in a region encompassing nucleotides 539 through 914. The MyoD cDNA was removed from plasmid pEMC11s, containing a long terminal repeat element, and ligated into pCDNA3 (Invitrogen), generating the plasmid pCMV-MyoD. 5′ and 3′ deletions were made in MyoD sequences (16). Numerical values present in the names of each plasmid denote the location of the deletions generated with respect to the 5′ and 3′ ends of the MyoD cDNA. Wild-type MyoD (lanes 1 and 2) or deletion plasmids (lanes 3 through 10), each at 0.5 μg, were individually transfected in 10T1/2 fibroblasts with or without a p65 expression plasmid (1.0 μg). RNA was extracted 4 hours after transfection, and MyoD was probed. Numerical values listed with arrows indicate the predicted sizes of exogenously expressed MyoD mRNA products. (B) p65 inhibits GAPDH expression containing MyoD sequences. Nucleotides 313 through 914 from MyoD were ligated into pCMVpGAPDH, generating the plasmid pCMVpGAPDH-MyoD. Transfections and RNA analysis were repeated as described in (A). The arrow indicates endogenous GAPDH expression in 10T1/2 fibroblasts. (C) MyoD regulation requires p65-dependent transcription. The upper portion shows transfections performed in 10T1/2 fibroblasts with a MyoD-responsive reporter (4RTK-Luc, containing four E-box sites of a minimal thymidine kinase promoter), the pCMV-MyoD expression plasmid, and epitope-tagged p65 expression plasmids containing either wild-type [Fp65(551)] or COOH-terminal deletions in the transactivation domain. Cell extracts were prepared with M-Per Extraction Buffer (Pierce) and tested in luciferase assays. The lower portion shows cell extracts from transfections that were probed with an antibody to FLAG (M2, Sigma) to verify the presence of equal p65 expression.
such as interleukin-1β (IL-1β), IL-6, and IFN-γ also induce cachexia in animal models (19), we investigated the possibility that one of these cytokines in combination with TNF could affect skeletal muscle cellular function. Treatments with IL-1β or IL-6 alone or in combination with TNF had no effect on skeletal muscle–specific gene expression (Fig. 4A) (15). In addition, IFN-γ treatment alone had no effect (15). However, differentiated myotubes treated with TNF plus IFN-γ exhibited significant reductions in both MyoD and MHC protein expression (Fig. 4A). Consistent with these results, immunostaining analysis displayed dramatic diminishment of MHC after treatment with TNF plus IFN-γ (Fig. 4C) (16), which correlated with the inability of myofibers to maintain their contractile activities in culture (15). Apoptosis was observed in cultures treated with TNF plus IFN-γ, but only in those myocytes unable to complete their differentiation program (16), indicating that the loss of muscle cell structure was not a result of cell death. Thus, IFN-γ and TNF together regulate myotube degeneration. When C2C12 myotubes devoid of NF-κB activity were treated with TNF plus IFN-γ, they maintained both MyoD and MHC expression (Fig. 4, B and C), indicating that repression of MyoD and impairment of skeletal muscle function are NF-κB–dependent processes.

To test whether TNF and IFN-γ regulate MyoD expression in vivo, we injected cyto- kines directly into mouse skeletal muscle. MyoD mRNA levels were substantially reduced after injection of the cytokines (Fig. 4D). A mixture of tumor-forming Chinese hamster ovary (CHO) cells expressing TNF (Fig. 4D). These results suggest that MyoD is an in vivo target of cytokine signaling.

In adult skeletal muscle, MyoD is expressed at relatively low levels (21, 22). However, in response to injury, MyoD expression is induced from satellite cells, and genetic evidence clearly demonstrates that MyoD is required for these cells to proliferate and to reinitiate SMD necessary for the repair process (4, 21). Our data demonstrate that TNF, along with a second proinflammatory cytokine, IFN-γ, functions through NF-κB to suppress MyoD synthesis by repressing the accumulation of MyoD mRNA. With respect to an injury model such as cachexia, we propose that the cytokines TNF and IFN-γ are likely to affect skeletal muscle regulation at two phases: first, when TNF alone would be sufficient to suppress MyoD expression, thereby inhibiting the formation of new myofibers; and second, when combined signaling between TNF and IFN-γ would be required to inhibit MyoD and cause the degeneration of newly formed myotubes. The combined cytokine effects, working through NF-κB, would lead to the inability to repair damaged skeletal muscle, thereby contributing to the overall wasting process associated with cachexia. Thus, direct inhibition of NF-κB may prove beneficial in reducing the muscle wasting associated with cachexia in cancer patients as well as with other disorders.

**Fig. 4.** IFN-γ is required in addition to TNF to regulate skeletal muscle dysfunction. (A) MyoD and MHC were reduced in myotubes by treatment with TNF plus IFN-γ. C2C12 myocytes were differentiated for 3-day period. DM was replaced with either no additive (lane 1); TNF at concentrations of 20 or 100 ng/ml (lanes 2 and 3, respectively); TNF (20 ng/ml) plus IL-1β (20 ng/ml; Promega) (lane 4); TNF (20 ng/ml) plus IL-6 (20 ng/ml; Promega) (lane 5); or TNF (20 ng/ml) plus IFN-γ (100 U/ml; Life Technologies) (lane 6). Treatments were repeated at 6, 12, and 24 hours. At 48 hours, whole-cell lysates were prepared and probed for MyoD, MHC, and α-tubulin. (B through D) Regulation of MyoD and myosin by TNF and IFN-γ is NF-κB–dependent. Vector control or kBuSR-expressing myoblasts were differentiated as described in (A) and then treated with TNF and/or IFN-γ for 48 hours. Lysates were prepared and probed for MyoD and MHC (B); or cells were immunostained (24) for MHC (C). Scale bar, 15 μm. (D) MyoD was down-regulated by cytokines in vivo. Upper panels: BALB/c and B6/129 mice were given saline alone (vehicle) or a combination of 2 μg of TNF and 5000 U of IFN-γ in a volume of 0.1 ml in the right gastrocnemius muscles were dissected and quick-frozen in liquid nitrogen. At a later time, RNA was prepared with the use of TRIZOL reagent (Life Technologies), and Northern analyses were performed to detect MyoD and GAPDH expression. Lower panels: 1 × 10^7 vector control CHO cells or a mixture of CHO cells expressing human TNF (12) and mouse IFN-γ, clones 1 and 15 (20), were injected into right gastrocnemius muscles of nude mice. At day 12 after injection, gastrocnemius muscles were isolated, RNA was prepared, and Northern analysis was performed, probing for MyoD and GAPDH.

**References and Notes**

16. Supplemental figures and information are available at www.sciencemag.org/content/data/1052677.shl.
20. CHO cells were transfected with an IFN-γ expression plasmid along with a gene conferring neomycin re- sistance. Clones were obtained with neomycin treatment (500 μg/ml) and screened for cytokine expression by sandwich enzyme-linked immunosorbent as- say (Endogen, MA) and Northern analysis. Two clones were analyzed.
23. Mouse 10T1/2 fibroblasts were grown overnight in triplic- ate in 60-mm culture dishes. Cells were then trans- fected by means of Superfect (Qiagen) with 160 ng of either empty vector or a MyoD expression plasmid.
Increase of Maximum Life-Span in Sweden, 1861–1999

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A fundamental question in aging research is whether humans and other species possess an immutable life-span limit. We examined the maximum age at death in Sweden, which rose from about 101 years during the 1860s to about 108 years during the 1990s. The pace of increase was 0.44 years per decade before 1969 but accelerated to 1.11 years per decade after that date. More than 70 percent of the rise in the maximum age at death from 1861 to 1999 is attributable to reductions in death rates above age 70. The rest are due to increased numbers of survivors to old age (both larger birth cohorts and increased survivorship from infancy to age 70). The more rapid rise in the maximum age since 1969 is due to the faster pace of old-age mortality decline during recent decades.

The world record of human life-span seems to be moving upward over time, as suggested by the death in 1997 of Jeanne Calment at the documented age of 122.45 years (1). However, such events are a poor measure of the trend in achieved human life-span, which can be studied more effectively using data for well-defined populations. National demographic statistics suggest that the maximum age at death has been rising steadily in industrialized countries for more than 100 years (2).

Two important questions arise from this observation. First, has this upward trend been steady over time, or has it changed pace in recent years? Perhaps the increase has accelerated due to an intensification of efforts to promote the health of the elderly and to prevent or even cure ailments such as coronary heart disease, stroke, and cancer. Or perhaps the trend has decelerated because maximum ages now observed for humans are approaching a (hypothetical) biological limit. Second, what accounts for the increase in the maximum age at death? There are two competing explanations. One is that it is due merely to the larger size of contemporary populations, which increases the probability that at least one individual will survive to an extreme old age. Another possibility is that the increase reflects improvements in an individual’s probability of survival, especially at older ages.

We have investigated these questions using Swedish national demographic data from 1861 to 1999, which are the longest available series of reliable information on the upper limits of achieved human life-span. Recorded maximum ages at death for men and women in Sweden centered around 101 years during the 1860s and around 108 years during the 1990s (Fig. 1). A statistical analysis indicates that the increase in the maximum age at death accelerated markedly around 1969, rising at a rate of 0.44 years per decade from 1861 to 1969 and 1.11 years per decade from 1969 to 1999. In both time periods, the maximum age at death was on average about 1.7 years lower for men than for women, although time trends for both sexes are similar (3).

Apparent trends in the maximum age at death can be distorted by changes in data quality. Inaccurately reported ages of very old persons and decedents may qualify as the oldest for a birth cohort may or may not be the oldest in the year of death, and vice versa. Our analysis of the cohort trend in the maximum age at death relies on a reconstruction of Swedish mortality by year of birth back to 1751. The life tables and the methodology used to produce them are available on our Web site (5). Although there may be some question about the quality of period life tables from late 18th-century Sweden (especially at older ages), life tables for cohorts born in this era are more reliable because the quality of the statistical system improved over their lifetime. By the time such cohorts attained age 100 or 110, where problems of data quality are most severe, the Swedish statistical system was extremely accurate (7).

In order to determine probability distributions of the maximum age at death, a sequence of age-specific death rates for each cohort is needed. Death rates below age 80 were computed directly from national mortality statistics, reorganized in a cohort format. Death rates above age 90 were estimated by fitting a logistic function to observed death rates and then extrapolating this function to very high ages (9). It was neces-

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24. For immunofluorescence microscopy, cells were fixed directly in the culture wells for 30 min at room temperature with 4% paraformaldehyde diluted in phosphate-buffered saline (PBS). The cells were washed three times with PBS and then permeabilized for 5 min with 0.1% Triton X-100 and sodium citrate and blocked for 30 min in horse serum diluted 1:100 in PBS. After washes with PBS, cells were incubated for 60 min at room temperature with an antibody to anti-skeletal MHC (1:250, Sigma) diluted in 3% bovine serum albumin and PBS. After washes, cells were incubated for 30 min with a rhodamine-conjugated antibody to mouse immunoglobulin G (1:100 Oregon Red, Molecular Probes) diluted in 10% goat serum and PBS. Cells were kept in PBS to be photographed on a Olympus 1X70 fluorescence microscope.

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